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Review Article

PHYTOCHEMICAL COMPOSITION OF ESSENTIAL OILS ISOLATED FROM DIFFERENT SPECIES OF GENUS *NEPETA* OF LABIATAE FAMILY: A REVIEW

Ajay Sharma and Damanjit Singh Cannoo*

Department of Chemistry, Sant Longowal Institute of Engineering and Technology, Longowal, Sangrur, Punjab-148106, India

ABSTRACT

Aromatic and medicinal plants have been regarded as foremost source of secondary metabolites (SMs). These SMs (less toxic and biodegradable) and their derivatives have been recognised as versatile source of biologically active drugs. The most of the drugs used in present medicinal system have direct or indirect relation with secondary metabolites. So, during last few decades the interest has increased in the study of phytochemical composition of aforementioned plants to evaluate their prospective in modern medicinal system. Amongst various plant families which have been known for their medicinal and therapeutic values, Labiatae (Mint) family (Genus *Nepeta*) remains quite important. Essential oils extracted from various parts of species of above said genus have been a vital source of terpenoid and oxygenated terpenoid hydrocarbons especially sesquiterpene hydrocarbons and their oxygenated derivatives. These compounds have been known for their inherent biological activities viz. sedative, diaphoretic, feberifuge, expectorant, diuretic, stomach tonic, antispasmodic, antipyretics, anti-viral, anti-inflammatory, antimicrobial, fungicidal, insect repellent and antidote against snakes and scorpion bites etc. The present communication constitutes a review on the chemical composition of essential oils (only key constituents) extracted from genus *Nepeta* along with the details of their specific area of collection, height, specific time in a year, stage of plant collection, method used for extraction and technique of analysis.

Keywords: Secondary metabolites, Nepeta, Phytochemical composition, Essential oils.

INTRODUCTION

Plants have been a rich source of valuable, cost effective and easily available natural products. Natural products have played a pivotal role in pharmacy agriculture due and to which inexhaustible development has been witnessed to synthesise and isolate new herbal and useful drugs. A few families viz. myrtacea, lauraceae, rutaceae. lamiaceae. asteraceae. apiaceae. zingiberaceae cupessaceae, poceace, and piperaceae have proven to be efficient in providing precious essential oils which are lipophilic in nature and are rich source of

innumerable bioactive compounds. The latter different classes of secondary belong to metabolites viz. flavonoids, carotenoids, alkaloids, phenolics, terpenoids, tannins, phenolic acids etc. These potent biologically active compounds act as alternatives to numerous synthetic pharmaceutical drugs, agrochemicals etc. and it has been a competent approach for finding new bioactive compounds extracted from plants. Further, these natural products have been isolated in their pure form and their study involves identification and characterization of

their structures, in vitro synthesis and evaluation of their pharmaceutical and agrochemical activities viz. antioxidant, anti-inflammatory, antiseptic, anti-asthmatic, diuretic, antispasmodic, anticancer, antipyretic, diaphoretic, analgesic, sedative, antimicrobial, fungicidal, herbicidal, insecticidal and insect repellent. These biological activities depend upon the composition and nature of the natural products.¹ Secondary metabolites obtained from genus Nepeta have been used in various traditional medicines from prehistoric times. Essential oils isolated from different Nepeta species have been used as laxative for the treatment of dysentery, teeth troubles, ear pain, kidney, liver diseases^{2,3} and many heart problems such as cardiac thrombosis, tachycardia, angina pectoris and weakness of the heart.^{4,5} These have also been used as feberifuge, sedative, antiseptic, anti-tussive, diaphoretic. anti-asthmatic. antioxidant.6-10 Along with this genus Nepeta showed many other biological activities viz. antianti-inflammatory,¹¹⁻¹³ atherosclerotic. expectorant, diuretic, antispasmodic,^{8,14} feline and canine attractant ¹⁵ stomach tonic, ¹⁶ antipyretics, against snakes and scorpion bites^{10,14} insecticidal, antimicrobial,¹⁷ fungicidal anti-viral and agents.18,19

N. juncea plant has been known for its biological potential like antiglycation, cytotoxic, platelet aggregation, phytotoxic and antimicrobial.²⁰ N. glomerulosa had been used to cure digestive problem, pneumonia and itching.²¹ N. menthoides has been reported for its phytotherapic properties viz. febrifuge, sedative and a relief agent for stomach pains whereas N. cataria has been used as disinfectant, fortifier and to cure colds, 10 N. racemosa as disinfectant, carminative, stomachic bracteata and N. as antiasthmatic and carminative.⁹ Catnip plant's (N. cataria) flowers and leaves find utility for the preparation of calmative herbal tea to take sound sleep. It has shown varied medicinal properties like diaphoretic, nervine, antispasmodic, emmenagogue, stimulant and mild sedative. Further, these have been used for the treatment of colic, diarrhoea, cancer, common cold, anxiety and tension. Juvenile hormone activity has also

been reported from catnip plant extract.²² Joshandah prepared from *N. ciliaris* plant has been used as efficient remedy for curing common cold, catarrh, cough and associated respiratory distress and fever in many parts of India.²³ *N. ciliaris* plant has been also used for preparing squash and liquid extract recommended against phlegm and as antipyretic, antitussive agent.²⁴⁻²⁶ All these properties showed by different species of genus *Nepeta* have been ascribed to their flavonoids and essential oil composition.²⁷

GENUS NEPETA

Nepeta is multiregional genus and belongs to Labiatae (Mint) family. The name *Nepeta* was coined after the name of ancient Italian city Nephi.²⁸ Genus *Nepeta* has 280 species spread all over the world but extensively grown in many regions of Asia, Africa, North America, central and southern part of Europe out of which 30 are found in plains and foothills of the Himalayas in India.²⁹⁻³² Taxonomic position of genus *Nepeta* is as under:

Kingdom : Plantae

- Class : Magnoliopsida
- Order : Lamiales
- Family : Lamiaceae (Labiatae)
- Genus : Nepeta³³

Most of the species of genus *Nepeta* are hairy perennial herbs having dense leaves which are covered with soft hairs. Flowers are of various colours and shapes occur in clusters bloom from July to September.³⁴

CHEMICAL COMPOSITION

Many classes of compounds have been obtained Nepeta from genus like monoterpenes (nepetalactones, dehydronepetalactone, nepetalic acid, 1,8-cineole, α -terpineol, α -citral and geraniol etc);^{7,19,35,36} sesquiterpenes (β -farnesene, α -bisbolene, β -caryophyllene and α -humulene, etc)^{.37} flavonoids (luteolin 7-O-glucuronide; apigenin 7-O-glucuronide and 7-O-glucuronoglucoside etc.); phenolic acid (caffeic, rosmarinic and p-coumaric acids, vanillic acid, gallic acid etc);³⁸ steroids (ursolic acid, oleanolic acid, β sitosterol, stigmasterol, stigmasterol glucoside, glutinol and β -amyrin etc.),³⁹ but the essential oils

isolated from different species of this genus has been mainly enriched with terpenes (mono and sesquiterpenes).

The essential oils obtained from different parts of plants viz. roots, stem, leaves, flowering parts etc. of genus Nepeta have been found to be rich in terpenoid hydrocarbons, sesquiterpene including their hydrocarbons oxygenated analogues.^{37,40} Depending upon the composition of major compounds in the essential oils, Nepeta species have been divided into two groups one that contained different isomers of nepetalactone and the second that had compounds other than isomers of nepetalactone like 1,8-cineole, β caryophyllene, caryophyllene oxide, β -farnesene, β-citronellol etc α -citral, as their major constituents.⁷

Group 1:

Species of this group contain $4a\alpha$, 7α , $7a\alpha$ nepetalactone, $4a\alpha, 7\alpha, 7a\beta$ -nepetalactone, $4a\beta$, 7α , $7a\beta$ -nepetalactone, $4a\beta,7\alpha,7a\alpha$ nepetalactone as their key compounds. These species also contained isomers $4a\alpha, 7\beta, 7a\beta$ nepetalactone, $4a\alpha,7\beta,7a\alpha$ -nepetalactone, 4αdihydronepetalactone, 4β-dihydronepetalactone and 5,9-dihydronepetalactone along with first four isomers but their percentage was very low as compare to principal compounds. Isomer **4aα,7α,7aα-nepetalactone (1)** has found as major ingredient in species like *N. assurgens, N. cadmea, N. caesarea, N. cephalotes, N. crassifolia, N. x faassenii, N. govaniana, N. mirzayanni, N. persica, N. racemosa* and *N. teydea* (see table no. 1).

Further isomer $4a\alpha$, 7α , $7a\beta$ -nepetalactone (2) found as key constituent in species like N. argolica, N. atlantica, N. cataria, N. coerulea, N. granatensis, N. meyeri, N. nuda ssp nuda, N. nuda ssp. albiflora, N. nepetella, N. parnassica, N. racemosa, N. rtanjensis, N. septemcrenata, N. transcaucasica and N. tuberosa (see table no. 1). Furthermore, N. bornmuelleri, N. eremophila, N. persica, N. racemosa, N. saccharata and N. sintenissii have $4a\beta$, 7α , $7a\beta$ -nepetalactone (3); N. argolica, N. grandiflora and N. kotschyi contain $4a\beta,7\alpha,7a\alpha$ -nepetalactone (4); N. crassifolia have $4a\alpha,7\beta,7a\beta$ -nepetalactone (5); N. betonicifolia, N. crassifolia and N. nuda have 4aα,7β,7aα-nepetalactone (6); N. angustifoliates, N. cataria, N. mahanensis and N. nepetella have nepetalactone (7); N. mussini contain epinepetalactone (8): N. tuberosa subsp. tuberose have 5,9-dehydronepetalacone (9); and N. cataria and N. elliptica have (7R)-trans, trans-nepetalactone (10) as their main compound (see table no. 1).



