## **Pharmacophore**

ISSN-2229-5402



Journal home page: http://www.pharmacophorejournal.com

# ASSESSMENT OF CYTOTOXIC ACTIVITY TOWARDS PC3 CELL LINE OF PEPTIDE ESTERS OF GALANTAMINE: GAL-LEU AND GAL-VAL

Dobrina Tsvetkova<sup>1\*</sup>, Lyubomir Vezenkov<sup>2</sup>, Tchavdar Ivanov<sup>2</sup>, Dancho Danalev<sup>3</sup>, Ivanka Kostadinova<sup>4</sup>

- 1. Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Medical University Sofia, Sofia, Bulgaria.
- 2. Department of Organic Chemistry, University of Chemical Technology and Metallurgy, 1756 Sofia, Bulgaria.
- 3. Department of Biotechnology, University of Chemical Technology and Metallurgy, 1797 Sofia, Bulgaria.
- 4. Department of Pharmacology, Pharmacotherapy and Toxicology, Faculty of Pharmacy, Medical University of Sofia, Sofia 1000, Bulgaria.

## ARTICLE INFO

Received: 03 Jan 2023 Received in revised form: 17 Apr 2023 Accepted: 20 Apr 2023 Available online: 28 Apr 2023

*Keywords:* Peptide esters, Galantamine, MTT, PC3 cell line

## ABSTRACT

h: The current work's objective was to examine the cytotoxic activity of recently created peptide esters of Galantamine: GAL-LEU and GAL-VAL on prostate cancer PC3 cell lines. For the estimation of the cytotoxic effect of Galantamine derivatives, the MTT reduction assay was applied. PC3 cells were triplicate exposed separately to each of the peptide esters, applied in different concentrations (1.875  $\mu$ M ÷ 30  $\mu$ M). In the MTT test, the reduction of tetrazolium salt MTT resulted in the creation of formazan, whose absorbance was measured spectrophotometrically at wavelength 570 nm. The experimental results show that peptide ester GAL-LEU at 30  $\mu$ M inhibits 55.36% of PC3 cell growth with an index of cell viability of 44.64 %. The lower antiproliferative effect of derivative GAL-VAL was proven by the fact that 30  $\mu$ M inhibits 43.96 % of cell growth. The results showed that both of the tested esters had cytotoxic action against the PC3 cell line, but that GAL-LEU has a stronger antiproliferative impact than GAL-VAL (IC<sub>50</sub> >30  $\mu$ M), because of its lower value of IC<sub>50</sub> = 30.8  $\mu$ M.

> This is an **open-access** article distributed under the terms of the <u>Creative Commons Attribution-Non</u> <u>Commercial-Share Alike 4.0 License</u>, which allows others to remix, and build upon the work non commercially.

**To Cite This Article:** Tsvetkova D, Vezenkov L, Ivanov T, Danalev D, Kostadinova I. Assessment of Cytotoxic Activity Towards PC3 Cell Line of Peptide Esters of Galantamine: GAL-LEU and GAL-VAL. Pharmacophore. 2023;14(2):111-9. https://doi.org/10.51847/1uNNypZfX2

## Introduction

Cancer is a disease involving unregulated cell growth. An important risk factor for cancer is old age [1]. Other factors are smoking [2], environmental agents such as exposure to chemicals, alcohol, drugs, sunlight, ionizing radiation, electromagnetic fields [3], and infectious agents [4]. Cytotoxic chemotherapy is applied for different types of cancer including lung, pancreatic and colorectal cancer [5].

The most used anticancer drugs are antimetabolites [6], cytotoxic antibiotics anthracyclines [7], and inhibitors of topoisomerase [8]. Flutamide, Bicalutamide, and Nilutamide are applied for prostate cancer [9]. It was discovered that extracts from different plants like species of *Astragalus* possess anticancer activity [10].

Alzheimer's disease is treated using Galantamine [11] improves cognitive functions by the following mechanisms [12]: inhibition of acetylcholinesterase [13]; stimulation of  $\alpha$ 7-subtype binding position of nicotinic acetylcholine receptors [14] and antioxidant activity [15, 16].

In connection with that L-Leucyl-L-Leucine methyl ester induces apoptosis on cell lines [17] and because the discovery of cytotoxic properties of new compounds can increase the possibility of obtaining the new *in-vivo* effective pharmaceutical

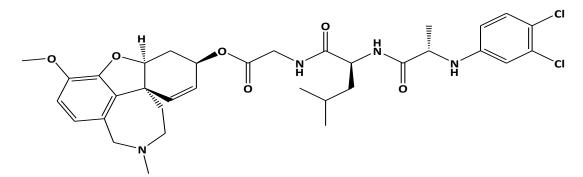
**Corresponding Author:** Dobrina Tsvetkova; Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Medical University – Sofia, Sofia, Bulgaria. E-mail: dtsvetkova@pharmfac.mu-spfia.bg.

