

CHEMICAL COMPOSITION AND ANTIOXIDANT, ANTI-INFLAMMATORY, AND ANTICANCER EFFECTS OF ETHANOL EXTRACT OF BLACK SHALLOT (*ALLIUM ASCALONICUM*)

Gia-Buu Tran^{1*}, Ngoc-Tuan Nguyen¹, Hoai-Nguyen Nguyen², Hong-Hieu Pham¹
Thanh Minh Thi Ngo³

1. *Institute of Biotechnology and Food-technology, Industrial University of Ho Chi Minh City, Vietnam.*
2. *Faculty of Biotechnology, Ho Chi Minh City Open University, Vietnam.*
3. *Laboratory of MICCO-NAMBO Mining Chemical Industry Company Limited, Vietnam.*

ARTICLE INFO

Received:

11 Jan 2020

Received in revised form:

01 May 2020

Accepted:

12 May 2020

Available online:

28 Jun 2020

Keywords: Anticancer activity, anti-inflammatory activity, antioxidant, black shallot, ESI-MS/MS

ABSTRACT

Introduction: Black shallot is the novel product of shallot (*Allium ascalonicum*), one of the most famous spices and remedies used in Vietnamese cuisine and in folk medicine, via aging process in a controlled temperature and humidity environment. However, the bioactivities of black shallot ethanol extract have not been studied yet. **Materials & Methods:** The aims of this study are determination of chemical composition and antioxidant, anticancer, anti-inflammatory effects of black shallot extract. The chemical constituents of black shallot extract were determined via ESI-MS/MS analysis and the antioxidant activity of extract was evaluated by 1,1-diphenyl-2-picrylhydrazil (DPPH) radical scavenging assay. The anticancer activity of black shallot against human cervical carcinoma (Hela) and human hepatocellular carcinoma (HepG2) cell lines was tested via the MTT assay. The anti-inflammatory effect of extract was measured through quantification of nitrite oxide production using the Griess reaction. **Results & Discussion:** We identified the presence of 7 bioactive compounds in black shallot extract, including quercetin 3,4'-diglucoside, isorhamnetin 3,4'-diglucoside, quercetin 3-glucoside, quercetin 4'-glucoside, isorhamnetin 4'-glucoside, quercetin aglycone, isorhamnetin. Moreover, black shallot extract exhibited the stronger antioxidant activity than fresh shallot extract ($25.67 \pm 0.65\%$ versus $18.21 \pm 0.35\%$, respectively). Of note, anticancer and anti-inflammatory effects of black shallot were also higher than those of fresh shallot. **Conclusion:** The data suggest black shallot as one of the promising functional foods for treatment of inflammatory and cancer related diseases.

Copyright © 2013 - All Rights Reserved - Pharmacophore

To Cite This Article: Gia-Buu Tran, Ngoc-Tuan Nguyen, Hoai-Nguyen Nguyen, Hong-Hieu Pham, Thanh Minh Thi Ngo, (2020), "Chemical composition and antioxidant, anti-inflammatory, and anticancer effects of ethanol extract of black shallot (*Allium ascalonicum*)", *Pharmacophore*, **10(3)**, 30-37.

Introduction

Plants used as traditional medicine contain a wide range of substances that can be used to treat chronic and infectious diseases [1, 2]. Shallot (*Allium ascalonicum* L.) is a member of the Amaryllidaceae family, which consists of several famous spices and remedies used in Vietnamese cuisine and in folk medicine such as garlic, chive, leek... Shallot is rich in starch, vitamin A, B, C, and bioactive compounds, such as selenium and sulfur-containing compounds, mannose-specific lectin, quercetin, and its glycosides, furostanol saponins, etc. [3-5]. In Nigerian folk medicine, shallot could be used to treat impaired vision, convulsion, and dysentery [6]. On the other hand, Amin et al. suggest that shallot exhibits strong antibacterial and anti-fungal activities and can be used to treat dermatomycosis and other infectious diseases [7]. The other beneficial effects of shallot including antioxidant, anticancer, anti-allergic, hypoglycemic, and hepatoprotective effects are well documented in the literature [4, 8-11]. For example, Mohammadi-Motlagh (2011) proves that the aqueous extract of shallot can inhibit the proliferation of human acute T cell leukemia (Jurkat cells), human erythroleukemia (K562 cells), and mouse fibrosarcoma (Wehi164) cell lines and possesses a remarkable anti-inflammatory effect [4]. Furthermore, Wongmekiat et al. (2008) suggested that shallot exhibits strong antioxidant and free radical scavenging activities, which in turn protects the kidney from cyclosporine-induced nephrotoxicity [10].

Corresponding Author: Gia-Buu Tran (Ph.D.), Department of Biotechnology, Institute of Biotechnology and Food-technology, Industrial University of Ho Chi Minh city, 12 Nguyen Van Bao Street, Ward 4, Go Vap District, Ho Chi Minh City, Vietnam.
Email: giabuu06cs@gmail.com

Recently, black shallot, a novel processed product of shallot, has been studied and developed for eliminating the offensive flavor of fresh shallot via the aging process in high temperature and humidity. Black shallot possesses a particular dark-brown color, gummy texture accompanied by a fruit-like sweet taste. The aging process alters not only the sensory attributes of shallot but also its chemical constituents. In a previous study, Moreno-Rojas *et al.* suggested that the concentration of some flavonoids, amino acids, and organosulfur compounds of black shallot changed during the aging process [12]. The difference in chemical composition between fresh and black shallot may lead to the difference in their bioactivities. However, the bioactivities of black shallot's ethanolic extract have not been studied yet. The aims of this study were the determination of chemical composition and some bioactivities such as anti-oxidative, anticancer, and anti-inflammatory effects of black shallot extract, which provides more information to utilize this product as one of the potential functional foods in future.

