

EVALUATION OF THE EFFECT OF INULIN ON METABOLIC PROCESSES

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ABSTRACT

The effect of introducing soluble dietary fiber (5% inulin) into the diet on correcting vitamin D and group B deficiency and its consequences has been studied. The study was conducted on growing male Wistar rats (body weight 51.4 ± 0.5 g) after they had a deficiency caused by a decrease in the content of vitamins D and group B in a vitamin mixture of a semi-synthetic diet for 23 days. The improvement of the rodent slim down with inulin did not influence the retention of vitamins A and D by lacking creatures amid the 7-day adjustment of vitamin status, whereas to some degree abating the reclamation of ordinary vitamins B1 and B6 urinary excretion, and B2 brain substance. At the same time, the substance of vitamin E within the liver was diminished by 1.48 times compared to the control bunch, the concentration of press within the blood plasma was expanded by 32.7, within the liver by 42.6, the substance of manganese within the brain was extended by 1.5 times, which does not surpass the physiological standard. This demonstrates the convenience of improving the slimming down of individuals with a pressing lack of inulin while enhancing with vitamins E and group B.

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Introduction

Adequate dietary fiber intake reduces cholesterol and glucose levels in the blood, normalizes the motility of the gastrointestinal tract, promotes obesity prevention, and reduces the risk of developing cardiovascular diseases, colon cancer, cholelithiasis, diabetes mellitus [1-3].

Many biologically active additives and fortified foods are created mechanically by combining (mixing) several components in one serving, each of which has beneficial properties for the body [4, 5]. At the same time, it is assumed that each ingredient that enters the body from the product will not only be absorbed but will also positively affect a particular function of the body [6, 7]. This approach does not always justify itself. For example, the presence of starch in a specialized product in the form of jelly containing all the vitamins prevented the absorption of vitamin B2 [8, 9]. In a specialized food product, which is positioned as a source of vitamins and soluble dietary fiber, the simultaneous presence of inulin, gum Arabic, and pectin, in a total dose of 300% of the adequate level of consumption of soluble dietary fiber, worsened the indicators of the provision of patients with vitamin E and beta-carotene, despite their presence in the product [10]. The high content of dietary fiber (43 g/day. oat bran) reduced the absorption of vitamins E, B2, and β -carotene but did not affect the bioavailability of vitamin C

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[11]. Enriching the diet with wheat or oat bran, chitosan, and pectin worsened the body's supply of vitamins E, B2, and beta-carotene [12-14].

Inulin is a natural linear polysaccharide (polyfructosan) of sweet taste and belongs to the group of dietary fibers obtained from chicory and Jerusalem artichoke roots [15]. Inulin is a substrate for beneficial bacteria in the colon, which is why it is used as a prebiotic [16, 17]. Inulin is a non-toxic, biodegradable, cheap ingredient with a variety of functions. In the food industry, it is used as a texture modifier, fat substitute, sugar substitute, and prebiotic [18, 19].

Despite the widespread use of inulin in the food industry, there is only some, sometimes contradictory, information about its effect on vitamin metabolism and the antioxidant status of the body. It has been shown that in humans and rats of different ages, inulin intake increases intestinal absorption of calcium (especially in older rats) and magnesium [20]. The inclusion of inulin in the diet of piglets led to an increase in the concentration of iron, copper, and zinc in the blood plasma compared with animals receiving the same vitamin and mineral premix but without inulin additives [21]. One explanation for the increased iron uptake may be the decrease in the concentration of hepcidin, a 25-amino acid peptide hormone produced in the liver and a central regulator of iron homeostasis, found in children after ingestion of inulin enriched with oligofructose [22]. At the same time, in women with anemia, the consumption of 20 g of inulin per day for 4 weeks did not cause an increase in iron absorption. However, changes in the composition of the intestinal microbiota and a decrease in fecal pH were observed [23].

In experiments on mice, it was shown that inulin does not hurt cholesterol metabolism [24]. Inulin consumption has a positive effect on the antioxidant status of laying hens, which was manifested in an increase in the activity of superoxide dismutase, catalase, and glutathione peroxidase in the blood and a decrease in the level of malondialdehyde (MDA) [25]. Reception by healthy persons for 7 weeks of inulin with *L. casei* had a positive effect on markers of oxidative stress (a decrease in blood concentrations of MDA, H₂O₂, oxidized glutathione and a significant increase in the concentration of reduced glutathione and SH groups compared with the control group) [26].