Group 2:

This group consists of species like *N*. *septemcrenata* which contain **1,2 benzene-**

dicarboxylic acid dibutyl ester (11); *N. curviflora, N. daenensis, N. fissa, N. nuda* L. subsp. *albiflora* and *N. oxyodonta* contain (**trans**

 β)-carvophyllene (12); N. Cilicia, N. or betonicifolia and N. nuda L. ssp. nuda have β caryophyllene oxide (13); N. baytopii, N_{\cdot} binaludensis, N. congesta, N. Crispa, N_{\cdot} denudate, N. discolor, N. fissa, N. flavida, N. haussknechtii. gloeocephala, N_{\cdot} N. heliotropifolia, N. involucrate, N. ispahanica, N. italica, N. menthoides, N. pannonica, N. pogonosperma, N. rivularis, N. royleana, N. schiraziana and N. sulfuriflora which contain 1.8cineole (14); N. cataria have α -citral (15); N. citnodora have β -citronellol (16);⁴¹ N. elliptica have β -elemene (17); *N. raphanorhiza* have (**Z**)**β-farnesene (18)**; *N. laevigata* and *N. ucraininca* have germacrene-D (19); N. pungens have geranyl acetate (20); N. deflersiana and N. nuda L. ssp albiflora have hexadecanoic acid (21); N. clarkei and N. leucophvlla have iridodial Bmonoenol acetate diastereomers and iridodial **β-monoenol acetate (22)**; *N. erecta* and *N.* govaniana have isoiridomyrmecin (23); N. clarkei have kaur-16-ene (24); N. cilicia have betonicifolia limonene (25); N. and N_{\cdot} satureioides have linalool (26); N. sessilifolia have linalool acetate (27); *N. floccosa* have neral (28); N. discolar, N. glomerulosa and N. laxiflora have α-pinene (29); N. govaniana have pregeijerene (30); *N. pratti* have pulegone (31); N. bracteata, N. cilicica, N. depauperata, N. macrosiphon and N. sessilifolia have spathulenol (32); N. sintenissii have α -terpinolene (33), N. asterotrichus have terpinen-4-ol and (34) N. makuensis have viridiflorol (35) as their major constituent (see table no. 2).



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Chemical Composition of Various Species of Genus *Nepeta*

Species with nepetalactone as principal constituents:

N. assurgens essential oil taken out from the air dried aerial parts (collected during full flowering stage in June, 2011 from Lalehzar Mountain in the Baft area, Iran) has been analysed by using GC and GC-MS revealed the presence of twentytwo compounds which form 98.4% of the total oil composition. The oil contained seven monoterpenes (17.6%), two sesquiterpene (1.3%), ten oxygenated monoterpenes (78.0%) and three oxygenated sesquiterpenes (1.5%). 4aa,7a,7aanepetalactone (44.6%), 1,8-cineole (21.3%), βpinene (36) (5.3%), α-terpineol (37) (3.8%), (Z)β-ocimene (38) (3.7%), α-pinene (3.1%) and $4a\alpha$, 7α , $7a\beta$ -nepetalactone (2.8%) have been the core constituents of the total oil.42 Dabiri and Sefidkon (2003)⁴⁸ reported the presence of twenty one components in the essential oil (extracted with hydrodistillation for 6 hours from air-dried aerial parts) of Nepta crassifolia (collected during full flowering stage in month of September from north part of Tehran). The composition of oil has been analysed with GC and GC-MS. ßcaryophyllene (0.58%), $4a\beta$, 7α , $7a\beta$ -nepetalactone (0.64%), germacrene D (0.73%), piperitone (39) linalyl acetate (40) (2.6%) (0.80%).and $4a\alpha$, 7α , $7a\alpha$ -nepetalactone (92.6%) have been found to be the major components present in the

oil. Mahboubi *et al.* $(2011)^{51}$ analysed the essential oil of Nepeta persica (collected from Central Province Novazen, Arak, Iran at flowering stage in July 2009) with GC and GC-MS and investigated fourteen compounds which form 97.3% of total oil composition. The oil has been isolated from air-dried aerial parts with distillation Clevenger-type steam (using 6 apparatus) for hours. 4aα,7α,7aαnepetalactone (80.0%), spiro [5,6]dodecane (41) (14.2%), β-bourbonene (42) (1.5%), 2-methoxy-4-methylphenol (43) (0.8%) and caryophyllene oxide (0.7%) have been the principal compounds present in the oil. Adiguzel *et al.* $(2009)^{56}$ examined the essential of Nepeta cataria (the plants were collected from Olur at height of 800m during flowering stage in August, 2004) extracted with methanol using Soxhlet and Clevenger-type apparatus from air dried plant material. Oil has been subjected to GC/MS analysis and 22 compounds have been identified which form the 93.00% of oil. $4a\alpha, 7\alpha, 7a\beta$ total nepetalactone (70.4%), $4a\alpha, 7\alpha, 7a\alpha$ nepetalactone (6.0%), $4a\beta$, 7α , $7a\beta$ -nepetalactone (2.5%), thymol (44) (2.3%) and pulegone (1.8%) have been the principal compounds present in the oil.

Zenasni *et al.* (2008)⁵⁵ examined the chemical composition of four *Nepeta* species collected from Moroccan regions viz. *Nepeta atlantica* (collected from Boumia-Tounfit), *Nepeta cataria* (collected from Kenitra region), *Nepeta*

granatensis and Nepeta tuberosa (collected from Iran region). The oil has been isolated from whole plant material with steam distillation and the distillate so obtained has been further extracted with ethyl acetate and analysed with GC-MS. $4a\alpha, 7\alpha, 7a\beta$ -nepetalactone has been the key compound in the four Nepeta species. Major compounds present in four Nepeta species have been given in table no. 3. Essential oil obtained (with hydrodistillation for 3 hours) from the air dried aerial part of Nepeta parnassica (wild population) collected during vegetative stage in June and flowering stage in September 2000 (from Mt Parnassos at an elevation of 1600 m) constitute about 94.8% and 98.7% of the total oil composition respectively. The oil composition has been analysed with GC-MS. Fifty five compounds have been identified from the extracted oil of these two stages. The oil extracted during vegetative stage have $4a\alpha, 7\alpha, 7a\beta$ nepetalactone (22.0%), 1, 8-cineole (21.1%), αpinene (9.5%) and $4a\alpha$, 7 β , 7 $\alpha\beta$ -nepetalactone (7.9%) as the main components, while **1,8-cineole** (34.6%), $4a\alpha$, 7α , $7a\alpha$ -nepetalactone (17.3%), α pinene (11.4%), $4a\alpha$, 7α , $7a\beta$ -nepetalactone (8.9%) and $4a\alpha$, 7 β , 7 $\alpha\beta$ -nepetalactone (2.0%) have been the key component of oil obtained from flowering stage.⁴⁰ Grbic et al. (2008)⁶⁴ analysed the essential oil (isolated by hydro-distillation with Clevenger-type apparatus for 2 hours) of Nepeta rtanjensis obtained from air-dried aerial parts with GC-MS. The plants have been collected from experimental fields of Institute for Biological Research Sinisa Stankovic, Belgrade, Serbia during pre-flowering stage. $4a\alpha,7\alpha,7a\beta$ nepetalactone (79.89%) has been the key compound and $4a\beta$, 7α , $7a\beta$ -nepetalactone (6.3%), α -pinene (3.3%), δ -cadinene (52) (2.1%), germacrene D (1.8%) have been the other major compounds present in the oil. Thirty nine compounds have been reported from the essential oil (isolated from fresh aerial parts with hydro distillation for 3 hours) of Nepeta septemcrenata

(collected during full flowering stage in March, 2006 from Kathrine, Egypt) when analysed with GC-MS. These form 98.3% of the total oil. The oil mainly comprises of $4a\alpha$, 7α , $7a\beta$ -neptalactone (24.2%), 4aα,7α,7aα-nepetalactone (19.9%), 1,8cineole (8.5%), elemol (53) (13.8%), linalool (5.6%), while β -bisbolene, terpine-4-ol and α terpinol have been present in low concentration.⁶⁵ Dabiri and Sefidkon (2003)⁸ collected aerial parts of Nepeta racemosa at flowering stage (from central parts of Iran in September 2000). The oil has been extracted from air-dried aerial parts with hydro-distillation using Clevenger-type apparatus for 6 hours. The oil has been analysed with GC and GC-MS and twenty two compounds have been recognized which form 99.3% of the total oil composition. 1,8-cineole (9%), 4aα.7α.7aαnepetalactone (24.4%), 4α , 7α , (25.6%) and $4a\beta.7\alpha.7a\beta$ -nepetalactone (33.6%)have been the key compounds present in the essential oil. The volatile essential oil extracted (with hydrodistillation for 3 hours) from air-dried aerial parts of Nepeta sintenisii (collected from Charat, Iran in the month of june, 2002 at an elevation of 2250 m) showed the presence of forty constituents when analysed with GC-MS. The oil mainly comprises of monoterpenes (27.7%) and sesquiterpenes (43.6%), while 4aβ,7α,7aβ-netalactone (23.4%), elemol (16.1%), E- β -farnesene (9.5%), 1.8-cineole (8.2%), germacrene-D (3.5%), β -bisabolene (54) and cis-sabinene hydrate (55) (6.5%) (4.2%)have been the major compounds present in the oil.⁷ Ricci et al. (2010)⁷⁹ with the help of GC- ^{13}C and NMR recognized FID, GC-MS trans, trans-nepetalactone (50.38%), trans, cisnepetalactone (56) (21.74%),cis.transnepetalactone (57) (6.66%) and nepetalactol (58) (0.49%) as key component in the essential oil (isolated with hydrodistillation for 4 hrs and extracted with dichloromethane) of Nepeta cataria leaves (collected from Serra Azul Street, 308 Piracicaba, SP, Brazil).