Materials and Methods

Chemicals and Materials

Shallot (*Allium ascalonicum* L.) was obtained from Vinh Chau Town, Soc Trang Province in March 2018, the voucher specimens were identified by a taxonomist at Institute of Biotechnology and Food-technology, Industrial University of Ho Chi Minh City, Vietnam

Lipopolysaccharide (LPS), 1,1-Diphenyl-2-picrylhydrazyl (DPPH), 3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide (MTT), Griess reagent, dimethyl sulfoxide (DMSO), Eagle's Minimum Essential Media (EMEM) were obtained from Sigma-Aldrich Chemical Company (St. Louis, MO, USA). Dulbecco's Modified Eagle Media, L-glutamine, streptomycin, penicillin, and fetal bovine serum (FBS) were provided by Thermo Fisher Scientific Inc. (Waltham, MA, USA). All reagents were of analytical or cell culture grades.

Preparation of black shallot extract

Shallots were rinsed with water and peeled off the outside husks to remove the dust and damaged parts. The whole shallots subsequently were fermented in the aging chamber (Shellab, USA) in accordance with Tran *et al.* procedure [13]. Concisely, shallots were incubated at 75°C in 90% relative humidity for 20 days to produce particular dark-brown products, black shallots (Figure 1). Black shallots or flesh shallots were sliced and immersed with 5 volumes of 98% ethanol solution for 1 week, and this step was repeated 2 times to obtain the maximum yield. The resulting extracts were combined and filtrated through a Whatman filter paper. Filtrates were eventually condensed in reduced pressure (130 mmBar) at 60°C to obtain the resulting extracts (about ½ of the initial filtrate volume). The ethanol residue in extracts was eliminated via sublimation drying, then the black shallot (BSE) and fresh shallot (FSE) extracts were stored at 4°C until further use.

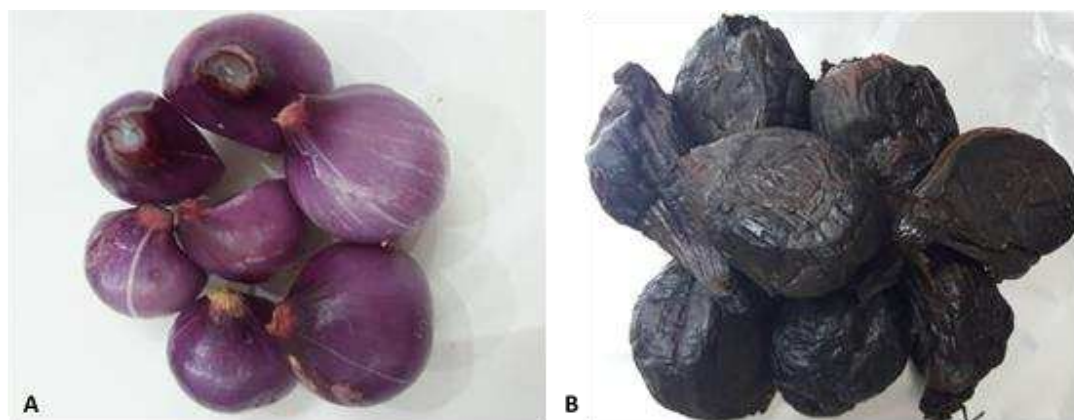


Figure 1. Black shallot is the novel processed product of fresh shallot via the aging process in a temperature- and humidity-controlled chamber. Fresh shallot (A), black shallot after aging for 20 days (B).

ESI-MS/MS analysis

ESI-MS/MS analysis was performed to elucidate chemical constituents of black shallot extract (BSE). In brief, the aliquots of BSE were injected into the Dionex UltiMate 3000 HPLC system coupled with the amaZon SL mass spectrometer (Bruker Daltonics Inc., Germany). Column C18 Poroshell (2.1 × 150 mm, 4 µm) was used as the stationary phase and the mobile phase was the mixture of 0.1% formic acid in water (solvent A) and acetonitrile (solvent B) in step gradient elution modified from the method of Bonaccorsi *et al.* [14]. The chromatographic separation was set as gradient elution from 5–20 % B (0-5 min), 20–30 % B (5-10 min), 30–35 % B (10-20 min), 35–50 % B (20-35 min), 50–95 % B (35-38 min), and staying at 95% B, then turning back to 5% B and equilibration for 4 minutes, with total run time about 45 minutes. The flow rate was 0.3 mL/min and the injection volume was 2 µL. The wavelengths were set at 370 and 260 nm. ESI-MS was equipped with a quadrupole ion trap. Nebulizer gas was set at 2bars, the capillary was set at 4,500V, and drying gas temperature was 220°C with a flow rate of 7.0 L/min. MS assessment was done in both negative and positive modes and scanned at the mass range

of m/z 80-1500 amu. HyStar, Trap control, and Chromelon were used as controlling programs. quercetin 3-glucoside (Chengdu Biopurify Phytochemicals Ltd.) and Quercetin hydrate (Sigma Aldrich) were used as standards.