According to other data, the inclusion of inulin in the diet of piglets did not improve the redox balance in the colon and led to a decrease in the activity of DNA repair enzymes [27]. Reception by children suffering from celiac disease, 10 g of inulin for 3 months. It contributed to an increase in the level of vitamins D and E in the blood and did not affect the concentration of vitamin A [28].

The purpose of the study: to study the effect of the introduction of inulin into the diet on the correction of vitamin D and group B deficiency and its consequences in growing rats deficient in these vitamins.

Materials and Methods

Experimental animals, male Wistar rats, were used in the work. The animals were kept in 2 individuals in transparent polycarbonate cages under controlled environmental conditions. environment (temperature 20-26 ° C, relative humidity 30-60%, in lighting mode 12/12 h) on a litter of sawdust. The animals were fed ad libitum and had constant access to distilled water [29].

Before the start of the experiment, during 5 days of quarantine, all animals (n=43) received a full-fledged semi-synthetic diet containing 20% food acid casein, 63% corn starch, 4.5% sunflower oil, 4.5% lard, 3.5% standard salt mixture, 2% microcrystalline cellulose, 1% dry vitamin mixture, 0.30% L-cysteine, 0.25% choline bitartrate and 0.95% sucrose.

At the end of isolate, the rats were arbitrarily separated into two bunches by body weight. The creatures of the control gather bunch 1 all through the test 30 days kept on get a full fledged slim down n 9, and the test bunch n 34 for 23 days gotten nourishment with a 5 fold decreased substance of vitamin D and all B vitamins within the vitamin blend of the diet (**Figure 1**).

The average feed intake in the control and experimental groups during the period of deficit creation did not differ ($p = 0.529$) and amounted to 20.7 ± 0.6 (Me = 20.5) and 21.1 ± 0.8 (Me = 21.3) g/day, respectively.

Then the animals of the experimental group with vitamin deficiency were randomly divided by body weight into 2 subgroups of 10 and 12 individuals each (**Figure 1**). During the next 7 days, these experimental animals were kept on "replenishing" diets aimed at correcting vitamin deficiency, replenishing the deficiency of vitamin D and B vitamins up to 100% of the content in the diet of the control group against the background of the standard diet (+D+B) and against the background of replacing 5% starch with inulin (+D+B+ Inulin).

Rats were placed in metabolic cells, deprived of food, and provided with water without restriction in order to collect urine 20 hours before slaughter. At the end of the experiment, rats previously anesthetized with ether were removed from the experiment by decapitation [30].