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Species with other than nepetalactone as principal constituents:

GC-MS evaluation of essential oil of Nepeta binaludensis collected from cultivated fields revealed the presence of twenty-three components

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which form 99.4% of the total oil. The oil has been extracted from dried aerial parts with hydrodistillation procedure using Clevenger-type apparatus for 3 hours. 1,8-cineole (63%) has been the major constituent present in the oil. $4a\alpha$, 7α , $7a\alpha$ -nepetalactone (19%), alcohol fenchyl (59) (5.3%), β-pinene (4.5%), α-terpineol (2.2%) and p-cymene-8-ol (60) (1.4%) have been the other key compounds found in the oil.92 Sonboli et al. (2004)⁹⁵ collected aerial parts of Nepeta crispa in July, 2003 at flowering stage from Alvand Mountains, Hamadan, Iran. The oil was isolated from air-dried aerial parts with hydrodistillation using Clevenger-type apparatus for 3 hours. The analysis of oil with GC and GC-MS revealed the presence of twenty three compounds which form 99.8 % of the total oil composition. The oil contained sesquiterpene hydrocarbons, hydrocarbons and oxygenated monoterpene monoterpenes in 2.2%, 13.7% and 83.9% concentration respectively, whereas 1,8-cineol (47.9%), $4a\alpha$, 7α , $7a\beta$ - nepetalactone (20.3%), β pinene (6.9%), α -terpineol (4.8%), 4-terpineol (2.8%) and α -pinene (2.5%) have been the major compounds reported in the oil. The examination of essential oils of Nepeta Crispa (collected from Alvand Mountains, province of Hamadan) and Nepeta menthoides (collected from Sabalan Mountains, province of Ardabil) obtained from the leaves and flowers (with hydro-distillation using Clevenger-type apparatus for 3 hours) showed the presence of 89.5% monoterpenoids, 0.9% sesquiterpenoids, 0.6% phenyl propanes in former species, while the latter species contained 87.2% monoterpenoids, 0.1% sesquiterpenoids, 0.9% phenyl propanes. The composition of the oil has been analysed with GC and GC-MS. 1,8cineole has been the major constituent present in both the species. **1,8-cineole** (71.0%), β -pinene (5.0%), α -terpineol (4.1%), δ -terpineol (61)(2.8%) and 4-terpineol (2.3%) have been the major components reported in the essential oil of Crispa, whereas 1,8-cineole N. (41.1%), dihydromyrcen-1-ol (62) (9.2%), 4-terpineol (7.1%), geranyl acetate (6.1%), α -terpineol (5.7%), β-pinene (5.6%) and sabinene (63) (2.4%) have been the key components found in the

essential oil of N. menthoides. The study of essential oil revealed that these species have no traces of nepetalactone. ⁹⁴ Essential oil obtained from the air-dried aerial parts of Nepeta pogonosperma (collected in May, 2008 from its wild locality in Qazvin provience, Iran) by hydrodistillation using Clevenger-type apparatus has been analysed with GC-MS by Ali et al. (2012).¹⁰³ Forty-one compounds have been characterised which represent 97.5% of the total oil. β-pinene (3.5%), linalool (4.5%), terpinen-4ol (4.8%), (E)-α-bisabolene (64) (5.4%), α- $4a\alpha,7\alpha,7a\alpha$ -nepetalactone terpineol (5.4%), (14.5%) and **1,8-cineole (31.2%)** have been the prime constituents present in the oil.

Sefidkon et al. (2002)⁸⁵ evaluated the essential oil with GC and GC-MS taken out from air-dried aerial parts of Nepeta fissa C.A. Mey (collected from Tehran Province in Jun, 2000 at full flowering season) with hydro-distillation using Clevenger-type apparatus for 4 hours. Forty two compounds forming 99.3% of the total oil have been characterised composition and spathulenol (4.1%), bicyclogermacrene (65) (4,9%),α-pinene (5.8%), β-pinene (6.0%), valencene (66) (6.6%), *γ*-muurolene (67) (7.9%), caryophyllene oxide (12.3%)and βcarvophyllene (17.4%) have been the major compounds. Sajjadi and Eskandari (2005)⁸⁷ studied the essential oil with GC-MS extracted (by hydrodistillation for 3 hours) from air-dried aerial parts of Nepeta oxyodonta. The plant material has been collected in June from Charmahal and Bakhtiari province, Iran from height of 2700m. Total fifty-eight compounds have been characterised and δ -candinene (2.8%), cineole α-pinene (3.2%),1.8-(3.3%),caryophyllene oxide (5.3%), T-cadinol (5.6%), germacrene-D-4-ol (68) (6.8%), α-cadinol (69) germacrene-D (7.4%), β -bourbonene (7.3%), (8.1%). spathulenol (8.5%) and **(E)**caryophyllene (12.6%) have been the principal compounds. Essential oil obtained from the young leaves (solvent extraction, percolation) of Nepeta cataria (collected in July 2006 from medicinal plant garden of Tehran University, Iran) has been analysed using GC and GC-MS by Saeidnia et al.

(2008).¹⁰⁵ Four compounds have been identified which represent 97.53% of the total oil. The oil has been rich in monoterpenes, while α -citral (51.95%), nerol (70) (32.24%), β-citronellol (9.03%) and geraniol (71) (4.31%) have been the major components of the oil. No isomer of nepetalactone has been found in the oil. The evaluation of essential oils of Nepeta elliptica and Nepeta laevigata (collected from on June 2009 from high Himalayas of J & K, India) isolated from air-dried aerial parts (inflorescences) with hydro-distillation (using Clevenger-type apparatus for 3 hours) showed the presence nineteen and twenty-four compounds forming 83.4% and 86.7% of the total oil composition respectively. The chemical composition of both oils have been analysed with capillary GC-FID and GC-MS. N. elliptica essential oil contained sesquiterpene hydrocarbon, 68.8% 12.2% oxygenated sesquiterpene, while α -terpineol (1.7%), spathulenol (2.6%), zingiberene (72)(2.8%), elemol (3.5%), γ -cadinene (3.3%), β bisabolene (3.2%), germacrene D (3.6%), β caryophyllene (5.5%), α -humulene (73) (11.8%), bicyclogermacrene (13.1%) and β -elemene (23.4%) have been the major components. Further, the essential oil of N. laevigata contained 20.7% oxygenated monoterpenes, 40.9% sesquiterpene hydrocarbon, 25.1% oxygenated sesquiterpene, whereas $4a\alpha.7\alpha.7a\alpha$ -nepetalactone (2.0%), caryophyllene oxide (3.2%), α -humulene spathulenol β-bourbonene (3.5%),(3.9%),(4.5%), α -bisabolol (74) (5.3%), α -bisabolol oxide B (75) (12.4%), β-caryophyllene (10.8%), germacrene D (19.4%) and **B-Citronellol** (16.5%) have been the prime components.¹⁰⁶ GC-MS examination of essential oil of Nepeta raphanorhiza collected in April, 2010 from Pulwama region of Kashmir Valley showed the presence of sixteen components which form 97.5% of the total oil. The oil has been extracted from the fresh aerial plant parts with hydrodistillation procedure using Clevenger-type apparatus for 3 hours. The oil contained 65.3% sesquiterpene hydrocarbons and 19.5% of monoterpenes hydrocarbons. Sesquiterpenes mainly composed of α -humulene (2.0%), α - bisbolene (9.4%), β -caryophyllene (12.7%) and **(Z)-\beta-farnesene (49.2%)**, whereas monoterpene hydrocarbons have δ -3-carene **(76)** (12.3%) as main compound. Sabinene (2.5%), caryophyllene oxide (3.4%) and germacrene-D-4-ol (5.8%) has been also found in the oil. Literature revealed that most of the Himalayan *Nepeta* species from Jammu and Kashmir showed either no or very low content of nepetalactone in their essential oil.³⁷