Determination of the antioxidant activity of black shallot extract

The antioxidant activity of BSE and FSE was measured via the 1,1-diphenyl-2-picryl-hydrazil (DPPH) radical scavenging assay according to the method of Kumar *et al.* with some modifications [15]. Briefly, the 0.1 mM DPPH solution was prepared in ethanol, and extracts were diluted in DMSO to obtain the 4 mg/mL solution. Then, 0.1 mM DPPH solution was mixed with a volume of ethanol extract to obtain the final extract concentration of about 200 µg/mL. The mixture was vigorously shaken and stand at room temperature for 30min, after which the absorbance of the sample was measured at 515 nm using a microplate reader (Infinite 50, Tecan, Switzerland). The mixture without ethanol extract was used as the control and ascorbic acid solution (final concentration of 44 µg/mL) was used as positive control. DPPH radical scavenging activity was calculated as the following formula: DPPH radical scavenging activity (%) = $\frac{A_0 - A_1}{A_0} \times 100\%$; in which, A_0 is the absorbance of control at 515 nm, and A_1 is the absorbance of the sample at 515 nm.

Cell culture

Human hepatocellular carcinoma (HepG2), human cervical carcinoma (Hela), and murine macrophage (Raw 264.7) cell lines were provided by American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were cultured in appropriate media (DMEM or EMEM) containing 10% heat-inactivated FBS, penicillin (100 units/mL), and streptomycin (100 µg/mL) and incubated at 37°C in a humidified incubator containing 5% CO₂.

Determination of the anti-inflammatory activity of black shallot extract

The anti-inflammatory activity of the extract was evaluated via quantification of nitrite oxide production using the Griess reaction [16]. In brief, mouse macrophage cells (Raw 264.7) were sub-cultured into 96-well plate with a density of about 2.5×10^4 cells/well for 24 hours and subsequently incubated with or without black shallot extracts (30-100 µg/mL) for 2 hours. Cardamomin (0.3-3.0 µM) was used as a positive control. Then cells were stimulated with 2 µL LPS (0.1 mg/mL) for 24 hours. The cell culture media was incubated with Griess reagent (ratio 1:1) and kept at room temperature for 10 min. The absorbance of the sample was recorded at 540 nm via a microplate reader (Infinite 50, Tecan, Switzerland). The nitrite concentration in each sample was calculated using a serial dilution standard calibration curve prepared with sodium nitrate. Cell viability of macrophages was measured by the 3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide MTT assay [17, 18]. Briefly, 264.7 cells after treatment with extracts or reagents were incubated with MTT solution (5 mg/mL in phosphate-buffered saline) at 37°C in a 5% CO₂ humidified atmosphere for 4 hours. The formazan crystal was dissolved by adding 100 µL of DMSO (Sigma-Aldrich, St. Louis, MO, USA) and absorbance of the solution was measured at 540 nm via a microplate reader (Infinite 50, Tecan, Switzerland). The cell viability and inhibition of nitrite oxide production were calculated and expressed as the average percentage of three analyses.

Determination of the anticancer effect of black shallot extract

Anticancer properties of the extract against human hepatocellular cancer (HepG2) and cervical cancer (Hela) cells were evaluated by the 3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide MTT assay [19]. Briefly, HepG2 and Hela were cultured in 96-well plates with appropriate density (0.8×10^4 and 0.4×10^4 cells/well, respectively) for 24 hours. Then, the media were carefully replaced with fresh media containing extracts (final concentration of 100 µg/mL) or positive control (Paclitaxel-final concentration 5 mM) and incubated for 48h. Media were removed and cells were incubated with MTT solution (5 mg/mL in phosphate-buffered saline) at 37°C in a 5% CO₂ humidified atmosphere for 4 hours. The formazan crystal was dissolved by the addition of 100 µL DMSO (Sigma-Aldrich, St. Louis, MO, USA) and the absorbance of the solution was recorded at 540 nm via a microplate reader (Infinite 50, Tecan, Switzerland). Cytotoxicity of the extract against HepG2 and Hela was determined and expressed as an average percentage of three analyses.

Statistical analysis

All determinations were done in triplicate. The experimental results were expressed as mean±standard deviation, and the Statgraphics Centurion XVI software (Statpoint Technologies Inc., Warrenton, Virginia, USA) was used to conduct statistical analysis. Difference between different treated groups were analysed using ANOVA test and multiple range tests with the criterion of statistical significance less than 0.05 ($p < 0.05$).

Results and Discussions

Chemical constituents of black shallot extract

To determine the chemical constituents of black shallot extract (BSE), ESI-MS/MS analysis was performed and the results are shown in Table1. HPLC chromatograms of quercetin 3-glucoside, quercetin hydrate, and shallot extracts are shown in Figure 2. Of note, the corresponding peak of quercetin aglycone eluted at the same retention time as quercetin hydrate was observed. Furthermore, the ESI-MS spectra in positive and negative modes of quercetin hydrate and quercetin 3- glucoside also confirmed the eluted peaks in shallot extract. ESI-MS spectra in the negative mode of BSE exhibited similar elution