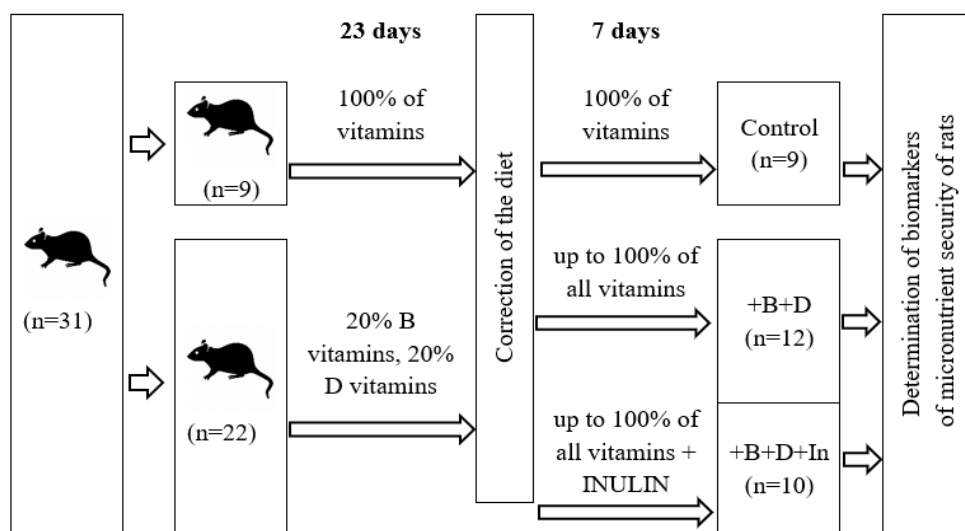


Figure 1. Experimental scheme

To determine by high-performance liquid chromatography, the concentration of vitamins A (retinol and retinol palmitate) and E (α -tocopherol) in blood plasma, freeze-dried liver, and the whole brain of rats [31]. Vitamins B1 and B2 (after acid enzymatic hydrolysis) in the liver, vitamin B2 in urine and plasma, and 4-pyridoxylic acid (4-PC) in urine were measured fluorometrically [32]. The concentration of 25-hydroxyvitamin D (25(OH)D) blood plasma was established by the enzyme immunoassay [33]. Plasma biochemical parameters (calcium, magnesium, iron, phosphorus, glucose, total bilirubin, direct bilirubin, urea, total protein, globulin, creatinine, uric acid, cholesterol (CS), triglycerides (TG), alanine aminotransferase (ALT) activity), aspartate aminotransferase (AST) and alkaline phosphatase were measured on a biochemical analyzer according to standard methods. Experimental data were processed using SPSS Statistics 23.0 (IBM, USA). The nonparametric Mann-Whitney U-test for independent variables and the nonparametric Kruskal–Wallace criterion were used to identify the statistical significance of the differences in continuous quantities. The differences between the analyzed indicators were considered statistically significant at $p < 0.05$.

Results and Discussion

The study of the effect of the introduction of 5% inulin into the diet on the absorption of vitamins and eliminating the consequences of vitamin deficiency was carried out during the correction of deficiency in rats lacking vitamins of groups D and B.

By the end of the stage of vitamin D and group B deficiency, which lasted 23 days, the average body weight of animals of the vitamin-deficient (–D –B) group was 198.5 ± 2.4 g ($I_u = 199.3$ g) and was statistically significantly 5.7% less ($p = 0.046$) than that of the control group (210.4 ± 4.6 g, $I_u = 203.5$ g), which was indirect evidence of the development of alimentary deficiency of these micronutrients in rats. Statistically significant differences in the absolute mass of organs (liver and brain) were observed, but no animals of the control and experimental groups were identified.

Supplementation of the deficient vitamin for 7 days with diets in the absence and presence of inulin completely reversed the growth retardation caused by vitamin deficiency. Also, it affected most of the plasma, liver, and brain parameters (Tables 1–3). The exceptions were iron and manganese. In animals after correction of vitamin status against the background of inulin introduction into the feed, the level of iron in blood plasma was statistically significantly higher by 32.7% compared with that in rats with eliminated vitamin deficiency who did not receive inulin, remaining within the physiological norm (17.4–61.0 mmol/L). Similar results were obtained when inulin was added to the diet of piglets, which led to an increase in the plasma concentration of not only iron but also copper and zinc compared with the indicator in animals receiving the same vitamin-mineral premix, but without the addition of inulin [34].

The addition of inulin to the diet of rats did not affect the level of cholesterol and other indicators of lipid metabolism, as well as the concentration of uric acid (Table 1). This fact does not agree with the data of other researchers who found that the consumption of inulin in approximately the same amounts for a longer period (4 weeks) by older rats of both sexes (9 weeks) led to a decrease in the concentration of cholesterol and uric acid in the blood plasma [35].

As follows from Table 3, even though there was no vitamin E deficiency in the diet of rats, the lack of other vitamins led to a decrease in its content in the liver, especially noticeable against the background of the addition of inulin to the diet, which persisted after 7 days of replenishing the lack of vitamins D and group B. At the same time, the level of α -tocopherol in the brain rats from different groups did not differ statistically significantly.

Inulin administration had no effect on the recovery of vitamin A, B1, and B2 levels in the liver. Still, the recovery of vitamin B2 levels in the brain was delayed after adding the deficient vitamin to the diet of vitamin-deficient rats.

Table 1. Biochemical parameters of rat blood plasma after correction of vitamin D and group B deficiency with and without addition of inulin to the diet ($M \pm m$)