Pharmacophore, 14(2) 2023, Pages 111-119

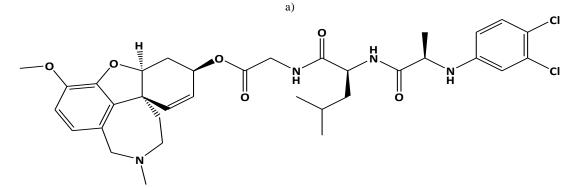
agents for anticancer therapy, the current work's objective was to examine the cytotoxic activity against on PC3 prostate cancer cells [18, 19] of newly synthesized peptide esters: 6-O-N-[N-(3.4-dichlorophenyl)-D, L-Alanyl]-L-Leucyl-Glycil-Galantamine (GAL-LEU) and 6-O-N-[N-(3.4-dichlorophenyl)-D, L-Alanyl]-L-Valil-Glycil-Galantamine (GAL-VAL) [20], for which has been observed to exert both inhibitory activities against acetylcholinesterase and  $\gamma$ -secretase [21] and which exhibit antioxidant qualities in the FRAP (ferric reducing/antioxidant power) process [22, 23].

## **Materials and Methods**

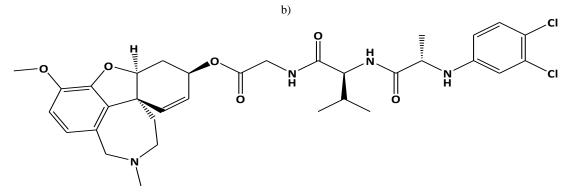
- 1. Tested Peptide Esters, Synthesized from Vezenkov et al. [20]
- 2. Investigated Peptide Esters GAL-LEU and GAL-VAL, Synthesized from Vezenkov et al. [20] (Figure 1)



(4aS,6R,8aS)-3-methoxy-11-methyl-4a,5,9,10,11,12-hexahydro-6*H*-benzo[2,3]benzofuro[4,3cd]azepin-6-yl (3,4-dichlorophenyl)-*L*-alanyl-*L*-leucylglycinate

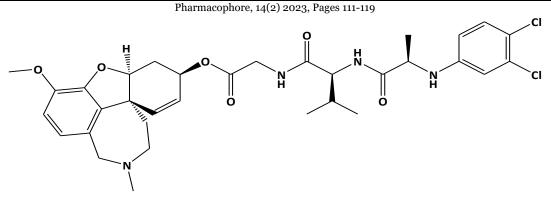


(4a*S*,6*R*,8a*S*)-3-methoxy-11-methyl-4a,5,9,10,11,12-hexahydro-6*H*-benzo[2,3]benzofuro[4,3*cd*]azepin-6-yl (3,4-dichlorophenyl)-*D*-alanyl-*L*-leucylglycinate



(4a*S*,6*R*,8a*S*)-3-methoxy-11-methyl-4a,5,9,10,11,12-hexahydro-6*H*-benzo[2,3]benzofuro[4,3*cd*]azepin-6-yl (3,4-dichlorophenyl)-*L*-alanyl-*L*-valylglycinate

c)



(4a*S*,6*R*,8a*S*)-3-methoxy-11-methyl-4a,5,9,10,11,12-hexahydro-6*H*-benzo[2,3]benzofuro[4,3*cd*]azepin-6-yl (3,4-dichlorophenyl)-*D*-alanyl-*L*-valylglycinate

d)

Figure 1. Chemical structures of GAL-LEU and GAL-VAL.

## 3. Reagents with Analytical Grade Quality

Standard MTT (3-[4.5-dimethylthiazole-2-yl]-2.5-diphenyl-tetrazolium bromide), dimethylsulfoxide, fetal bovine serum (FBS), 100 IU/ml Penicillin, 100 µg/ml Streptomycin, 0.25 % Trypsin EDTA 1X.

## 4. Preparation of Solutions of Peptide Esters

An accurately weighed quantities of the examined peptide esters GAL-LEU and GAL-VAL were dissolved separately in dimethylsulfoxide to obtain concentrations:  $1.875 \,\mu$ M;  $3.75 \,\mu$ M;  $7.5 \,\mu$ M;  $15 \,\mu$ M;  $30 \,\mu$ M.

## 5. Preparation of MTT Solution

An accurately weighed quantity of MTT was dissolved in phosphate buffer solution to obtain a solution with a concentration of 5 mg/ml. This solution is stable for 1 month at storage at 4 °C.

#### 6. In vitro Cancer Test Systems - Cell Lines

Dulbecco's Modified Eagle Medium was used for the cultivation of the PC3 prostate cancer cell line for the assessment of the cytotoxic activity of peptide esters.

#### 7. MTT-Test

To examine the PC3 cell line's susceptibility to the cytotoxicity of Galantamine peptide esters, the MTT test of Mosmann was applied [18]. Into each well separately were added 200  $\mu$ l of solution of respective ester in fresh medium in different dilutions (1.875  $\mu$ M ÷ 30  $\mu$ M). For each of the examined esters in each concentration experiments were performed in triplicates. The average values were calculated. After 48 hours of exposure on esters 200  $\mu$ l 0.5 mg/ml MTT were added directly to each well. The plates were then incubated for a further 4 hours at 37°C with 5% CO2. To solubilize the produced formazan, 100  $\mu$ l of dimethylsulfoxide was added to each well after the supernatant was removed. The formazan's absorbance was measured at  $\lambda = 570$  nm.