order and m/z of the chemical compounds in line with the results of a study conducted by Bonaccorsi *et al.*, 2005. We identified the presence of seven compounds in black shallot extract, such as (1) quercetin 3,4'-diglucoside (MW=626) and (2) isorhamnetin 3,4'-diglucoside (MW=640) in RT for 7.6-7.9 min, (3) quercetin 3-glucoside (MW=464) with RT for about 9 min, (4) quercetin 4'-glucoside (MW=464) and (5) isorhamnetin 4'-glucoside (MW=478) in RT for 10.4-10.9 min, (6) quercetin aglycone (MW=302) and (7) isorhamnetin (MW=316) in RT for 16.8 min. The representative molecular ions in the negative mode shallot extracts, corresponding to m/z of 477, 463, 639, and 625 are presented in Figure 3. The molecular weight of quercetin aglycone is 302 and that of quercetin 3-glucoside is 464. Quercetin 3-glucoside could lose glycosyl moiety leading to fragment ion m/z 302. The typical fragmentation pattern of m/z 302 gave fragment ions 179 and 151. ESI-MS/MS of BSE revealed the same fragmentation pattern as those of quercetin aglycone and the fragment ion of quercetin 3-glucoside (Figure 4).

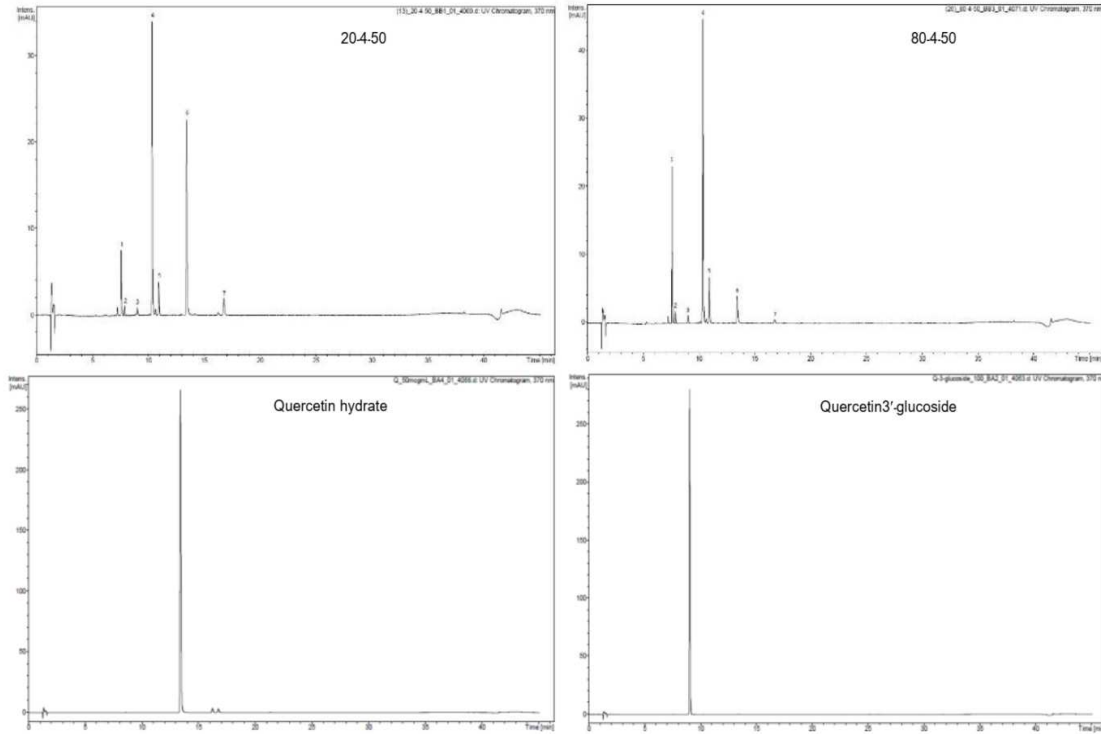
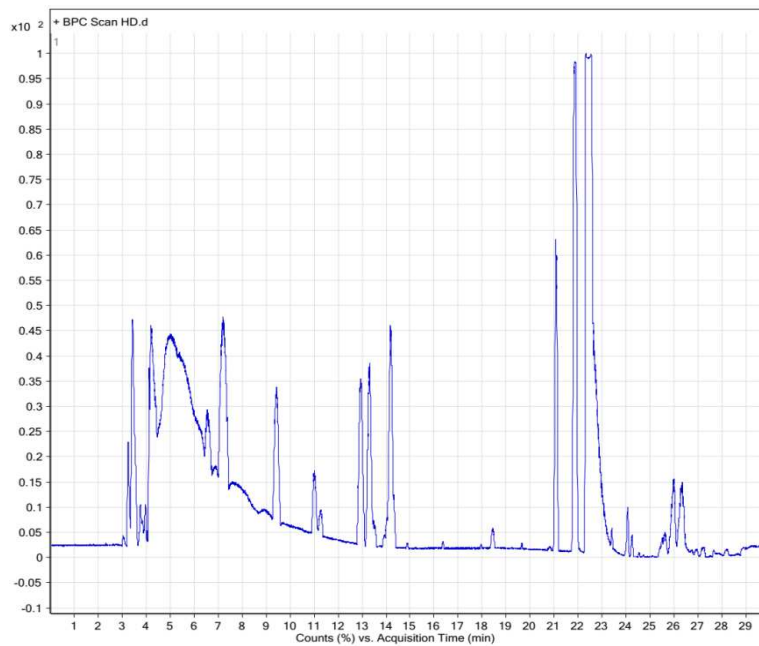