Farjam (2012)¹⁰⁸ investigated the essential oil of Nepeta pungens collected from Sepidan mountain, Fars, Iran in August, 2011. The oil has been extracted from air-dried leaves using Clevenger-type apparatus for 4 hours. GC and GC-MS analysis of the oil revealed the presence of forty nine compounds which constitute 97.2% of the total oil composition. Nervlacetate (77) (2.5%), β -sesquiphellandrene (78) (2.8%), β ocimene (3.9), sabinene (3.9%), spathulanol (4.2%), citronellal (4.9%), bornylacetate (79) (5.3 %), eucalyptol (1,8-Cineole) (5.8 %), limonene (12.0 %) and geranyl acetate (17.0 %) have been the key compounds reported in the oil. Airdried aerial parts of Nepeta deflersiana (collected in May, 2007 from the vicinity of Sana'a-Yeman) have been subjected to hydro-distillation using Clevenger-type apparatus for 3 hours. The obtained essential oil has been analysed by using GC and GC-MS which showed the presence of fifty-one compounds mainly oxygenated monoterpenes oxygenated (31.4%)and sesquiterpenes (28.2%). Hexadecanoic acid (8%), caryophyllene oxide (6.4%), 2-methoxy-pcresol (5.6%), camphor (80) and eugenol (81) (4.7% each) have been the main constituents reported in the total oil.¹⁰⁹ Bisht et al. (2010)⁸⁰ examined chemical composition of essential oil isolated with steam distillation from fresh flowering aerial parts of six Himalayan Nepeta species viz. Nepeta clarkei Hook. F. (collected from Malari, Chamoli at height of 2800m), Nepeta discolor Royle ex Benth. (collected from Malari, Chamoli at height of 2800m), Nepeta elliptica Royle ex Benth. (collected from Clips, Nainital at height of 2700m), Nepeta erecta Benth. (collected from Hemkund at height of

3250 m), Nepeta govaniana Benth. (collected from Bhundiar, Chamoli at height of 2600m) and Nepeta leucophylla Benth (collected from Nainital at height of 2400 m). The oils have been analysed by using GC and GC-MS technique. The major compounds present in six Himalayan Nepeta species have been given in table no. 4. GC-MS and GC-FID analysis of oil extracted with Clevenger-type apparatus for 3 hours from the fresh plant material (aerial parts) of Nepeta govaniana (collected in July, 2010 from Gulmarg region of Kashmir Valley) showed the presence of seventeen components which form 87.3% of the total oil composition. The oil contains hydrocarbons sesquiterpene and oxygenated sesquiterpenes which form 80.5% and 6.5% composition, respectively. β -bourbonene (3.6%), β -caryophyllene (6.1%), germacrene D (9.4%) and pregeijerene (56.9%) have been the major constituents of sesquiterpene hydrocarbon present in the oil, whereas oxygenated sesquiterpenes have torreyol (85) (5.1%) as principal compound and other minor components include elemol, germacrene-D-4-ol, α -cadinol and α -eudesmol.¹¹⁷ Thirteen components have been identified (which forms 97.55% of the total oil composition) from the essential oil of Nepeta cilicia (collected from Basconus Mountain at an altitude 1450 m in month of June) when analysed by GC-MS. The oil has been isolated from air-dried flowering parts and leaves using hydro-distillation method with Clevenger-type apparatus for 3 hours. Limonene (44.75%), β-pinene (13.6%), βcaryophyllene (11.2%), caryophyllene oxide (9%), germacrene D (5.2%) and α -pinene (3.5%) have been the prime components of the oil. Less than 9% of the total oil composition consists of other compounds.¹¹² The examination of essential oil of Nepeta depauperata using GC-MS showed the presence of thirty three compounds which form 82.52% of the total oil composition. The oil has been obtained from the air-dried aerial parts with hydro-distillation using Clevenger-type apparatus for 3 hours. Plants have been collected from northern slopes of Khabr Mountain, Kerman Province, Iran at flowering season from height of 3500m in Jun, 2002. β-eudesmol (86) (2.16%), δcadinene (2.80%), 1,8-cineole (3.97%), α candinol (5.41%), caryophyllene oxide (10.27%), β -caryophyllene (12.93%) and **spathulenol** (**31.84%**) have been the principal constituent present in the oil.²⁷

Ghannadi *et al* $(2003)^{121}$ isolated the essential oil of Nepeta macrosiphon Boiss from air-dried flowering aerial parts by Likens-Nickerson's simultaneous distillation-extraction procedure. The plant material has been collected from northern slopes of Dalakhani mountain, Songhur, Iran at height of 2300 m. GC-MS analysis of the oil revealed the presence of forty-five compounds which constitute 95.1% of the total oil composition. Torreyol (2.0%), myrtenol (87) (2.0%), δ-cadinene (2.3%), germacrene A (2.4%), α -cadinene (88) (3.0%), citronellyl acetate (89) (3.8%), γ -eudesmol (90) (3.9%), linalool (4.1%), verbenone (91) (4.4%), aromadendrene (92) $(4.8\%), \alpha$ -cadinol (4.9%),bicyclogermacrene (5.7%), α -muurolene (93) (6.0%), caryophyllene oxide (8.1%), germacrene D (9.2%) and spathulenol (14.1%) have been the prime compounds present in the oil. GC-MS analysis of essential oil of Nepeta sintenisii taken out from Clevenger-type the air-dried leaves (with apparatus for 6 hours) showed the presence of twenty one components which form 97.82% of the total oil. Plants have been collected in Jun, 2010 from Darkesh protected area, Bojnourd, North Khorassan Province Iran at the flowering Germacrene-D stage. (2.27%), β-bisabolene (3.65%), nepetalactone (4.93%), E-β-farnesene (22.67%) and α -terpinolene (47.86) have been the main compounds present in the oil.¹²³ Fresh aerial parts of Nepeta septemcrenata [collected from the wadi gebal in Sant cathrin (Sinai), Egypt on August, 2002] have been subjected to hydrodistillation using Egptian Pharmacopoeia apparatus and analysed by GC-MS. The obtained essential oil have sixteen key compounds [ten oxygenated (70.67%) compounds and six hydrocarbons (24.29%)] forming 94.96% of total oil composition. 1,2 benzene-dicarboxylic acid dibutyl ester (33.16%), 4-methyl-2,6-ditertbutyl phenol (94) (23.30%), thujone (95) (5.45%), hexadecanoic acid (3.64%) and 2-decyn-1-ol (96)

(3.25%) have been the principal oxygenated compounds, whereas 4-nonyne (97) (9.00%), n-heneicosane (98) (6.69%), 3-octyne (99) (5.14%)

and 2-methyl-pentadecane (100) (2.36%) have been the main hydrocarbons present in the essential oil.⁸¹



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CONCLUSION

The composition of the essential oil has been varying according the region, soil type and environmental condition from where the plant species have been collected.^{126,127} It has been found from the literature that the most of species growing in the Himalayas region have been contained compounds other than nepetalactone as major constituents in their essential oils as compare to the species growing in other part of the world (Iran, Tehran, Serbia, Egypt, Turkey, Brazil, USA etc.), which have both nepetalactone along with its derivatives and other than nepetalactone compounds as the major ingredient

of their essential oils. The essential oil isolated from different plant parts viz. stem, leaves, flowers (fresh or dry) collected at different age and vegetative cycle stage have no major effect on the composition of the oil. It has been also found from the literature that the oil of different species of *Nepeta* have been mostly isolated from aerial parts mainly collected during full flowering stage with hydrodistillation using Clevenger type apparatus gives good results. For the sustainable growth and development, a gradual shift from the use of synthetic drugs and agrochemicals to the natural ones have been witnessed as the latter have proven to be non-toxic, cheap and easily available. Active ingredients isolated from genus

Nepeta has been reported to show wide array of biological activity in medicinal and agriculture field. Depending upon the multiple uses of secondary metabolites obtained from genus *Nepeta*, their structure-activity relationship and activity screening goes unabated. The present review would be supportive in the enhancement of today's research in the development of new biologically potent compounds derived from

plants which would find many applications in medicinal and agricultural fields.

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Table 1: Species which	contained nepetalactone	e as principal constituents
	1	1 1

C. N.	S	Principal Compounds (%) A B C D E F G H I J K L M N O P R																
Sr. No.	Species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Ref.
1	N. assurgens	44.6	2.8						21.3						3.1/	3.8	22/	42
	(aerial parts)														5.3	/-	98.4	
2	N. cadmea*																ľ	
	i (herbal parts)	74.96															84/	43
																	94	
	ii (herbal parts)	44.51		-													-	ļ
3	N. caesarea*																	
	1	91.2-																44
		95.3																45
4	ll No control of control	92-95															ļ	45
4	N. cephalotes"	25.1							114						/		10/	16
	1	33.1							11.4						18.2		10/ 78 5	40
															10.2		8	
	ii (aerial narts)	90.1													_/		6	47
	n (act iai parts)	2011													7.5		'	.,
5	N. crassifolia	92.6													,		21/	48
	(aerial parts)																99.49	
6	N. x faassenii	67.8				-/			6.6				4.8		2.7		109	49
	(dry leaves)					2.3									/-		/95.9	
7	N. govaniana	25.9									17.5		20.5					50
											/-							
8	N. mirzayanni	61.0								-/							22	47
	(aerial parts)									7.8							/-	
9	N. persica	80															14/	51
10	(aerial parts)	(1.0	7.4	1.7													97.3	50
10	N. racemosa	64.9	7.4	1./													ļ′	52
11	N. teyaea **	00 54																53
	a) leaves (before flowering)	89.54													-	-		
	by leaves and nowers	-													7 87	12.04		
12	N. argolica ssp		68-	1											1.07	,	62	54
	Argolica		94.5														/-	