## 8. Incubation of PC3 Cells

96 Microplates with flat bottoms were used. By using the established MTT colorimetric test, the compounds' cytotoxic activity was assessed. PC3 cells were added to Dulbecco's Modified Eagle Medium in 75 cm<sup>2</sup> flasks. After addition to a medium of 5 % fetal bovine serum, 100  $\mu$ g/ml Streptomycin, and 100 IU/ml Penicillin, PC3 cells were incubated in a fully humidified atmosphere of 5% CO<sub>2</sub> at 37 °C for 24 h. In the obtained phase of exponential growth, the cells were trypsinized, ad centrifuged. Haemocytometer was used for the determination of the content of cells. Cell culture with a concentration of 1.10<sup>5</sup> cells/ml was obtained by diluting with a particular volume of medium. 100  $\mu$ l/well was added to 96-well plates. Samples were incubated for 24 hours at 37 °C in a thoroughly humidified 5% CO2 environment. The culture media was withdrawn after incubation.

#### **Results and Discussion**

The cytotoxic activity of peptide esters GAL-LEU and GAL-VAL against PC3 prostate cells was estimated by using the standard MTT colorimetric test.

As  $(A_{(+)})$  control PC3 line was treated with MTT without peptide esters. As  $(A_{(-)})$  control PC3 cell line was dissolved in a culture medium without MTTand peptide esters. For the investigation of the antiproliferative effect of peptide esters, the PC3 cell line was treated separately and duplicated with GAL-LEU in different concentrations (1.875  $\mu$ M  $\div$  30  $\mu$ M) and triplicated with GAL-VAL in 30  $\mu$ M. MTT assay was applied. As a standard was used Doxorubicin. In Table 1. absorbances of positive

Pharmacophore, 14(2) 2023, Pages 111-119 A (+) and negative A (-) controls and of formazan obtained after treatment of PC3 cell line with tested compounds are summarized.

Cell line	PC3						
N:	GAL-LEU			GAL-VAL			
1.	0.947	0.065		1.155	0.064		
2.	0.909	0.063		1.007	0.063		
3.	0.883	0.061		1.019	0.059		
4.	0.930	0.063		1.021	0.065		
5.	0.930			1.053			
6.				1.077			
$\overline{\mathbf{X}}$	0.920	0.063		1.055	0.063		
SD	0.025	0.002		0.055	0.003		
C <sub>GAL-LEU</sub> [µM]	Absorbances of formazan [AU]						
	1	2	3	$\overline{\mathbf{X}}$	SD		
3.75	0.816	0.830		0.823	0.010		
7.5	0.864	0.802		0.833	0.044		
15	0.751	0.837		0.794	0.061		
30	0.465	0.426		0.446	0.028		
Cgal-val [µM]	Absorbances of formazan [AU]						
30	0.666	0.615	0.576	0.619	0.045		

Table 1. Absorbances of controls and of formazan produced from the GAL-LEU- and GAL-VAL-treated PC3 cell line.

In **Figure 2** is illustrated that the increase of concentration correlates with the decrease of formazan's absorbance and of cell viability.

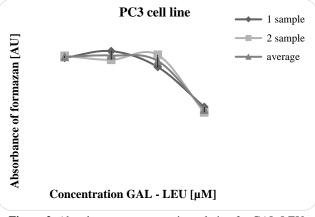


Figure 2. Absorbance – concentration relation for GAL-LEU.

Experiments were applied in duplicate. Sample  $1 = 1^{st}$  experiment Sample  $2 = 2^{nd}$  experiment

In Table 2 the data for the activity of esters on inhibition of cell growth (%) are summarized.

Cgal-leu [µM]	Inhibition PC3 cell growth [%]					
	1	2	3	$\overline{\mathbf{X}}$	SD	
3,75	12.11	10.48		11.30	1.15	
7,5	6.51	13.75		10.13	5.12	
15	19.70	9.66		14.68	7.10	
30	53.08	57.63		55.36	3.22	
C <sub>GAL-VAL</sub> [µM]	Inhibition PC3 cell growth [%]					
	1	2	3	$\overline{\mathbf{X}}$	SD	
30	39.22	44.36	48.29	43.96	4.55	

## **Tsvetkova** *et al.*, **2023** Pharmacophore, 14(2) 2023, Pages 111-119

In **Figure 3** the cytotoxic effect of GAL-LEU against the PC3 cell line is illustrated.

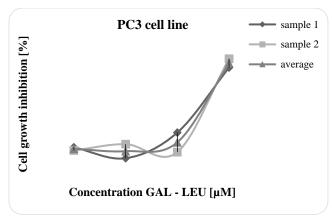


Figure 3. Cytotoxic effect of GAL-LEU against PC3 cell line.