Figure 2. HPLC chromatograms of quercetin 3-glucoside, quercetin hydrate, and shallot extracts at 370nm



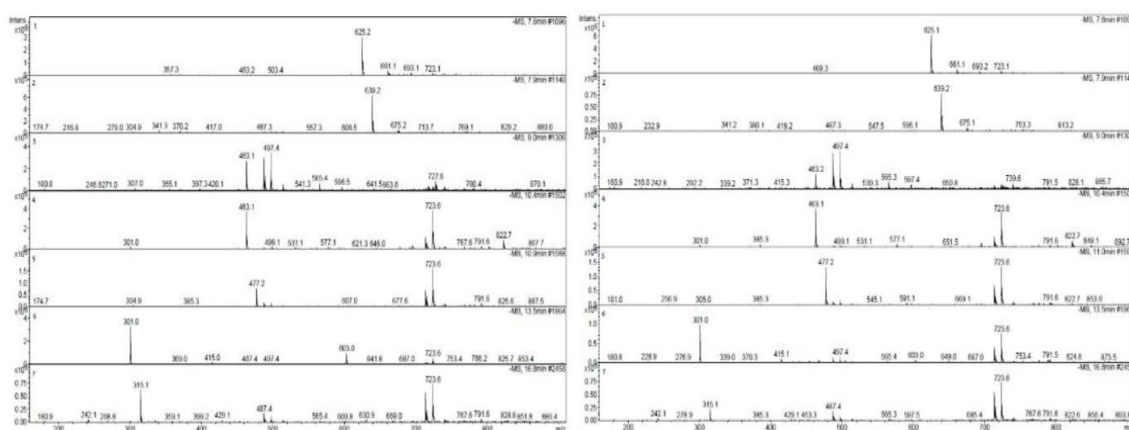


Figure 3. LC and ESI-MS spectra of shallot extracts

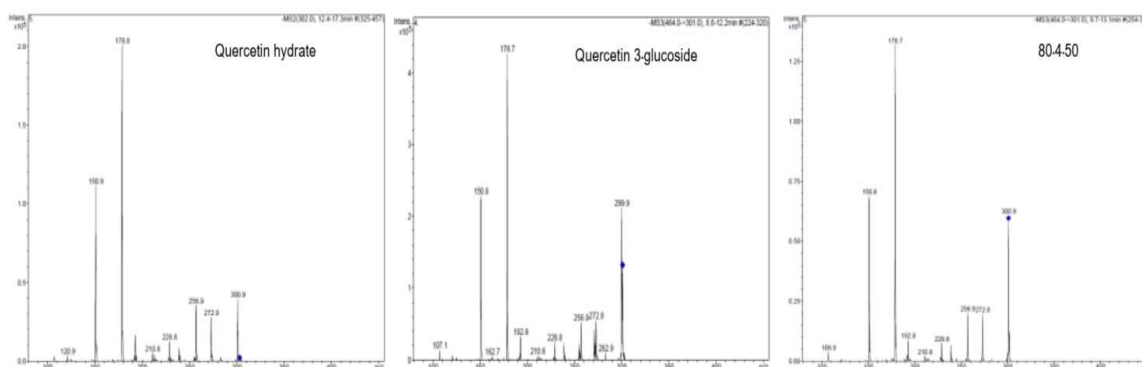


Figure 4. MS/MS fragmentation of $m/z=302$, quercetin hydrate, quercetin-3-glucoside.

The biological effects of those compounds have been well documented in the literature. For example, Zhou *et al.* reported that isorhamnetin exerted anticancer activity against human hepatoma cells (HepG2) and murine colon carcinoma cells (CT26), as well as free radical scavenging and anti-inflammatory activities [20]. Furthermore, quercetin aglycone is the ubiquitous flavonoid, which exists in several fruits and vegetables and exerts many bioactivities, such as anticancer, antioxidant, anti-diabetic, and anti-inflammatory effects [21]. According to Kang *et al.* (2013), quercetin aglycone could protect microglial cells from LPS induced nitric oxide production via downregulation of the NF- κ B signaling pathway and activating of the Nrf2-dependent HO1-1 pathway, which in turn alleviates the nitrosative stress in the nerve system [22]. In previous studies, both quercetin 3-glucoside and quercetin 4'-glucoside exhibited the anti-oxidative effect via suppressing iron ion driven lipid peroxidation in the intestinal tract, but quercetin 4'-glucoside was suggested as the more efficient antioxidant as compared to quercetin 3-glucoside [23]. Williamson *et al.* also suggested the anticancer effect of quercetin 4'-glucoside via induction of quinone reductase, the anticarcinogenic phase II marker enzyme in mouse hepatoma (Hepalclc7) cells [24]. Moreover, Boyle *et al.* also indicated that flavonoid glycosides, such as quercetin-3-glycoside and isorhamnetin 4'-glucoside, increased in plasma level after onion meal and were related with the enhancement of resistance to DNA strand breakage in lymphocytes [25]. In this study, ESI-MS/MS has identified some chemical constituents in black shallot extracts; however, further experiments have to undertake in order to clarify the relationship with their bioactivities.