Indicator	Group 1 (control)	Group 2 (+D+B)	Group 3 (+D+B+inulin)
HDL, mmol	1.45 ± 0.11	1.32 ± 0.07	1.45 ± 0.10
Cholesterol, mmol	1.82 ± 0.15	1.70 ± 0.08	1.85 ± 0.11
Triglycerides, mmol	0.86 ± 0.15	0.82 ± 0.09	0.81 ± 0.14
Glucose, mmol/l	8.0 ± 0.4	8.4 ± 0.3	8.7 ± 0.3
AST, Units/l	223 ± 12	197 ± 6	196 ± 5
ALT, Units/l	56.9 ± 2.4	51.8 ± 1.1	48.9 ± 2.1
AST/ALT	4.0 ± 0.3	3.8 ± 0.1	3.9 ± 0.2
LDG, Unit/l	1506 ± 108	1486 ± 88	1599 ± 120
Total protein, g/l	67.6 ± 1.6	67.2 ± 1.1	67.4 ± 1.7
Total bilirubin, µmol/l	4.5 ± 0.6	4.9 ± 0.5	6.3 ± 1.2
Straight bilirubin, µmol/l	3.3 ± 0.4	3.9 ± 0.5	4.5 ± 0.5
Globulins, g/l	32.7 ± 0.9	34.2 ± 0.9	33.9 ± 1.3
Iron, µmol/l	21.6 ± 3.9	33.6 ± 3.7	44.6 ± 4.7
Calcium, mmol/l	3.01 ± 0.09	2.76 ± 0.13	2.85 ± 0.12
Magnesium, mmol/l	1.03 ± 0.02	1.01 ± 0.02	0.99 ± 0.02
Phosphorus, mmol/l	3.19 ± 0.08	2.97 ± 0.09	2.97 ± 0.13
Alkaline phosphatase, Units/l	745 ± 88	588 ± 57	620 ± 50
Osteocalcin, ng/ml	1066 ± 43	1050 ± 34	947 ± 43
Albumin, g/l	33.3 ± 0.6	33.0 ± 0.5	33.5 ± 0.5
Creatinine, µmol/l	47.2 ± 0.4	48.3 ± 0.7	47.0 ± 1.3
Uric acid, µmol/l	47.9 ± 4.9	44.0 ± 3.6	50.3 ± 4.4
Urea, mmol/l	5.3 ± 0.5	6.1 ± 0.3	6.4 ± 0.4

HDL – high-density lipoproteins, AST – aspartate aminotransferase, ALT – alanine aminotransferase, LDH – lactate dehydrogenase.

Table 2. Effect of correction of the combined deficiency in the diet of rats of vitamins D and group B on the background of the addition and without the addition of inulin to the diet on the concentration of vitamins in blood plasma ($M \pm m$)

Indicator	Group 1 (control)	Group 2 (+D+B)	Group 3 (+D+B+inulin)
25(OH)D, ng/ml	9.8 ± 0.5	8.8 ± 0.4	9.5 ± 0.7
Riboflavin, ng/ml	38.2 ± 1.9	41.4 ± 4.4	37.4 ± 1.4
Retinol, µg/dl	35.4 ± 2.0	34.6 ± 2.1	38.4 ± 2.4
α-Tocopherol, mg/dl	1.15 ± 0.14	1.00 ± 0.08	1.18 ± 0.11
α-Tocopherol/TG, µmol/mmol	34.0 ± 4.1	30.0 ± 3.5	32.6 ± 2.4
α-Tocopherol/CS, µmol/mmol	15.3 ± 2.1	13.7 ± 1.0	15.0 ± 1.2
α-Tocopherol/(TG+CS), µmol/mmol	10.1 ± 1.1	9.3 ± 0.8	10.3 ± 0.6

CS – cholesterol, TG – triglycerides

Table 3. Effect of correction of combined deficiency in the diet of rats of vitamins D and group B on the background of inulin addition and without addition to the diet on biomarkers of vitamin and mineral availability in the liver and brain of rats (mcg per 1 g of raw tissue) ($M \pm m$)

Indicator	Group 1 (control)	Group 2 (+D+B)	Group 3 (+D+B+inulin)
	<i>Liver</i>		
Retinol palmitate, µg RE	10.5 ± 0.6	9.0 ± 0.6	9.8 ± 1.0
α-Tocopherol	194 ± 26	137 ± 19	113 ± 11
Vitamin B1	10.0 ± 0.8	9.9 ± 0.6	10.3 ± 0.6
Vitamin B2	27.6 ± 1.1	28.6 ± 0.5	27.5 ± 0.3
Calcium	1200±50	1330±50	1200±30

<i>The whole brain</i>			
Magnesium	187±0.008	200±0.003	186±0.007
Iron	48.4±4.4	55.6±3.7	69.0±6.0
Manganese	1.56±0.10	1.67±0.06	1.73±0.07
Zinc	34.8±1.0	34.7±1.4	33.6±1.3
Copper	3.27±0.17	3.11±0.14	3.13±0.23
<i>The whole brain</i>			
α-Tocopherol	17.9 ± 0.8	18.9 ± 0.6	19.3 ± 0.6
Vitamin B1	4.59 ± 0.24	4.88 ± 0.29	4.39 ± 0.14
Vitamin B2	2.68 ± 0.09	2.44 ± 0.08	2.26 ± 0.08
Calcium	753±50	750±34	767±31
Magnesium	142±7	136±4	134±3
Iron	22.5±1.9	19.6±1.2	20.7±1.1
Manganese	1.00±0.20	1.46±0.17	1.51±0.11
Zinc	11.6±0.3	11.0±0.2	11.1±0.3
Copper	1.31±0.30	1.44±0.19	1.18±0.26