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Sr. No	Species	Principal Compounds (%) A B C D E F G H I J K L M N O P R																
Sr. 190.	Species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Ref.
13	N. atlantica (whole plant)		71.4							8.2/-		-/ 2.5					26/ 99.9	55
14	<i>N. cataria*</i> i (whole plant)	-	77.4			-			-	-				4.1 /-			26/ 99.7	55
	ii (leaves)	6.0	70.4			-/ 2.5			-	-				-			22/ 93.0	56
	iii	11.9	28.8			-			13.5 /-	5.7 /-				-			-	57
15	N. coerulea	11.9	21.5	3.7	19.3													58
16	N. granatensis (whole plant)		39.4						24.0						6.3 /2.3		26/ 98.9	55
17	<i>N. meyeri</i> (aerial parts)	8.83	83.4														14/ -	59
18	N. nuda * i. ssp nuda** a) leaves oil	-	24.7						16.7	- /16.3								60
	b) verticillaster oil ii. <i>ssp. albiflora</i>	- 37.6	75.7 37.6						-	- -/ 4.4								61
19	N. nepetella ssp Aragonensis	3.5	57.7															58
20	<i>N. parnassica</i> ** a) vegetative stage	1.5	22.0			7.9			21.1	- /4.8					9.5 /3.5	5.0 /-	35/ 94.8	40
	b) flowering stage	17.3	8.9			2.0			34.6	-					11.4 /6.4	4.3 /-	46/ 98.7	
21	N. racemosa		31.5- 91.6															62
22	N. rtanjensis* i (aerial parts)		86.4	6.3									-		1.4 /-		19/ 95	63
	ii (aerial parts)		79.9	-									1.8		3.3 /-		8/ 96.2	64

S- No	Smanian							Pri	ncipal (Compou	nds (%)							
Sr. 10.	Species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Ref.
23	<i>N</i> .	19.9	24.2						8.5		-			-/		2.5	39/	65
	septemcrenata										/13.8			5.6		/2.7	98.3	
	(aerial parts)	•																
24	N. transcaucasica	28	39										15				27/	66
25	(aerial parts)	-	7(0						1.2						-		97.69	<i></i>
25	N. tuberose		76.8						1.2								26/	55
26	(whole plant) N bornmuallari			64.0					7.1								28/	47
20	(aerial narts)			04.0					/.1								20/	4/
27	N. eremonhila			73.3					13.1								26/	67
,	(aerial parts)																-	
28	N. persica**																	68
	a) flower		33.0	58.5											3.6			
															/-			
	b) leaf		28.3	62.											-			
			24.0	3											16			
	c) stem		24.9	66.2											4.0			
	d) root		7.6	27.1											40 4			
	u) 1001		7.0	27.1											/-			
29	N. racemosa	24.4	25.6	33.6					9								24/	8
	(aerial parts)																99.3	
30	N. saccharata			66.9									12.9				18/	69
	(aerial parts)																98.2	
31	N. sintenissii			23.4					8.2		-	9.5	3.5			2.3	40/	7
	(aerial parts)		1.0.0								/16.1	/-				/2.5	96.5	
32	N. argolica	26.5	12.9	14.5	29.4													70
33	N. grandiflora	_	2.4		41.0				2.6						-		11/	71
34	N. Kotschyl				92.0				2.6								07.7	12
35	(aeriai parts)	16.3	77	9.6		27.2			9.0								91.1	73
55	IV. Crassijona	10.5	1.1	9.0		/_			9.0									15
36	N. betonicifolia					-/			3.2				6.0				33/	69
	(aerial parts)					42											97.9	
37	N. crassifolia	5.9				-/												57
						81.1												

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Sr.	Stranian							Pri	ncipal (Compou	inds (%)							
No.	Species	Α	В	С	D	Е	F	G	Η	Ι	J	K	L	Μ	Ν	0	Р	Ref.
38	N. nuda					-/ 18.10					-/ 14.38		15.68					74
39	N. angustifaliathe						75.37 /-										29/ -	75
40	<i>N. cataria*</i> i						>77 /-		-	-	-	-		-	-	-	-	41
	ii (aerial parts)						27.5 /-		10.8	5.5 /-	2.5 /-	2.6 /-	9.2	-/ 3.6	2.5 /-	5.3 /-	47/ 91.2	76
41	<i>N. mahanensis</i> (aerial parts)						37.6 /-		27.2				6.5				18/ -	67
42	<i>N. nepetella</i> (leaves and flowers)						76.5 /-											77
43	N. mussini						-/ 70											41
44	<i>N. tuberosa subsp. tuberose</i> (inflorescences)							69 /-		5.0 /-								78
45	N. cataria (leaves)							-/ 50.38									4/ 79.27	79
46	<i>N. elliptica</i> (aerial parts)							-/ 83.4										80

*sub heading i, ii, iii etc. represent the essential oil composition of same plant (having same compound as major component of the oil) investigated from different localities by same or different worker.

** sub heading a), b), c) etc. represent the essential oil composition of different parts of same plant investigated by same worker.

*** species in the above table have been alphabetically enlisted according to the major constituent due to which repetition in the name of species may be observed. (because same species from different locality may have different major constituents)

A: $4a\alpha,7\alpha,7a\alpha$ -nepetalactone; B: $4a\alpha,7\alpha,7a\beta$ -nepetalactone; C: $4a\beta,7\alpha,7a\alpha$ -nepetalactone; D: $4a\beta,7\alpha,7a\beta$ -nepetalactone; E: $4a\alpha,7\beta,7a\beta$ -nepetalactone; F: nepetalactone; F: nepetalactone; F: nepetalactone; F: nepetalactone; C: $4a\beta,7\alpha,7a\beta$ -nepetalactone; C: $4a\beta,7\alpha,7a\beta$ -nepe

Species with sr. no. (1-11) have $4a\alpha,7\alpha,7a\alpha$ -nepetalactone, sr. no. (12-25) have $4a\alpha,7\alpha,7a\beta$ -nepetalactone, sr. no. (26-31) have $4a\beta,7\alpha,7a\alpha$ -nepetalactone, sr. no. (32-34) have $4a\beta,7\alpha,7a\beta$ -nepetalactone, sr. no. (35) have $4a\alpha,7\beta,7a\beta$ -nepetalactone, sr. no. (36-38) have $4a\alpha,7\beta,7a\alpha$ -nepetalactone, sr. no. (39-42) have nepetalactone, sr. no. (43) have epinepetalactone, sr. no. (44) have 5,9-dehydronepetalacone and sr. no. (45-46) have trans, trans-nepetalactone as the key constituent in their essential oil.

Key compounds (other than compounds given in table from A to O) present in different species given in above table:

1: *N. assurgens* [(Z)- β -ocimene (3.7%)]; 5: *N. crassifolia* [linalyl acetate (2.6%)]; 6: *N. x faassenii* [(Z)- β -ocimene (2.6%)]; 9: *N. persica* [Spiro[5.6]dodecane(14.2%)]; 10: *N. racemosa* [(Z)- β -ocimene (9.5%), (E)-nerolidol (8.8%)]; 11: *N. teydea* b) [camphor (5.94%), 1,8-cineole + limonene (14.67%)]; 13: *N. atlantica* [dihyderonepetalactone (3.1%)]; 14: *N. cataria* i [dihyderonepetalactone (5.0%), terpinene (4.2%)]; iii [citronelly acetate (5.2%)]; 16: *N. granatensis* [α -phellandrene (5.8%), ρ -cymene (3.8%), dihyderonepetalactone (2.8%)]; 20: *N. parnassica* a) vegetative stage [trans-verbenol (6.4%), verbenone (4.5%)], b) flowering stage [δ -cadinene (2.8%), ρ -mentha-1,5-diene-8-ol (2.6%)]; 23: *N. septemcrenata* [β -bisabolene (3.7%)]; 25: *N. tuberose* [dihyderonepetalactone (5.9%)]; 28: *N. persica* b) leaf [β -ocimene (3.6%)] d) [α -amorphene (5.3%), γ -cadinene (2.8%)]; 30: *N. saccharata* [sabinene (6.5%), trans-caryophyllene (3.3%)]; 31: *N. sintenissii* [cis-sabinene hydrate (6.5%), β -bisabolene (4.2%), β -sesquiphellandrene (2.8%)]; 36: *N. betonicifolia* [triplal (5.2%), 1-norbourbonanone (4.0%)]; 39: *N. angustifaliathe* [spiro(5,6) dodecane (3.47%), 2,4-dimethyl-1,3-pentadiene (2.7%)]; 40: *N. cataria* ii [spathulenol (2.8%)]; 44: *N. tuberosa subsp. tuberose* [geranyl acetate (17%)]; 45: *N. cataria* [trans-cis nepetalactone (21.74%), cis-trans nepetalactone (6.66%)].