Experiments were applied in duplicate. Sample  $1 = 1^{st}$  experiment Sample  $2 = 2^{nd}$  experiment

The obtained data for the inhibition concentration IC<sub>50</sub> are GAL-LEU: 30.8  $\mu$ M; GAL-VAL: > 30  $\mu$ M. In **Table 3** the data for activity of esters on the index of cell viability V (%) and are presented.

Cgal-leu [µM] —	Index of cell viability of PC3 cell line [%]						
	1	2	3	X	SD		
3.75	87.89	89.52		88.70	1.15		
7.5	93.49	86.25		89.87	5.12		
15	80.30	90.34		85.32	7.10		
30	46.92	42.37		44.64	3.22		
IC <sub>50</sub>	29.84	31.76		30.80	1.38		
CGAL-VAL [µM] —	Index of cell viability of PC3 cell line [%]						
	1	2	3	$\overline{\mathbf{X}}$	SD		
30	60.78	55.64	51.71	56.04	4.55		

In Figure 4 the accordance between the concentration of GAL-LEU and the index of cell viability V (%) is demonstrated.

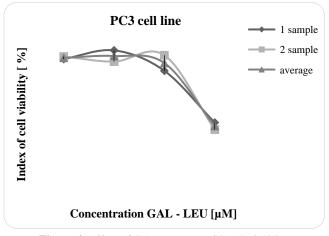


Figure 4. Effect of GAL-LEU on PC3 cell viability.

Experiments were applied in duplicate. Sample  $1 = 1^{st}$  experiment Sample  $2 = 2^{nd}$  experiment

#### Pharmacophore, 14(2) 2023, Pages 111-119

Prostate adenocarcinoma is considered to be the second most commonly diagnosed malignancy [24] and the second highest factor of cancer-related death in men [25]. Prostate cancer cell proliferation is regulated by androgens via the androgen receptor [26]. Aggressive prostate cancer is associated with up-regulation of caveolin-1 [27]. The most often applied therapy for patients with metastatic prostate cancer is the combination of Docetaxel and Prednisone [28]. Other treatments include surgery, radiation, and hormonal therapy, naturally occurring compounds: lycopene, curcumin [25], genistein [29], and natural polyphenol Gallic acid [30]. Liposomal hydroxy aluminum phthalocyanine gel has the potential for effective photodynamic therapy of prostate cancer since vitamin D can be utilized to treat and prevent prostate cancer since vitamin D deficiency has been associated with increased risk of illness [32, 33].

The PC3, LNCaP, and DU145 cells [34] are being widely used to analyze and characterize the development of human prostate cancer *in vitro* and *in vivo* [35]. An epithelial prostatic adenocarcinoma cell line called PC3 is utilized to study the biochemical alterations in advanced prostate cancer cells and gauge how well they respond to chemotherapy drugs. The PC3 cell line was created in 1979 [36].

For the investigation of cell viability and proliferation [37] widely is applied the MTT test of Moosmann, in which soluble yellow MTT is reduced by the mitochondrial enzyme activity of viable cells into soluble in dimethylsulfoxide violet formazan, which absorbance can be measured spectrophotometrically at  $\lambda = 570$  nm [18]. The index of cell viability V (%) and cell growth inhibition I (%) were calculated. The amount of activity is a gauge of the viability of the cells since the decrease of MTT can only occur in metabolically active cells. A decrease in the rate of cell growth is shown by absorbance values that are lower than those of the control cells. An increase in cell proliferation is indicated by a greater absorbance rate.

It has been demonstrated that indole-3-carbinol inhibits the growth of PC3 and DU145 cells leading to apoptosis [38]. Pinocembrin (5, 7-dihydroxy flavanone) shows a potent antiproliferative effect against PC3, DU-145 and LNCaP cells [39]. Propolis extracts suppresses the viability of human prostate cancer cells [40].

According to studies, arachidonic acid and its metabolites, which are created by the enzyme 5-lipoxygenase, promote the proliferation of prostate cancer cells. The natural products that act as 5-lipoxygenase inhibitors include caffeic acid [41], curcumin [42], quercetin [43], luteolin [44], resveratrol [45], rosmarinic acid [46], nordihydroguaiaretic acid [47].

It has been observed that Caffeic acid phenethyl ester dosage dependently inhibits of LNCaP, DU-145, and PC3 cells [48]. LNCaP and DU 145 cell-growth is suppressed by Curcumin from *Curcuma longa L*. and genistein, daidzein, and glycitein. Isoflavones alone do not have as strong an inhibitory effect on cell growth as does the combination with Curcumin [49].

Curcumin suppresses proteins Bcl-2 and Bcl-x what inhibit apoptosis. Curcumin activates pro-apoptotic proteins from the Bcl-2 family and caspases [50]. According to research, luteolin inhibits the growth of human prostate tumors [43]. Quercetin is known to cause death of prostate cancer cells, which is a result of stimulation of caspases [44]. Nordihydroguaiaretic acid increases  $Ca^{2+}$  concentration by releasing of  $Ca^{2+}$  from the endoplasmic reticulum [47].