Table 1. Phytochemical composition of ethanol extract of black shallot

No	Compounds	Chemical formulas	Molecular Weight	Retention Time (min)	Bioactivities
1	Quercetin 3,4'-diglucoside	$C_{27}H_{30}O_{17}$	626	7.6-7.9	
2	Isorhamnetin 3,4'-diglucoside	$C_{28}H_{32}O_{17}$	640	7.6-7.9	
3	Quercetin 3-glucoside	$C_{21}H_{20}O_{12}$	464	9.0	Anti-oxidative effect
4	Quercetin 4'-glucoside	$C_{21}H_{20}O_{12}$	464	10.4-10.9	Anti-oxidative, anti-cancer effects
5	Isorhamnetin 4'-glucoside	$C_{22}H_{22}O_{12}$	478	10.4-10.9	DNA protective effect
6	Quercetin aglycone	$C_{15}H_{10}O_7$	302	16.8	Anti-oxidative, anti-inflammatory, anticancer, anti-diabetic effects
7	Isorhamnetin	$C_{16}H_{12}O_7$	316	16.8	Anti-inflammatory, anti-oxidative, anti-cancer effects

DPPH radical scavenging activity

The antioxidant effect of black shallot extract was evaluated via DPPH radical scavenging activity. As shown in Figure 5, ascorbic acid exhibited the strongest antioxidant activity with $89.27 \pm 1.10\%$, next coming with black shallot extract and fresh shallot with $25.67 \pm 0.65\%$, versus $18.21 \pm 0.35\%$, respectively. The antioxidant activity of black shallot extract may be explained through the existence of some antioxidant compounds including isorhamnetin, quercetin 4'-glucoside, quercetin 3-glucoside, and quercetin aglycone. Of note, the antioxidant activity of black shallot extract is 1.4 folds as high as that of fresh shallot extract, which provides the evidence of changes of chemical composition, as well as bioactivity of black shallot via the aging process.

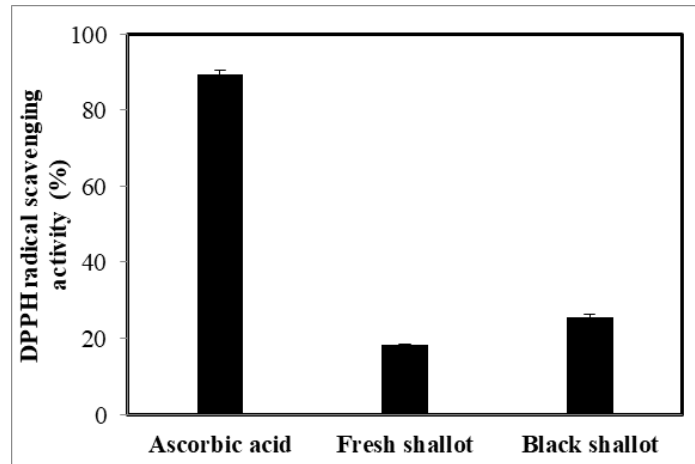


Figure 5. Antioxidant activity of black shallot ethanol extract was determined via DPPH radical scavenging activity.

Anti-inflammatory effect of black shallot extract

The anti-inflammatory effect of black shallot extract was measured via inhibition of nitric oxide (NO) production in Raw 264.7 cells. It was observed that the elevation of inhibition action of NO production by LPS-stimulated macrophages was associated with the increase of concentration of black shallot extract, fresh shallot extract, and Cardamonin. Moreover, the capacity of black shallot extract to regulate the NO production was higher than that of fresh shallot extract at the same concentration (Table 2). For example, BSE at the concentration of 100 µg/mL could inhibit $32.57 \pm 0.64\%$ of the NO production while FSE only inhibited $15.39 \pm 0.54\%$ at the same concentration. On the other hand, both shallot extracts (BSE and FSE) exhibited low toxicity to macrophages (cell viability over 85%), and cell viability of BSE was not significantly different from reference drug (Cardamonin, 0.3-3.0 µM). These results were in line with the data from Mohammadi-Motlagh *et al.* experiments [4]. In a previous study, shallot aqueous extract exerted the low toxicity in human umbilical vein endothelial cells (HUVEC) and a significant anti-inflammatory effect *in vivo* via the acetic acid-induced vascular permeability model. Of note, the cell viability of BSE at the concentration of 30 µg/mL was not remarkably different from that of FSE at the concentration of 100 µg/mL but the anti-inflammatory effect of BSE was higher than that of the FSE ($19.84 \pm 0.67\%$ and $15.39 \pm 0.54\%$, respectively), which implied black shallot as a natural anti-inflammatory reagent.

Table 2. Anti-inflammatory effect of black shallot ethanol extract

		Inhibition of NO production (%)	Cell viability (%)
Cardamonin	0.3 µM	17.03 ± 0.85^a	87.39 ± 1.24^c
	3.0 µM	96.29 ± 0.51^b	86.58 ± 1.09^c
Fresh shallot extract	30 µg/mL	8.76 ± 0.41^c	90.07 ± 1.18^a
	100 µg/mL	15.39 ± 0.54^d	89.72 ± 1.14^{ab}
Black shallot extract	30 µg/mL	19.84 ± 0.67^e	87.74 ± 1.27^{bc}
	100 µg/mL	32.57 ± 0.64^f	86.52 ± 1.32^c

^{a,b,c,d,e,f} Different letters in same column indicate significant differences among different groups (p<0.05).