C. N.	C								Prin	cipal C	ompoun	ds (%)							
Sr. No.	Species	Α	B	С	D	Ε	F	G	Н	I	J	K	L	Μ	Ν	0	Р	Q	Ref.
1	N. septemcrenata (fresh aerial	33.16																16/ 94.96	81
	parts)																		
2	N. curviflora*																		
	i		41.6 /			11.0												-	82
			9.5			/-													
	ii (aerial parts)		50.2/			5.3												35/	83
			6.4			/-												93.2	
3	N. daenensis		27.1/			_/					14.5							23	84
			2.8			11.4					/-							/-	
4	N. fissa		17.4/								5.8/		4.1/			2.7/		42/	85
	(aerial parts)		12.3								6.0		-			-		99.3	
5	N. nuda L.		23.9	6.4		-/					-/		7.35					24/	86
	subsp. <i>albiflora</i>		/-	/-		2.75					10.01		/-					93.46	
	(aerial parts)																		
6	N orvodonta		12.6	33		_/					3.2		8.5		_/	73		58/	87
U	(aerial narts)		/5 3	/_		74					5.2		/_		81	/_		96 1	07
7	N cilicia		15.7/	,		,							,		0.1	,		23/	88
,	(leaves &		40 7															96.6	00
	flowers)																	2010	
8	N. betonicifolia		_/										97					42/	89
	1		39.2										/-					90.7	•
9	N. nuda L. ssp.		5.4/										13.8						90
	nuda		21.8										/-						
10	N. baytopii		5.6	23.2		_/					3.1/		3.9/			5.3		46/	76
	(aerial parts)		/4.4	/-		4.5					3.8		2.8			/-		92.2	
11	N.																		
	binaludensis*																		
	i			42					-/		-/						-	-	91
				/-					4.0		3.0								
	ii (aerial parts)			63 /-					-		- /4.5						19.0 /-	23/ 99.4	92

Table 2: Species which contained constituents other than nepetalactone as principal constituents

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Sr. No	Spacios								Prin	cipal C	Compour	1ds (%)							
Sr. 10.	species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	М	Ν	0	Р	Q	Ref.
12	<i>N. congesta</i> (aerial parts)			29.9 /-		-/ 20.3							-/ 10.3						93
13	N. crispa* i (leaves & flowers) ii (aerial parts) iii (aerial parts)			71 /- 47.9 /- 62.8 /-							-/ 5.0 2.5 /- -						-/ 20.3 10.3 /-	12/ 91.0 23/ 99.8 28 /-	94 95 67
14	N. denudata			48.0 /-							-/ 4.6							21/ 85.7	46
15	<i>N. discolor</i> (fresh flowering aerial parts)		18.8/-	25.5 /-															80
16	<i>N. fissa</i> (aerial parts)		9.2/ 8.2	24.3 /-		3.1/ 5.4					-/ 3.2		3.0/ 3.2					49/ 92.5	76
17	N. flavida			38.9 /-					-/ 25.1									68/ 96.4	6
18	<i>N.</i> <i>gloeocephala</i> (aerial parts)			35.2 /-							7.1/ 21.8		-/ 7.8					29/ 99.98	96
19	<i>N.</i> <i>haussknechtii</i> (aerial parts)			36.7 /-														27/ 94.2	97
20	N. heliotropifolia		11.3/ 14.2	19.0 /-		5.1 /-							8.3 /-					25/	98
21	<i>N. involucrate</i> (aerial parts)			23.1 /-		-/ 15.1					-/ 12.2							48/ 97.2	99

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Sr. No	Species								Prin	cipal C	Compour	nds (%)							
Sr. 10.	Species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Q	Ref.
22	N. ispahanica*																		
	Ι			66														-	91
				/-															(7
	ii (aerial parts)			71.7														27	67
22	N italiaa			/-														/-	100
23				/-														/-	100
24	N. menthoides*																		
	i (leaves &			41.1			6.1				-/						-	27/	94
	flowers)			/-			/-				5.6							89.5	
	ii (aerial			33.8			-				-						23.2	29/	101
	flowering			/-													/-	97.6	
	parts)																		100
25	N. pannonica			28.9														60	102
26	(aerial parts)			/-					1		/			1	5.4		14.5	/-	102
26	<i>I</i> N.			31.2					-/		-/			-/	5.4		14.5	41/	103
	<i>pogonosperma</i> (aerial parts)			/-					4.3		5.5			4.0	/-		/-	97.5	
27	N. rivularis			38.5							_/		_/					22	67
_,	(aerial parts)			/-							10.7		14.8					/-	
28	N. royleana			75															50
				/-															
29	<i>N</i> .																		104
	schiraziana**																		
	a) stem		11.7	45.6		-/												14	
	1. 6		/-	/-		17.4												/-	
	b) flower		10.6	34.4		-/												14	
			/-	/-		15.8												/-	
	c) leaves		/11 7	30.5 /-		-												10	
30	N. sulfuriflora		/11./	61.5														28	100
20				/-														/-	100
31	N. cataria			_/	9.03													4/	105
	(young leaves)			51.95	/-													97.53	

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C. No	S-reading.								Prin	cipal C	Compour	nds (%)							
Sr. No.	Species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Q	Ref.
32	N. elliptica		5.5		-/	-/							2.6		3.2			19/	106
	(aerial parts)		/-		23.4	3.6							/-		/-			83.6	
33	<i>N</i> .		3.2			49.2							-/		9.4			16/	37
	raphanorhiza		/3.4			/-							2.5		/-			97.5	
	(aerial parts)																		
34	N. laevigata		10.8		16.5	-/							3.9		-/			24/	106
	(aerial parts)		/3.2		/-	19.4							/-		4.5			86.7	
35	N. ucraininca					-/							5.6		-/			41/	107
						39.7		-					/-		5.8	,		89.5	100
36	N. pungens			5.8	4.9		17.0		12				4.2			-/		49/	108
	(aerial parts)		,	/-	/-		/-		/-				/3.9			3.9		97.2	100
37	N. deflersiana		-/				-/		-/									53/	109
20	N / T		6.4				8.0		2.6			1			7.0			90.1	00
38	N. nuda L. ssp		-/				-/					-/			7.8				82
	albiflora*		1.3				10.1					1.2			/-				
	(collected from																		
20	Tannourine)					/		25.3			-				+	+			80
39	IV. Clurkel					-/		23.3											80
	flowering					15.0		/-											
	aerial narts)																		
40	N. leucophylla							25.4											80
10	(fresh							/-											00
	flowering							,											
	aerial parts)																		
41	N. erecta*																		
	i (fresh		-					_/										-	80
	flowering							66.7											
	aerial parts)																		
	ii (aerial parts)		-/					-/										34/	110
			9.6					70.6										94.6	
42	N. govaniana							_/				20.7							80
	(fresh							35.2				/-							
	flowering																		
	aerial parts)																		

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S. No	Smaataa								Princ	cipal Co	ompoun	ds (%)							
Sr. No.	Species	А	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Q	Ref.
43	N. clarkei***		-/ 14.1															20/ 95.3	111
44	<i>N. cilicia</i> (flowering parts and leaves)		11.2 /9.0			-/ 5.2			44.75/-		3.5 /13.6							13/ 97.55	112
45	<i>N. betonicifolia</i> (aerial parts)		-/ 9.2	20.8 /-					-/ 40.5									34/ 89.6	113
46	N. satureioides		6.6/ 6.4			3.4 /-			-/ 23.8									45/ 97.4	114
47	<i>N. sessilifolia</i> (aerial parts)								-/ 14.2	14.7 /-								33/ 97.4	97
48	N. floccosa									-/ 33.1									50
49	N. discolar										18.5 /12.6								50
50	N. glomerulosa										18.3 /-								115
51	N. laxiflora			11.8 /-							19.7 /-							57 /-	116
52	<i>N. govaniana*</i> i (aerial parts) ii (aerial parts)		6.1 /- -			-/ 9.4 -						56.9 /- 38.0 /-			-/ 3.6 -			17/ 87.3 >50 /-	117 118
53	N. pratti											-/ 45.21						15/ 76.8	119
54	<i>N. bracteata</i> (aerial parts)		11.2 /12.3										14.0 /-					28 /-	47
55	<i>N. cilicica</i> (aerial parts)						-/ 14.0						15.1 /-					75/ 96.8	120
56	<i>N. depauperata</i> (flowering aerial parts)		12.93/ 10.27	3.97 /-									31.84/-			5.41 /-		33/ 82.5	27

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Sr.	Spacing		Principal Compounds (%)																
No.	species	Α	В	С	D	Е	F	G	Н	Ι	J	K	L	Μ	Ν	0	Р	Q	Ref.
57	N. macrosiphon*																		
	i (flowering																		
	aerial parts)		_/			_/			_/				14.1			4.9		45/	121
	ii (aerial parts)		8.1			9.2			4.1				/-			/-		95.1	
			9.6			-			_/				28.8			13.6		73/	122
			/-						5.6				/-			/-		96.9	
58	N. sessilifolia												25.75/-					57/-	116
59	N. sintenissii					22.67/								47.86/-	3.65			21/	123
	(aerial parts)					2.27									/-			97.82	
60	N. asterotrichus			17.4					_/					-/				15	124
				/-					12.5					22.8				/-	
61	N. makuensis***												9.0/-					28/	125
																		92.9	

*sub heading i, ii, iii etc. represent the essential oil composition of same plant (having same compound as major component of the oil) investigated from different localities by same or different worker. ** sub heading a), b), c) etc. represent the essential oil composition of different parts of same plant investigated by same worker.

*** The major constituent of Species N. clarkei with sr. no. 43 and N. makuensis with sr. no. 61 have kaur-16-ene (36.6%) and viridiflorol (17.5%).