Tumors can be a result of stem cells mutations [51]. MTT test of Moosmann is used for the estimation of effect of plant extract against cancers. Some tumors such as lung, breast, prostate, and cervical are high spread in population, while others like urinary bladder primary squamous cell carcinoma are with rare cases [52]. In Saudi Arabia are spread cervical cancer, thalassemia and sickle-cell anaemia [53].

It has been reported that for the estimation of the citotoxic activity of extracts of *Rheum ribes L*. towards cell lines A-549 and KB [16], *Acalypha wilkesiana* against cervical cancer HeLa cell line [19] and MCF-7 breast cancer cell line [23] and ginger on cell viability in breast and pancreatic cancer [54], was used MTT test. For these assay a specific cell-line-grow mediun was applied [55].

In our previous experiments, the survival of 3T3 cells was assessed using Moosmann's MTT assay. In concentration 30  $\mu$ M GAL-VAL inhibits 88.32 % of 3T3 cell growth and exerts cytotoxic activity with IC<sub>50</sub> = 23.17  $\mu$ M [56]. GAL-LEU in concentration 30  $\mu$ M suppresses 99.9 % of 3T3 cells and possesses antiproliferative cytotoxic activity with IC<sub>50</sub> = 19  $\mu$ M [57]. The current investigation of the Galantamine peptide esters GAL-LEU and GAL-VAL [58] effects on the inhibition of cell growth PC3 cell line that was treated triplicated separately with each of the examined peptide esters in different concentrations (1.875  $\mu$ M  $\div$  30  $\mu$ M). In the applied MTT test of Mosmann, the accordance of formazan is proportional to viability cell lines. The cytotoxic effect was estimated as concentration that suppresses 50 % of PC3 cells (IC<sub>50</sub>). Cell growth inhibition I (%) and index of cell viability V (%) by the following equations were calculated:

$$I(\%) = 100 - \frac{A(t) - A(-)}{A(+) - A(-)}$$
(1)

$$V(\%) = \frac{A(t) - A(-)}{A(+) - A(-)}$$
(2)

V (%) – index of cell viability

I (%) – cell growth inhibition

At – mean absorbance value of formazan, obtained after treatment of the examined cells with test compounds

 $A_{(+)}$  — mean absorbance of formazan, obtained with the positive control (treated with an MTT solution without the test chemicals in the studied cell line

A(-) – mean absorbance value of formazan, obtained with negative control (MTT, without the examined chemicals.

#### Pharmacophore, 14(2) 2023, Pages 111-119

The experimental results show that GAL-LEU at concentration 30  $\mu$ M inhibits 55.36 % of PC3 cell growth with an index of cell viability of 44.64 %. GAL-VAL in concentration 30  $\mu$ M inhibits 43.96 % of cell growth with an index of cell survival of 56.04 %. These results prove the lower antiproliferative effect of GAL-VAL in comparison with GAL-LEU.

## Conclusion

The treatment of the PC3 cell line with peptide esters decreased formazan concentration and absorbance, indicating their ability to suppress growth. The outcomes of the experiment demonstrated that GAL-LEU slowed the growth of PC3 cancer cells. The experimental findings demonstrated that both esters had cytotoxic effects on the PC3 cell line, but that GAL-LEU has a stronger cytotoxic impact than GAL-VAL (IC<sub>50</sub> > 30  $\mu$ M) because of its lower IC<sub>50</sub> of 30.8  $\mu$ M. The antiproliferative activity of peptide esters is lower compared to standard Doxorubicin activity (IC<sub>50</sub> = 1.698  $\mu$ M ± 0.285  $\mu$ M).

Acknowledgments: The authors are gratefully acknowledged Rizwana Malik, Sadia Siddiq, and Prof. Iqbal Choudhary from "Dr. Panjwani Center for Molecular Medicine and Drug Research, ICCBS, University of Karachi, Pakistan, for skillful technical assistance, great experimental support, extending laboratory competence and for their continued scientific correspondence, cooperation, and help. The authors gratefully acknowledged Prof. Danka Obreshkova for scientific consultation and cooperation.