The presence of vitamin D and group 2 in the diet of deficient laboratory animals and subsequent supplementation of the levels of these vitamins to sufficient values without the addition of inulin to the diet (group 2) has no effect on the content of measured elements in edible rat liver (**Table 3**). Against the background of inulin in rats after correction of the combined vitamin deficiency (group 3) was observed statistically significantly higher (by 42.6%) concentration of iron in the liver relative to the indicator in the control and by 24.4% ($p < 0.10$) relative to the indicator in the experimental group of animals that did not receive inulin (group 2). Against the background of inulin consumption, the level of manganese in the liver of rats was slightly higher compared to the control by about 10% ($p < 0.10$) in the absence of statistically significant differences in this indicator in animals of experimental groups 2 (+D+B) and 3 (+D+B+inulin).

In rats, after correcting the combined deficiency in the diet of vitamins D and group B by replenishing their content to an adequate level, the manganese content in the brain of rats of the experimental group 2 (+D+B) exceeded 1.46 times ($p < 0.10$) the corresponding indicator in animals of the control group. After the correction of vitamin deficiency on the background of inulin, the level of manganese in the brain was increased by 1.51 times ($p < 0.05$) in rats compared with the indicator of the control group rats. The absence of statistically significant differences between the indicators of animals of experimental groups 2 and 3 allows us to conclude that the combined lack of vitamins D and group B in the diet, and not inulin, which, apparently, only enhances this process, affects the accumulation of manganese in the brain of rats. Since statistically significant deviations in the content of the remaining studied elements in the brains of animals of experimental groups 2 and 3 from the corresponding indicators in group 1 (control) were not detected (**Table 3**), it can be assumed that the combined deficiency of vitamins D and group B selectively leads to the accumulation of manganese in the brain of rats, which requires further research to clarify mechanisms of this organ-specific influence, especially since excessive exposure to manganese can lead to neurodegenerative diseases similar to Parkinson's disease [36, 37].

Thiamine excretion (based on creatinine released) in rats against the background of inulin added to the diet and increased to an adequate level of missing vitamins (+D+B+Inulin) was 2.1 times lower compared to the values in rats of the control group and animals from the vitamin deficiency correction group without adding inulin (+B+D) to the feed. Similarly, the excretion of the vitamin B6 – 4-PC metabolite was reduced by about 2.2 times (**Table 4**).

Table 4. Biomarkers of micronutrient status in rat urine after correction of vitamin deficiency with and without addition of inulin to the diet ($M \pm m$)

Indicator	Group 1 (control)	Group 2 (+D+B)	Group 3 (+D+B+inulin)
Thiamine / creatinine, $\mu\text{g/g}$	1.1 ± 0.2	1.6 ± 0.3	0.52 ± 0.14
Riboflavin/creatinine, $\mu\text{g/g}$	8.8 ± 0.6	8.7 ± 0.5	10.1 ± 1.0
4-PC / creatinine, $\mu\text{g/g}$	11.2 ± 1.0	9.8 ± 1.0	5.0 ± 1.1
Glucose, mmol	3.6 ± 0.7	3.2 ± 0.6	3.6 ± 0.5
Calcium, mg	0.93 ± 0.21	0.68 ± 0.13	0.86 ± 0.25
Creatinine, mg	4.4 ± 0.3	4.2 ± 0.2	4.6 ± 0.2
Calcium/creatinine, mg/g	0.20 ± 0.04	0.16 ± 0.03	0.18 ± 0.05
Magnesium, μmol	67.4 ± 11.1	45.9 ± 5.4	61.7 ± 6.3
Phosphorus, μmol	0.36 ± 0.04	0.41 ± 0.05	0.50 ± 0.03

Phosphate reabsorption, %	84.5 ± 2.6	82.2 ± 1.7	80.3 ± 2.3
Uric acid, μmol	11.0 ± 0.7	10.0 ± 0.6	10.9 ± 0.6
Urea, mmol	3.2 ± 0.3	2.8 ± 0.2	3.3 ± 0.2

Conclusion

During the study of the effect of the introduction of soluble dietary fiber into the diet on the correction of vitamin D and group B deficiency and its consequences in deficient rats, the effect of inulin on the absorption of vitamins A, E, B1, and B2 and several minerals was evaluated. Judging by the content of vitamins in the organs, the enrichment of the diet. inulin did not affect the absorption of vitamin A by animals deficient in vitamins D and group B during the correction of vitamin status.