**** species in the above table have been alphabetically enlisted according to the major constituent due to which repetition in the name of species may be observed. (because same species from different locality may have different major constituents)

A: 1,2 benzene-dicarboxylic acid dibutyl ester; B: β -caryophyllene/ β -caryophyllene oxide; C: 1, 8-cineole (eucalyptol)/ α -citral; D: β -citronellol/ β -elemene; E: (E-Z)- β -farnesene/germacrene-D; F: geranyl acetate/hexadecanoic acid: G: iridodial β -monoenol acetate diastereomers and iridodial β -monoenol acetate/ isoiridomyrmecin; H: limonene/ linalool; I: linalool acetate/neral; J: α -pinene/ β -pinene; K: pregeijerene/ pulegone; L: spathulenol/sebinene; M: α -terpinolene/ terpinen-4-ol, N: (α/β)-bisabolene/ β -bourbonene; O: α -cadinol/ β -ocimene ; P: 4a α ,7 α ,7a α -nepetalactone/4a α ,7 α ,7a β -nepetalactone; Q: total compounds/representing % of total oil.

Species with sr. no. (1) have 1,2 benzene-dicarboxylic acid dibutyl ester, sr. no. (2-6) have β -caryophyllene, sr. no. (7-9) have β -caryophyllene oxide, sr. no. (10-30) have1, 8-cineole (eucalyptol), sr. no. (31) have α -citral, sr. no. (32) have β -elemene, sr. no. (33) have (E-Z)- β -farnesene, sr. no. (34-35) have germacrene-D, sr. no. (36) have geranyl acetate, sr. no. (37-38i) have hexadecanoic acid (38 ii have α -bisabolene), sr. no. (39-40) have iridodial β -monoenol acetate diastereomers and iridodial β -monoenol acetate, sr. no. (41-42) have isoiridomyrmecin, sr. no. (43) have kaur-16-ene, sr. no. (44) have limonene, sr. no. (45-46) have linalool, sr. no. (47) have linalool acetate, sr. no. (48) have neral, sr. no. (49-51) have α -pinene, sr. no. (52) have pregeijerene, sr. no. (53) have pulegone, sr. no. (54-58) have spathulenol, sr. no. (59) have α -terpinolene, sr. no. (60) have terpinen-4-ol, sr. no. (61) have viridiflorol as the principal constituent in their essential oil.

Key compounds (other than given in above table from A to P) present in different species:

3: *N. daenensis* [bicyclogermacrene (9.6%), bicycloelemene (8.1%), myrcene (5.6%)]; 4: *N. fissa* [γ-muurolene (7.9%), valencene (6.6%), bicyclogermacrene (4.9%), δ-cadinene (3.0%)]; 5: *N. nuda* L. subsp. *albiflora* [isopulegone (12.6%), cis-sabinol (10.1%), β-myrcene (2.95%), calarene (2.65%), α-campholene aldehydes (2.56%); 6: *N. oxyodonta* [germacrene-D-4-ol (6.8%), T-cadinol (5.6%), δ-cadinene (2.8%)]; 7: *N. Cilicia* [αcopaene (7.3%), βcubebene (6.6%)]; 8: *N. betonicifolia* [caryophyllenol-II (5.1%), humulene epoxide-II (4.7%), isocaryophyllene oxide (4.3%)]; 9: *N. nuda* L. ssp. *nuda* [allo-aromadendrene (9.0%)]; 10: *N. baytopii* [nepetalactone (12.8%), α-terpineol (3.1%), β-Gurjunene (2.5%)]; 11: *N. binaludensis* i [nepetalactone (25%), α-terpineol (4.0%)], ii [alcohol fenchyl (5.3%)]; 13: *N. Crispa* i [α-terpineol (4.1%), δ- terpineol (2.8%)]; 16: *N. fissa* [nepetalactone (17.6%), α-terpineol (2.8%)]; 18: *N. gloeocephala* [(E)-β-ocimene (7.1%), (Z)-β-ocimene (6.9%)]; 19: *N. haussknechtii* [elemol (11.4%)]; 20: *N. heliotropifolia* [myrtenol (5.9%), *trans*-pinocarveol (4.2%), α-terpineol (5.7%)]; 24: *N. menthoides* i [dihydromyrecen-1-ol (9.2%), 4-terpineol (3.2.4%), geraniol (4.31%)]; 32: *N. elliptica* [bicyclogermacrene (13.1%), α-binsabolol (5.3%), γ-cadinene (3.3%), gibeerne (2.8%)]; 31: *N. cataria* [nerol (3.2.4%), geraniol (4.31%)]; 32: *N. elliptica* [bicyclogermacrene (13.1%), α-binsabolol (5.3%), γ-cadinene (3.3%), gibeerne (2.8%)]; 33: *N. cataria* [nerol (3.2.4%), geraniol (4.31%)]; 32: *N. elliptica* [bicyclogermacrene (13.1%), α-binsabolol (5.3%), α-terpineol (5.3%)]; 35: *N. ucrainica* [abisabolol oxide B (12.4%), α-bisabolol (5.3%)]; 35: *N. ucrainica* [palmitic a-bisabolol oxide B (12.4%), α-bisabolol (5.3%)], γ-cadinene (3.5%)]; 35: *N. ucrainica* [palmitic a-bisabolol oxide B (12.4%), α-bisabolol (5.3%)]; 35: *N. ucrainica* [palmitic a-bisabolol oxide B (12.4%), α-bisabolol (5.3%)]; 35: *N. ucrainica* [palmitic a-bisabolol oxide B (12.4%), α-bisabolol (5.3%)]; 35: *N. u*

(6.9%)]; 39: *N. clarkei* [β -sesquiphellandrene (22.0%), α -guaiene (10.0%)]; 40: *N. leucophylla* [dihydroiridodial diacetate (18.2%), iridodial diacetate (7.8%)]; 43: *N. clarkei* [kaur-16ene (36.6%), pimara-7,15-dien-3-one (19.7%), methylabietate (5.3%), manoyl oxide (4.4%)]; 46: *N. satureioides* [(Z,E)-farnesol (14.7%), linalyl acetate (11.1%), Lavandulol acetate (6.6%)]; 48: *N. floccose* [geranial (32.4%)]; 49: *N. discolar* [linalyl acetate (12.3%), myrcene (10.7%)]; 52: *N. govaniana* i [torreyol (5.1%), α -muurolene (3.6%)] ii [cis-trans iridolactone (14.0%), gejerenene (6.8%), two nepetalactone (4.8%)]; 53: *N. pratti* [menthone (19.9%)]; 54: *N. bracteata* [bicyclogermacrene (11.4%)]; 55: *N. cilicica* [δ -cadinene (5.5%), α - copaene (4.5%)]; 56: *N. depauperata* [δ -cadinene (2.80%)]; 57: *N. macrosiphon* i [α -muurolene (6.0%),bicyclogermacrene (5.7%), aromadendrene (4.8%), verbenone (4.4%), γ -eudesmol (3.9%), citronellyl acetate (3.8%), α -cadinene (3.0%)], ii [bicyclogermacrene (10.1%)]; 58: *N. sessilifolia* [lavandulyl acetate (16.7%)]; 59: *N. sintenissii* [nepetalactone (4.93%)]; 60: *N. asterotrichus* [4a β ,7 α ,7 $a\beta$ nepetalactone (14.8%), γ -terpinene (10.6%)]; 61: *N. makuensis* [viridiflorol (17.5%), T-cadinol (10.7%)].

N. atlantica	N. cataria	N. granatensis	N. tuberose
4aα,7α,7aβ-nepetalactone (71.4%)	4aα,7α,7aβ-nepetalactone (77.4%)	4aα,7α,7aβ-nepetalactone (39.4%)	4aα,7α,7aβ-nepetalactone (76.8%)
dihydronepetalactone (45) (3.1%)	dihydronepetalactone (5.0%)	eucalyptol (1,8-cineole) (24.0%)	dihydronepetalactone (5.9%)
β-caryophyllene (8.2%)	terpinene (46) (4.2%)	α-pinene (6.3%)	menthol (47) (1.6%)
farnesol (48) (2.5%)	limonene (4.1%)	α-phellandrene (49) (5.8%)	α-pinene (1.3%)
α-curcumene (50) (1.3%)	Thymol (1.3%)	ρ-cymene (51) (3.8%)	eucalyptol (1,8-cineole)(1.2%)

Table 3: Major compounds present in the essential oils of four Nepeta species

Table 4: Major compounds present in six Himalayan Nepeta species:

Species Name	Major compounds
N. clarkei	α-guaiene (82) (10.0%); germacrene D (13.0%); β-sesquiphellandrene (22.0%) iridodial β-monoenol acetate diastereomers (25.3%)
N. discolor	<i>ρ</i> -cymene (9.8%); <i>β</i> -caryophyllene (18.6%); 1,8-cineole (25.5%)
N. elliptica	(7 <i>R</i>)- <i>trans,trans</i> -nepetalactone (83.4%)
N. erecta	isoiridomyrmecin (66.7%)
N. govaniana	pregeijerene (20.7%); isoiridomyrmecin (35.2%)
N. leucophylla	iridodial dienol diacetate (83) (7.8%); dihydroiridodial diacetate (84) (18.2%) iridodial β-monoenol acetate (25.4%)