## Conflict of interest: None

## Financial support: None

Ethics statement: None

## References

- 1. Anisimov VN, Sikora E, Pawelec G. Relationships between cancer and aging: A multilevel approach. Biogerontol. 2009;10(4):323-38. doi:10.1007/s10522-008-9209-8
- 2. Kuper H, Boffetta P, Adami HO. Tobacco use and cancer causation: Association by tumor type. J Int Med. 2002;252(3):206-24. doi:10.1046/j.1365-2796.2002.01022.x
- 3. Irigaray P, Newby JA, Clapp R, Hardell L, Howard V, Montagnier L, et al. Lifestyle-related factors and environmental agents causing cancer: An overview. Biomed Pharmacother. 2007;61(10):640-58. doi:10.1016/j.biopha.2007.10.006
- 4. Samaras V, Rafailidis PI, Mourtzoukou EG, Peppas G, Falagas ME. Chronic bacterial and parasitic infections and cancer: A review. J Infect Dev Ctries. 2010;4(5):267-81. doi:10.3855/jidc.819
- 5. Corrie PG. Cytotoxic chemotherapy: Clinical aspects. Medicine. 2008;36(1):24-8. doi:10.1016/j.mpmed.2007.10.012
- 6. Parker WB. Enzymology of purine and pyrimidine antimetabolites used in the treatment of cancer. Chem Rev. 2009;109(7):2880-93. doi:10.1021/cr900028p
- Minotti G, Menna P, Salvatorelli E, Cairo G, Gianni L. Anthracyclines: Molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. Pharmacol Rev. 2004;56(2):185-229. doi:10.1124/pr.56.2.6
- Nitiss JL. Targeting DNA topoisomerase II in cancer chemotherapy. Nature Rev Cancer. 2009;9(5):338-50. doi:10.1038/nrc2607
- 9. Wilt TJ, MacDonald R, Hagerty K, Schellhammer P, Kramer BS. Five-alpha-reductase Inhibitors for prostate cancer prevention. J Cochrane Database Syst Rev. 2008;2:CD007091. doi:10.1002/14651858.CD007091
- 10. Teyeb H, Zanina N, Neffati M, Douki W, Najjaar MF. Cytotoxic and antibacterial activities of leaf extracts of Astragalus gombiformis Pomel (Fabaceae) growing wild in Tunisia. Turk J Biol. 2012;36(1):53-8. doi:10.3906/biy-1010-131
- 11. Danchev N, Nikolova I. Pharmacological treatment of cognitive impairments in Alzheimer's disease. Autonomic Autocoid Pharmacol. 2006;26(1):46-9.
- 12. Woodruff-Pak DS, Vogel RW III, Wenk G. Galantamine: Effect on nicotinic receptor bin ding, acetylcholinesterase inhibition, and learning. Proc Natl Acad Sci USA. 2001;98(4):2089-94. doi:10.1073/pnas.98.4.2089
- Ago Y, Koda K, Takuma K, Matsuda T. Pharmacological aspects of the acetylcholinesterase inhibitor Galantamine. J Pharmacol Sci. 2011;116(1):6-17. doi:10.1254/jphs.11r01cr
- Sharp BM, Yatsula M, Fu Y. Effects of galantamine, a nicotinic allosteric potentiating ligand, on nicotine-induced catecholamine release in hippocampus and nucleus accumbens of rats. J Pharmacol Exp Ther. 2004;309(3):1116-23. doi:10.1124/jpet.103.063586
- 15. Tsvetkova D, Obreshkova D, Zheleva-Dimitrova D, Saso L. Antioxidant activity of galanthamine and some of its derivatives. Curr Med Chem. 2013;20(36):4595-608. doi:10.2174/09298673113209990148
- Azadpour M, Farajollahi MM, Varzi AM, Hadipour F, Barati M. The evaluation of cytotoxicity effects of Rheum ribes L. (rhubarb) extract on cancer cell lines and its antibacterial and mutagenicity activity. Entomol Appl Sci Lett. 2020;7(3):7-12.