The introduction of inulin into the diet was accompanied by an increase in the concentration of iron in blood plasma and liver, but worsened the provision of vitamin E (in terms of liver content, reduced by 1.48 times compared to the indicator of the control group) and slowed down the restoration of normal body supply with vitamins B1, B6 and B2, which manifested itself in a reduced level of vitamin B2 in the brain and reduced excretion of thiamine and 4-PC. An increase in the excretion of water-soluble vitamins occurs when the body is fully saturated with vitamins, and a decrease in the excretion of water-soluble vitamins is an earlier sign of their deficiency. One possible explanation is a more delayed elimination of the deficiency of B vitamins (B1 and B6) against the background of the introduction of inulin into the diet. Against the background of inulin consumption, there was some lag in the restoration of vitamin B2 content in the brain, and the manganese content increased by 1.5 times. A positive aspect is an increase in the concentration of iron in the blood plasma and liver. This reflects an improvement in its absorption when inulin is included in the diet of rats and is consistent with the opinion of other authors about the benefits of inulin enrichment in the diet of people with iron deficiency conditions.

The data obtained indicate that the inclusion of soluble dietary fiber (inulin) in the diet must be accompanied by synchronous enrichment with vitamin E and vitamins of group B. This is also done when using chitosan [38], the addition of which to the diet of rats, without affecting the metabolism of vitamins C, B1, B2, and A, led to a decrease in the concentration of vitamin E in blood plasma. The inclusion of bran in the diet of rats also contributed to a significant decrease in the concentration of vitamin E in blood plasma and liver. Thus, if dietary fibers of various natures manifest themselves differently about B vitamins, then, regardless of their nature, their common property is a deterioration in the body's vitamin E supply during their consumption. Based on the results obtained, it is also important to note that the simultaneous presence of inulin and dietary fiber in the composition of dietary supplements or a food product does not guarantee the complete assimilation of all added vitamins, especially against the background of existing multiple vitamin deficiencies, and can also lead to unforeseen consequences (an increase in the level of manganese in the brain).

In other words, the expected effectiveness of an enriched product or dietary supplement for the correction of vitamin and mineral status may be significantly reduced, which indicates the need for clinical testing, confirming the bioavailability of enriching components and effectiveness for maintaining health.

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References

1. Coelho DB, Lopes LMP, de Oliveira EC, Becker LK, de Paula Costa G, Hermsdorff HHM, et al. Baseline diet quality is related to changes in the body composition and inflammatory markers: An intervention study based on resistance training and nutritional advice. *Biomed Res Int.* 2021;2021:6681823. doi:10.1155/2021/6681823
2. Xie A, Song J, Lu S, Liu Y, Tang L, Wen S. Influence of diet on the effect of the probiotic lactobacillus paracasei in rats suffering from allergic asthma. *Front Microbiol.* 2021;12:737622. doi:10.3389/fmicb.2021.737622
3. Nagdalian AA, Pushkin SV, Lodygin AD, Timchenko LD, Rzhepakovsky IV, Trushov PA. Bioconversion of nutrients and biological active substances in model systems chlorella-insect-livestock. *Entomol Appl Sci Lett.* 2018;5(1):103-10.
4. Chandra S, Saklani S, Kumar P, Kim B, Coutinho HDM. Nutraceuticals: Pharmacologically active potent dietary supplements. *Biomed Res Int.* 2022;2022:2051017. doi:10.1155/2022/2051017
5. Mason SA, Trewin AJ, Parker L, Wadley GD. Antioxidant supplements and endurance exercise: Current evidence and mechanistic insights. *Redox Biol.* 2020;35:101471. doi:10.1016/j.redox.2020.101471
6. Ahmed MM, Montaser SA, Elhadary A, Elaragi GGM. Another concept of cancer interpretation in view of the interaction between plasma radiation and DNA. *Clin Cancer Investig J.* 2022;11(4):33-43.