Anti-cancer effect of black shallot extract

The anti-cancer effects of black shallot extract and fresh shallot extract were evaluated via MTT assay and the results are presented in Table 3. The anti-cancer activities of black shallot extract against Hela cells and HepG2 ($18.58 \pm 0.45\%$ and $19.94 \pm 0.58\%$, respectively) were higher than those of fresh shallot extract ($13.65 \pm 0.42\%$ and $14.85 \pm 0.49\%$, respectively). The anticancer effect of *Allium* species has been proven in several previous studies. For example, Li *et al.* proved that intraperitoneal injection of raw garlic extract (equivalent to 100mg wet weight) for 21days could cure the highly aggressive malignancies (sarcoma 180 and EL4-induced ascites) in mouse models [26]. According to Fragis *et al.* onion extracts exhibited cytotoxic, anti-proliferation, anti-migratory effects against human breast cancer (MDA-MB-231 and MCF-7 cells) cell lines [27]. Moreover, the anticancer activity of the aqueous extract of shallot against mouse fibrosarcoma cells (Wehi164), human acute T-cell leukemia (Jurkat), and erythroleukemia (K562) cells was also reported [4]. In a

previous study, Pan *et al.* (2018) also indicated that quercetin glucosides, constituents of *Allium* species such as garlic, onion, Chinese onion, Welsh onion, could inhibit the growth of HepG2, HT-29, and PC-3 cancer cells [28]. To the best of our knowledge, this study is the first work that proved the anticancer effect of black shallot and compared the efficiency of black shallot and fresh shallot against cancer cells. These data provide the scientific basis for further application of black shallot for the treatment of cancer-related diseases.

Table 3. Anti-cancer effect of black shallot ethanol extract

	Inhibition rate on Hela cells (%)	Inhibition rate on HepG2 cells (%)
Paclitaxel	75.79 ± 0.57 ^c	97.24 ± 0.62 ^c
Fresh shallot extract	13.65 ± 0.42 ^a	14.85 ± 0.49 ^a
Black shallot shallot extract	18.58 ± 0.45 ^b	19.94 ± 0.58 ^b

^{a,b,c} Different letters in the same column indicate significant differences among different groups ($p < 0.05$).

Conclusion

In this study, the presence of 7 compounds in black shallot extract was identified, including quercetin 3,4'-diglucoside, isorhamnetin 3,4'-diglucoside, quercetin 3-glucoside, quercetin 4'-glucoside, isorhamnetin 4'-glucoside, quercetin aglycone, isorhamnetin. Moreover, black shallot extract exhibited stronger antioxidant activity than fresh shallot extract ($25.67 \pm 0.65\%$ versus $18.21 \pm 0.35\%$, respectively). Of note, anticancer and anti-inflammatory effects of black shallot were also higher than those of the fresh shallot. The data indicate black shallot as the potential functional foods for treating inflammatory and cancer-related diseases.

Acknowledgment

The authors gratefully thank the Board of Directors of the Institute of Biotechnology and Food Technology, Industrial University of Ho Chi Minh for their support of equipment during this research project.

Conflict-of- interest Notification Page:

The authors declare there is no conflict of interest regarding the publication of this paper.

Key Messages: Black shallot, a novel processing product from shallot, exhibits the stronger anti-oxidant, anti-inflammatory and anticancer properties than fresh shallot. These findings suggest black shallot ethanol extract as a promising functional food to alleviate inflammatory and cancer related diseases.

References

- Ahmad MS, Shawky A, Ghobashy MO, Felifel RH. Effect of Some medicinal plants on life cycle of Citrus Brown Mites (*Eutetranychus orientalis*). *Int. J. Pharm. Res. Allied Sci.* 2018;7(4):13-7.
- Alasmari A. Phytomedicinal potential characterization of medical plants (*Rumex nervosus* and *Dodonaea viscosa*). *J Biochem Tech.* 2020;11(1):113-21.
- Sun W, Shahrajabian MH, Cheng Q. The insight and survey on medicinal properties and nutritive components of Shallot. *Journal of Medicinal Plants Research.* 2019;13(18):452-457.
- Mohammadi-Motlagh HR, Mostafaie A, Mansouri K. Anticancer and anti-inflammatory activities of shallot (*Allium ascalonicum*) extract. *Arch Med Sci.* 2011;7(1):38-44.
- Fattorusso E, Iorizzi M, Lanzotti V, Tagliatalata-Scafati O. Chemical Composition of Shallot (*Allium ascalonicum* Hort.). *Journal of agricultural and food chemistry.* 2002;50(20):5686-5690.
- Odugbemi T. A textbook of medicinal plants from Nigeria. Yaba-Laos: University of Laos Press; 2008.
- Amin M, Montazeri EA, Mashhadizadeh, MA, Sheikh AF. Characterization of shallot, an antimicrobial extract of *Allium ascalonicum*. *Pakistan Journal of Medical Sciences.* 2009;25(6):948-952.
- Arpornchayanon W, Klinprung S, Chansakaow S, Hanprasertpong N, Chaiyasate S, Tokuda M, et al. Antiallergic activities of shallot (*Allium ascalonicum* L.) and its therapeutic effects in allergic rhinitis. *Asian Pac J Allergy Immunol* [journal on gthe Internet]. 2019 Aug 18 [cited 2020 Jun 9]. Available from: <http://apjai-journal.org/wp-content/uploads/2019/08/AP-300319-0529.pdf> [Epub ahead of print]
- Kourjampa W, Luangpirom A. Protective effect of shallot (*Allium ascalonicum*) on paracetamol-induced hepatotoxicity in mice. *Journal of Physiological and Biomedical Sciences.* 2011;24(1 Suppl):S74-77.
- Wongmekiat O, Leelarugrayub N, Thamprasert K. Beneficial effect of shallot (*Allium ascalonicum* L.) extract on cyclosporine nephrotoxicity in rats. *Food Chem Toxicol.* 2008;46(5):1844-1850.