Pharmacophore, 14(2) 2023, Pages 111-119

- 17. Thiele DL, Lipsky PE. Modulation of human natural killer cell function by L-Leucine methyl ester: Monocyte dependent depletion from human peripheral blood mononuclear cells. J Immunol. 1985;134(2):786-93.
- 18. Mosmann T. Rapid colorimetric assay for cellular growth and survival. Application to proliferation and cytotoxicity assays. J Immunol Methods. 1983;65:55-63.
- 19. Halimah E, Hendriani R, Ferdiansyah F. Antiproliferative activity of Acalypha wilkesiana against human cervical cancer cell lines HeLa. J Adv Pharm Educ Res. 2021;11(4):7-10. doi:10.51847/jsMgvvrBMs
- 20. Vezenkov LT, Georgieva MG, Danalev DL, Ivanov TB, Ivanova GI. Synthesis and characterization of new Galanthamine derivatives comprising peptide moiety. Protein Pept Lett. 2009;16(9):1024-8. doi:10.2174/092986609789055412
- Vezenkov L, Sevalle J, Danalev D, Ivanov T, Bakalova A, Georgieva M, et al. Galantamine-based hybrid molecules with acetylcholinesterase, butyrylcholinesterase, and γ-secretase inhibition activities. Curr Alzheimer Res. 2012;9(5):600-5. doi:10.2174/156720512800618044
- Tsvetkova D, Zheleva-Dimitrova D, Obreshkova D. Estimation of antioxidant activity of new peptide esters of Galanthamine by applying of Ferric reducing antioxidant power (FRAP) method. Compt Rend Acad Bulg Sci. 2013;66(3):445-50.
- 23. Halimah E, Hendriani R, Indradi B, Sofian FF. Cytotoxicity of ethanol extract and its fractions from Acalypha wilkesiana against breast cancer cell MCF-7. J Adv Pharm Educ Res. 2022;12(1):17-20. doi:10.51847/G2bMkvc6PO
- 24. Jemal A, Murray T, Ward E, Samuels A, Tiwari RC, Ghafoor A, et al. Cancer statistics. CA Cancer J Clin. 2005;55(1):10-30. doi:10.3322/canjclin.55.1.10
- 25. Bommareddy A, Eggleston W, Prelewicz S, Antal A, Witczak Z, Mccune DF, et al. Chemoprevention of prostate cancer by major dietary phytochemicals. Anticancer Res. 2013;33(10):4163-74.
- 26. Feldman BJ, Feldman D. The development of androgen-dependent prostate cancer. Nat Rev Cancer. 2001;1(1):34-45. doi:10.1038/35094009
- 27. Sugie S, Mukai S, Tsukino H, Toda Y, Yamauchi T, Nishikata I, et al. Increased plasma caveolin-1 levels are associated with the progression of prostate cancer among Japanese men. Anticancer Res. 2013;33(5):1893-7.
- 28. Schallier D, Decoster L, Braeckman J, Fontaine C, Degrève J. Docetaxel in the treatment of metastatic castration-resistant prostate cancer (mCRPC): An observational study in a single institution. Anticancer Res. 2012;32(2):633-41.
- 29. Adjakly M, Ngollo M, Boiteux JP, Bignon YJ, Guy L, Bernard-Gallonn D. Genistein and Daidzein: Different molecular effects on prostate cancer. Anticancer Res. 2013;33(1):39-44.
- 30. Russell LH, Mazzio E, Badisa RB, Zhu ZP, Agharahimi M, Oriaku ET, et al. Autoxidation of Gallic acid induces ROSdependent death in human prostate cancer LNCaP Cells. Anticancer Res. Anticancer Res. 2012;32(5):1595-602.
- Sutoris K, Rakusan J, Karaskova M, Mattova J, Benes J, Nekvasil M, et al. Novel topical photodynamic therapy of prostate carcinoma using hydroxy aluminum phthalocyanine entrapped in liposomes. Anticancer Res. 2013;33(4):1563-8.
- 32. Ray R, Banks M, Abuzahra H, Eddy VJ, Persons KS, Lucia MS, et al. Effect of dietary Vitamin D and calcium on the growth of androgen-insensitive human prostate tumor in a murine model. Anticancer Res. 2012;32(3):727-31.
- 33. Chen TC, Sakaki T, Yamamoto K, Kittaka A. The roles of cytochrome P450 enzymes in prostate cancer development and treatment. Anticancer Res. 2012;32(1):291-8.
- Alimirah F, Chen J, Basrawala Z, Xin H, Choubey D. DU-145, and PC-3 human prostate cancer cell lines express androgen receptor: Implications for the androgen receptor functions and regulation. FEBS Lett 2006;580(9):2294-300. doi:10.1016/j.febslet.2006.03.041
- 35. Moulay M, Liu W, Willenbrock S, Sterenczak KA, Carlson R, Ngezahayo A, et al. Evaluation of stem cell marker gene expression in canine prostate carcinoma and prostate cyst-derived cell lines. Anticancer Res. 2013;33(12):5421-31.
- Kaighn ME, Narayan KS, Ohnuki Y, Lechner JF, Jones LW. Establishment and characterization of a human prostatic carcinoma cell line (PC-3). Invest Urol. 1979;17(1):16-23.
- 37. Çelik K, Toğar, B, Türkez H, Taşpinar N. In vitro cytotoxic, genotoxic, and oxidative effects of acyclic sesquiterpene farnesene. Turk J Biol. 2014;38(2):253-9. doi:10.3906/biy-1309-55
- Chinni SR, Li Y, Upadhyay S, Koppolu PK, Sarkar FH. Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest, and apoptosis in prostate cancer cells. Oncogene. 2001;20(23):2927-36. doi:10.1038/sj.onc.1204365
- 39. Chen Z, Rasul A, Zhao C, Millimouno FM, Tsuji I, Yamamura T, et al. Antiproliferative and apoptotic effects of pinocembrin in human prostate cancer cells. Bangladesh J Pharmacol. 2013;8(3):255-62. doi:10.3329/bjp.v8i3.14795
- 40. Li H, Kapur A, Yang JX, Srivastava S, McLeod DG, Paredes-Guzman JF, et al. Antiproliferation of human prostate cancer cells by ethanolic extracts of Brazilian propolis and its botanical origin. Int J Oncol. 2007;31(3):601-6.
- Chuu CP, Lin HP, Ciaccio MF, Kokontis JM, Hiipakka RA, Liao S, et al. Caffeic acid phenethyl ester suppresses the proliferation of human prostate cancer cells through the Inhibition of p70S6K and Akt signaling networks. Cancer Prev Res. 2012;5(5):788-97. doi:10.1158/1940-6207.CAPR-12-0004-T
- 42. Mukhopadhyay A, Bueso-Ramos C, Chatterjee D, Pantazis P, Aggarwal BB. Curcumin downregulates cell survival mechanisms in human prostate cancer cell lines. Oncogene. 2001;20(52):7597-609. doi:10.1038/sj.onc.1204997
- 43. Lee DH, Szczepanski M, Lee YJ. Role of Bax in quercetin-induced apoptosis in human prostate cancer cells. Biochem Pharmacol. 2008;75(12):2345-55. doi:1016/j.bcp.2008.03.013