7. Domiaty DMM. Gum Arabic mitigates AlCl₃-induced nephrotoxicity by upregulating the XRCC1 gene and downregulating Ki67 and P53 expressions. *Clin Cancer Investig J.* 2022;11(4):44-51.
8. Özüdoğru S, Tosun G. Evaluation of microleakage and fatigue behaviour of several fiber application techniques in composite restorations. *Ann Dent Spec.* 2022;10(2):60-6.
9. Shawky M, Aljahdali E, Alkhanbashi R. Medication-related osteonecrosis of the jaw: Evaluation of knowledge and attitude among Saudi dental students and interns. *Ann Dent Spec.* 2022;10(2):52-9.
10. Sadovoy VV, Selimov MA, Shchedrina TV, Nagdalian AA. Usage of biologically active supplements in the technology of prophylactic meat products. *Res J Pharm Biol Chem Sci.* 2016;7(5):1861-5.
11. Kristensen M, Bügel S. A diet rich in oat bran improves blood lipids and hemostatic factors and reduces apparent energy digestibility in young healthy volunteers. *Eur J Clin Nutr.* 2011;65:1053-8. doi:10.1038/ejcn.2011.102
12. Jefferson A, Adolphus K. The effects of intact cereal grain fibers, including wheat bran on the gut microbiota composition of healthy adults: A systematic review. *Front Nutr.* 2019;6:33. doi:10.3389/fnut.2019.00033
13. Nefitullayeva A, Azimova S, Maskurova Y, Tsimigigova R, Papanova A, Dachava S, et al. Investigation of the yield of biologically active substances during the ultrasound and electro-discharge extraction of medicinal herbs of the foothills of the North Caucasus. *Potr S J Food Sci.* 2023;17:217-30. doi:10.5219/1843
14. Martazanova L, Maslova A, Ulikhanov K, Khadaeva D, Shemshedinova A, Abdullayeva AM, et al. The study of the effect of drinks based on extracts of herbal adaptogens on the functional status of athletes during physical activity. *Potr S J Food Sci.* 2023;17:30-42. doi:10.5219/1804
15. Qin YQ, Wang LY, Yang XY, Xu YJ, Fan G, Fan YG, et al. Inulin: Properties and health benefits. *Food Funct.* 2023;14(7):2948-68. doi:10.1039/d2fo01096h
16. Alaghemandan H, Ferdosi M, Savabi O, Yarmohammadian MH. Proposing a framework for accreditation of dental clinics in Iran. *J Organ Behav Res.* 2022;7(2):161-70.
17. Dumitru M, Vranceanu D, Banica B, Cergan R, Taciuc IA, Manole F, et al. Management of aesthetic and functional deficits in frontal bone trauma. *Medicines.* 2022;58(12):1756.
18. Delcea C, Siserman C. Validation and standardization of the questionnaire for evaluation of paraphilic disorders. *Rom J Leg Med.* 2020;28(1):14-20.
19. Ahmed W, Rashid S. Functional and therapeutic potential of inulin: A comprehensive review. *Crit Rev Food Sci Nutr.* 2019;59(1):1-13. doi:10.1080/10408398.2017.1355775
20. Tawfik MM, Xie H, Zhao C, Shao P, Farag MA. Inulin fructans in diet: Role in gut homeostasis, immunity, health outcomes, and potential therapeutics. *Int J Biol Macromol.* 2022;208:948-61. doi:10.1016/j.ijbiomac.2022.03.218
21. Li B, Schroyen M, Leblais J, Wavreille J, Soyeurt H, Bindelle J, et al. Effects of inulin supplementation to piglets in the suckling period on growth performance, postileal microbial and immunological traits in the suckling period and three weeks after weaning. *Arch Anim Nutr.* 2018;72(6):425-42. doi:10.1080/1745039X.2018.1508975
22. Hughes RL, Alvarado DA, Swanson KS, Holscher HD. The prebiotic potential of inulin-type fructans: A systematic review. *Adv Nutr.* 2022;13(2):492-529. doi:10.1093/advances/nmab119
23. Le Bastard Q, Chapelet G, Javaudin F, Lepelletier D, Batard E, Montassier E. The effects of inulin on gut microbial composition: a systematic review of evidence from human studies. *Eur J Clin Microbiol Infect Dis.* 2020;39(3):403-13. doi:10.1007/s10096-019-03721-w
24. Illippangama AU, Jayasena DD, Jo C, Mudannayake DC. Inulin as a functional ingredient and their applications in meat products. *Carbohydr Polym.* 2022;275:118706. doi:10.1016/j.carbpol.2021.118706
25. Shang H, Zhang H, Guo Y, Wu H, Zhang N. Effects of inulin supplementation in laying hens diet on the antioxidant capacity of refrigerated stored eggs. *Int J Biol Macromol.* 2020;153:1047-57. doi:10.1016/j.ijbiomac.2019.10.234
26. Kleniewska P, Hoffmann A, Pniewska E, Pawliczak R. The influence of probiotic lactobacillus casei in combination with prebiotic inulin on the antioxidant capacity of human plasma. *Oxid Med Cell Longev.* 2016;2016:1340903. doi:10.1155/2016/1340903
27. Barszcz M, Taciak M, Tuśnio A, Świąch E, Bachanek I, Kowalczyk P, et al. The effect of dietary level of two inulin types differing in chain length on biogenic amine concentration, oxidant-antioxidant balance, and DNA repair in the colon of piglets. *PloS one.* 2018;13(9):e0202799. doi:10.1371/journal.pone.0202799
28. Drabińska N, Krupa-Kozak U, Abramowicz P, Jarocka-Cyrta E. Beneficial effect of oligofructose-enriched inulin on vitamin D and E status in children with celiac disease on a long-term gluten-free diet: a preliminary randomized, placebo-controlled nutritional intervention study. *Nutrients.* 2018;10(11):1768. doi:10.3390/nu10111768
29. Lyashenko EN, Uzbekova LD, Polovinkina VV, Dorofeeva AK, Ibragimov S-US-u, Tatamov AA, et al. Study of the embryonic toxicity of TiO₂ and ZrO₂ nanoparticles. *Micromachines.* 2023;14(2):363. doi:10.3390/mi14020363
30. Verevkina M, Goncharov V, Nesmeyanov E, Kamalova O, Baklanov I, Pokhilko A, et al. Application of the Se NPs-Chitosan molecular complex for the correction of selenium deficiency in rats model. *Potr S J Food Sci.* 2023;17:455-66. doi:10.5219/1871
31. Kozyreva F, Tuaeava I, Baklanova O, Bondarenko N, Bernyukevich T, Semkina E, et al. Improving milk quality to prevent microelement deficiencies: A socio-hygienic perspective on adding bioavailable trace elements. *Potr S J Food Sci.* 2023;17:433-43. doi:10.5219/1872