11. Jalal R, Bagheri SM, Moghimi A, Rasuli MB. Hypoglycemic effect of aqueous shallot and garlic extracts in rats with fructose-induced insulin resistance. *J Clin Biochem Nutr.* 2007;41(3):218-223.
12. Moreno-Rojas JM, Moreno-Ortega A, Ordóñez JL, Moreno-Rojas R, Pérez-Aparicio J, Pereira-Caro G. Development and validation of UHPLC-HRMS methodology for the determination of flavonoids, amino acids and organosulfur compounds in black onion, a novel derived product from fresh shallot onions (*Allium cepa* var. *aggregatum*). *LWT-Food Science and Technology.* 2018;97 :376-383.
13. Tran, GB, Dam SM, Le TNT. Amelioration of single clove black garlic aqueous extract on dyslipidemia and hepatitis in chronic carbon tetrachloride intoxicated Swiss albino mice. *International Journal of Hepatology.* 2018;2018:9383950.
14. Bonaccorsi P, Caristi C, Gargiulli C, Leuzzi U. Flavonol glucoside profile of southern Italian red onion (*Allium cepa* L.). *J Agric Food Chem.* 2005;53(7):2733-2740.
15. Kumar GP, Navya KT, Ramya EM, Venkataramana M, Anand T, Anilakumar KR. DNA damage protecting and free radical scavenging properties of *Terminalia arjuna* bark in PC-12 cells and plasmid DNA. *Free Radicals Antioxid.* 2013;3(1): 35-39.
16. Joo T, Sowndhararajan K, Hong S, Lee J, Par SY, Kim SM et al. Inhibition of nitric oxide production in LPS-stimulated RAW 264.7 cells by stem bark of *Ulmus pumila* L. *Saudi J Biol Sci.* 2014;21(5):427-435.
17. Lee SY, Kim HJ, Han JS. Anti-inflammatory effect of oyster shell extract in LPS-stimulated Raw 264.7 cells. *Prev Nutr Food Sci.* 2013;18(1):23-29.
18. Krishnaa PK, Thangavelu L, Roy A. Anti-inflammatory activity of *Caralluma Fimbrata*-raw 264 macrophage cell lines. *J. Adv. Pharm. Educ. Res.* 2018;8(2):21-4.
19. Prashanthi K, Suryan S, Varalakshmi KN. In vitro Anticancer Property of Yellow Pigment from *Streptomyces griseoaurantiacus* JUACTION 01. *Brazilian Archives of Biology and Technology.* 2015;58(6):869-876.
20. Zhou JL, Fang XY, Wang JQ, Zhao LG, Li Y, Tang F, et al. Structures and bioactivities of seven flavonoids from *Osmanthus fragrans* 'Jinqiu' essential oil extraction residues. *Nat Prod Res.* 2018;32(5):588-591.
21. Kawabata K, Mukai R, Ishisaka A. Quercetin and related polyphenols: new insights and implications for their bioactivity and bioavailability. *Food Funct.* 2015;6(5):1399-1417.
22. Kang CH, Choi YH, Moon SK, Kim WJ, Kim GY. Quercetin inhibits lipopolysaccharide-induced nitric oxide production in BV2 microglial cells by suppressing the NF- κ B pathway and activating the Nrf2-dependent HO-1 pathway. *Int Immunopharmacol.* 2013;17(3):808-813.
23. Murota K, Mitsukuni Y, Ichikawa M, Tsushida T, Miyamoto S, Terao J. Quercetin-4'-glucoside is more potent than quercetin-3-glucoside in protection of rat intestinal mucosa homogenates against iron ion-induced lipid peroxidation. *J Agric Food Chem.* 2004;52(7):1907-1912.
24. Williamson G, Plumb GW, Uda Y, Price KR, Rhodes MJ. Dietary quercetin glycosides: antioxidant activity and induction of the anticarcinogenic phase II marker enzyme quinone reductase in Hepalcl7 cells. *Carcinogenesis.* 1996;17(11):2385-2387.
25. Boyle SP, Dobson VL, Duthie SJ, Kyle JA, Collins AR. Absorption and DNA protective effects of flavonoid glycosides from an onion meal. *Eur J Nutr.* 2000;39(5):213-223.
26. Li Z, Le W, Cui Z. A novel therapeutic anticancer property of raw garlic extract via injection but not ingestion. *Cell Death Discov.* 2018;4 :108.
27. Fragis M, Murayyan AI, Neethirajan S. Cytotoxic activity and anti-cancer potential of Ontario grown onion extracts against breast cancer cell lines. *Functional Food in Health and Disease.* 2018;8(3):159-174.
28. Pan Y, Zheng YM, Ho WS. Effect of quercetin glucosides from *Allium* extracts on HepG2, PC-3 and HT-29 cancer cell lines. *Oncol Lett.* 2018;15(4):4657-4661.