Pharmacophore, 14(2) 2023, Pages 111-119

- 44. Pratheeshkumar P, Son YO, Budhraja A, Wang X, Ding S, Wang L, et al. Luteolin inhibits human prostate tumor growth by suppressing vascular endothelial growth factor receptor 2-mediated angiogenesis. PLoS One. 2012;7(12):e52279. doi:10.1371/journal.pone.0052279
- Wang TTY, Hudson TS, Wang TC, Remsberg CM, Davies NM, Takahashi Y, et al. Differential effects of resveratrol on androgen-responsive LNCaP human prostate cancer cells in vitro and in vivo. Carcinogenesis. 2008;29(10):2001-10. doi:10.1093/carcin/bgn131
- Yesil-Celiktas O, Sevimli C, Bedir E, Vardar-Sukan F. Inhibitory effects of rosemary extracts, carnosic acid, and rosmarinic acid on the growth of various human cancer cell lines. Plant Foods Hum Nutr. 2010;65(2):158-63. doi:10.1007/s11130-010-0166-4
- 47. Huang JK, Chen WC, Huang CJ, Hsu SS, Chen JS, Cheng HH, et al. Nordihydroguaiaretic acid-induced Ca2+ handling and cytotoxicity in human prostate cancer cells. Life Sci. 2004;75(19):2341-51. doi:10.1016/j.lfs.2004.04.043
- McEleny K, Coffey R, Morrissey C, Fitzpatrick JM, Watson RW. Caffeicacid phenethyl ester-induced PC-3 cell apoptosis is caspase-dependent and mediated through the loss of inhibitors of apoptosis proteins. BJU Int. 2004;94(3):402-6. doi:10.1111/j.1464-410X.2004.04936.x
- 49. Horie S. Chemoprevention of prostate cancer: Soy isoflavones and curcumin. Korean J Urol. 2012;53(10):665-72. doi:10.4111/kju.2012.53.10.665
- Dorai T, Gehani N, Katz A. Therapeutic potential of curcumin in human prostate cancer. I. Curcumin induces apoptosis in both androgen-dependent and androgen-independent prostate cancer cells. Prostate Cancer Prostatic Dis. 2000;3(2):84-93. doi:10.1038/sj.pcan.4500399
- 51. Al Mojel SA, Ibrahim SF, Alshammari LK, Zadah MH, Ghamdi RNA, Thaqfan DAA. Saudi population awareness and attitude regarding stem cell donation. Arch Pharm Pract. 2021;12(1):85-9. doi:10.51847/X6pE71yCtN
- 52. Iqbal B, Kumar H, Vishwanathan V, Zaheer M, Gore C. Urinary bladder primary squamous cell carcinoma: A rare case description and literature review. Clin Cancer Investig J. 2023;12(1):11-3. doi:10.51847/NnM18XSmgJ
- Nancy A, Sukinah A, Maram A, Sara A, Hiba A, Manar A. Dental and skeletal manifestation of Sickle-cell anaemia and thalassemia in Saudi Arabia; A systematic review. Int J Pharm Res Allied Sci. 2021;10(3):1-7. doi:10.51847/MqER5p763n
- 54. Sarami S, Dadmanesh M, Hassan ZM, Ghorban K. Study on the effect of ethanol ginger extract on cell viability and p53 level in breast and pancreatic cancer. Arch Pharm Pract. 2020;11(3):115-21.
- 55. Dsouza TS. Cell culture and microscopy as research aids in conservative dentistry and endodontics. Ann Dent Spec. 2021;9(4):12-5. doi:10.51847/jFEn5AEnsF
- 56. Tsvetkova DD, Obreshkova DP, Petkova VB, Atanasov PY, Malik R, Siddiq S, et al. Investigation of antiproliferative activity of Galanthamine peptide derivate GAL-VAL against 3T3 cell lines. World J Pharm Pharm Sci. 2014;3:10-9.
- 57. Tsvetkova D, Klisurov R, Obreshkova D, Atanasov P. Effect of 6-O-N-[N-(3,4-dichlorophenyl)-D, L-Alany]-L-Leucyl-Glycine-Galantamine on 3T3 cells viability. Pharmacia. 2014;61(1):22-6.
- 58. Vezenkov LT, Georgieva MG, Danalev DL, Ivanov CB, Bakalova AT, Hristov KK, et al. Galantamine derivatives, a method for their obtaining and use. European Patent: EP 2 123 328 A1. Bulletin 48/2009. Application N: 09472001.8.