32. Obeid R, Geisel J, Nix WA. 4-Pyridoxic Acid/Pyridoxine ratio in patients with type 2 diabetes is related to global cardiovascular risk scores. *Diagnostics (Basel)*. 2019;9(1):28. doi:10.3390/diagnostics9010028
33. Han S, Qiu W, Zhang J, Bai Z, Tong X. Development of a chemiluminescence immunoassay for quantification of 25-hydroxyvitamin D in human serum. *J Anal Methods Chem*. 2020;2020:9039270. doi:10.1155/2020/9039270
34. Samolińska W, Grela ER. Comparative effects of inulin with different polymerization degrees on growth performance, blood trace minerals, and erythrocyte indices in growing-finishing pigs. *Biol Trace Elem Res*. 2017;176(1):130-42. doi:10.1007/s12011-016-0796-y
35. Massot-Cladera M, Azagra-Boronat I, Franch À, Castell M, Rodríguez-Lagunas MJ, Pérez-Cano FJ. Gut health-promoting benefits of a dietary supplement of vitamins with inulin and acacia fibers in rats. *Nutrients*. 2020;12(8):2196. doi:10.3390/nu12082196
36. Khandia R, Pandey MK, Zaki MEA, Al-Hussain SA, Baklanov I, Gurjar P. Application of codon usage and context analysis in genes up-or down-regulated in neurodegeneration and cancer to combat comorbidities. *Front Mol Neurosci*. 2023;16:1200523. doi:10.3389/fnmol.2023.1200523
37. Khandia R, Ali Khan A, Alexiou A, Povetkin SN, Verevkina MN. Codon usage analysis of pro-apoptotic bim gene isoforms. *J Alzheimers Dis*. 2022;86(4):1711-25. doi:10.3233/JAD-215691
38. Gim SY, Jung J, Kwon Y, Kim MJ, Kim G, Lee J. Effects of chitosan and collagen containing α -tocopherol on the oxidative stability in bulk oil and oil-in-water emulsion. *Food Sci Biotechnol*. 2018;27(4):947-56. doi:10.1007/s10068-018-0345-x