REFRENCES

- Cardona, J; Celis, A; Cuca, L; Delgado, W et al. (2008), "Extractos vegetales utilizados como biocontroladores conenfasis enla familia Piperacea", Una Revision Agronomia Colombiana, Vol. 26, 97-106.
- Baser, KHC; Demirci, B; Kirimer, N and Kurkcuoglu, M (2000), "Essential oils of *Nepeta* species growing in Turkey", *Chem Nat Compd*, Vol. 36, 356–359.
- 3. Stuart, GA (1911), "*Chinese Materia Medica*", Shanghai, 281.
- 4. Avicenna (1956), "*Kitabul Adviyatul Qalbia*", National Printing Co, Aligarh.
- 5. Sherani, AW (1887), "*Tazkira Imam Sawedi*", Illahiya Press, Egypt.
- Daferera, D; Polissiou, M; Sokmen, A; Tepe, AS *et al.* (2007), "Antioxidant activity of the essential oil and various extracts of *Nepeta flavida* Hub. Mor from Turkey", *Food Chem*, Vol. 103 (4), 1358-1364.
- Sajjadi, SE (2005), "Analysis of the essential oil of *Nepeta sintenisii* bornm. from Iran", *Daru*, Vol. 13 (2), 61-64.
- Dabiri, M and Sefidkon, F (2003), "Chemical composition of the essential oil of *Nepeta racemosa* Lam. from Iran", *Flav Fragr J*, Vol. 18, 157-158.
- Amin, GR (1991), "Popular Medicinal Plants of Iran", Vol. 1, Iranian Ministry of Health Publications, Tehran, 40-44.
- Zargari, A (1990), "*Medicinal Plants*", Vol. 4, Tehran University Publications, Tehran, 4th ed. 106 -108.
- Agarwal, OP; Arora, RB and Khanna, DS (1978), "Studies of antiatherosclerotic action of *Nepeta hindostana* in pigs, *Arterry*, Vol. 4, 487-496.
- Miceli, N; Giuffrida, D; Taviano, MF; Trovato, A *et al.* (2005), "Antiinflammatory activity of extract and fractionsfrom *Nepeta sibthorpii* Bentham", *J Ethanopharma*, Vol. 97 (2), 261-266.
- 13. Prokopenko, SA and Spiridonov, AV (1985), "Betulin from Transcaucasian

clover (Nepeta)", *Farm Zhurnal*, Vol. 6, 70.

- 14. Flores, M; Galati, EM; Rapisarda, A; Tzakou, O et al. (2001), "Nepeta sibthorpii Bentham (Lamiaceae): Micromorphological analysis of leaves and flowers", *Farmaco*, Vol. 56, 413– 15.
- 15. Tucker, AO and Tucker, SS (2009), "Catnip and catnip response", *Econ Bot*, Vol. 42, 214.
- 16. Kafaru, EO (1994), "*Simple ways of staying healthy*", Elikaf Health Services Ltd, 32-33.
- 17. Edewor, TI and Usman, LA (2011), "Phytochemical and antibacterial activities of leaf extracts of *Nepeta cataria*", *Afr J Pure and Applied Chem*, Vol. 5 (16), 503-506.
- Bourrel, C; Bessiere, JM; Michel, G and Perineau, F (1993), "Catnip (*Nepeta cataria* L.) essential oil: analysis of chemical constituents, bacteriostatic and fungistatic properties", *J Essen Oil Res*, Vol. 5, 159.
- 19. Bankova, V; Galabov, A; Ignatova, A; Kujumgiev, A *et al.* (1995), "Chemical composition and biological activity of leaf exudates from some Lamiaceae plants", *Pharmazie*, Vol. 50, 62–65.
- 20. Abbas, G; Ahmed, S; Gilani, SA; Hussain, J et al. (2009), "Platelet aggregation, antiglycation, cytotoxic, phytotoxic and antimicrobial activities of extracts of *Nepeta juncea*", Afr J Biotechnol, Vol. 8 (6), 935-940.
- 21. Nadkarni, KM (1976), "*Indian Materia Medica*", Vol. 1, 845-846.
- 22. James, K; Louey, J; Petersen, N; Salotti, D *et al.* (2001), "*Oil of catnip by supercritical fluid extraction*" Chemistry Department, Sacred Heart University 5151 Park Avenue, Fairfield, CT 06432-1000.
- 23. Vohora, SB (1986), "Unani joshandah drugs for common cold, catarrh, cough, and associated fevers", J *Ethnopharmacol*, Vol. 16, 201.
- 24. Chopra, IC; Handa, KL and Sobti, SN (1957), "Aromatic plant resources of

Jammu and Kashmir", *J Sci and Indust Res*, Vol. 16 (A), 18.

- 25. Israili, AH (1980), "Therapeutic basis of Unani Mufferrahat", *Eastern Pharmacist*, Vol. 23 (272), 39.
- 26. Khare, CP (2004), "Indian herbal remedies: rational western therapy, ayurvedic and other traditional usage, botany", 259.
- Mehrabani, M; Asadippour, A and Amoli, SS (2004), "Chemical constituents of the essential oil of *Nepeta depauperata* benth. from Iran" *DARU*, Vol. 12 (3), 98-100.
- Hussain, J; Hussain, ST; Shah, MR; Ullah, F *et al.* (2008), "Z Naturforsch C", *J Bio sci*, Vol. 63 (b), 591.
- 29. Pojarkova, AI (1954), "Flora of the U.S.S.R. Moscow", Vol. 20, Izdatel'stvo Akademii Nauk SSSR, 191-293.
- 30. The Wealth of India (1966), "*Raw materials. Publication and information Directorate*", Vol. 8, CSIR, New Delhi, 12-13.
- Rechinger, KH (1982), "Nepeta, Flora Iranica, Akademische Druck-u," Verlagsanstal Graz, Vol. 150 (2), 108-216.
- 32. Mabberley, DJ (1997), "*The plant Book*", Cambridge University Press.
- 33. GRIN (November 9, 2001), "Nepeta discolor information from ARS/GRIN", Taxonomy for Plants. National Germplasm Resources Laboratory, Beltsville, Maryland: USDA, ARS, National Genetic Resources Program. Retrieved October 12, 2009.
- 34. Dar, GH; Hassan, T and Khuroo, AA (2011), "Taxonomic status of genus Nepeta L. (Lamiaceae) in Kashmir Himalaya, India", Iran J Bot, Vol. 17 (2), 181-188.
- 35. Inouye, H (1991), "Iridoids", *Methods Plant Biochem*, Vol. 7, 99–143.
- Chapman, JV; Clark, LJ; Hamilton, JGC; Hallahan, DL *et al.* (1997), "Analysis of monoterpenoids in glandular trichomes of the catmint *Nepeta racemosa*", *The Plant J*, Vol. 11, 1387–1393.

- 37. Dar, BA; Ganai, BA; Hassan, T; Qurishi, MA *et al.* (2012), "Essential oil composition of *Nepeta raphanorhiza* Benth growing in Kashmir valley", *Rec Nat Prod*, Vol. 6 (1), 67-70.
- 38. Modnicki, D; Klimek, B and Tokar, M (2007), "Flavonoids and phenolic acids of nepeta cataria var. citriodora (becker) balb. (lamiaceae)", Acta Poloniae Pharmaceut Drug Res, Vol. 64 (3), 247-252.
- Alwahsh, MAA; Haider, S; Jamila, N; Ullah, R; *et al.* (2011), "Secondary metabolites from *Nepeta juncea*", *Afr J Biotech*, Vol. 10 (77), 17884-17886.
- 40. Gkinis, G; Iliopoulou, D; Roussis, V and Tzakou, O (2003), "Chemical composition and biological activity of *Nepeta parnassica* oils and isolated Nepetalactones", *Z Naturforsch*, Vol. 58 (c), 681-686.
- 41. Eisenbraun, EJ; Regnier, FE and Waller, GR (1967), "Studies on the composition of the essential oils of three *Nepeta* Species", *Phytochem*, Vol. 6 (9), 1281–1289.
- Akhgar, MR; Jafari, S and Moradalizadeh, M (2012), "Chemical composition of the essential oil of *Nepeta assurgens* Hausskn. ex Bornm", *Trends in Modern Chemistry (TMC)*, Vol. 2 (1), 31-35.
- 43. 43, Altintas, A; Baser, KHC; Demircakmak, B and Duman, H (1998), "Composition of the essential oils of *Nepeta cadmea* Boiss', *J Essent Oil Res*, Vol. 10 (3), 327-328.
- 44. Baser, KHC and Ozek, T (1994), "Composition of the essential oil of *Nepeta caesarea* Boiss. from Turkey", *J Essent Oil Res*, Vol. 6, 645-646.
- 45. Aydin, A; Baser, KH; Beis, R and Ozturk, Y (1998), "Nepetalactone: a new opioid analgesic from *Nepeta caesarea* Boiss", *J Pharm Pharmacol*, Vol. 50 (7), 813-817.
- 46. Komeilizadah, H; Monfared, A; Nadji, K and Rustaiyan, A (2000), "Volatile constituents of *Nepeta denudata* Benth. and *N. cephalotes* Boiss. from Iran", *J Essent Oil Res*, Vol. 12 (4), 462-466.

- 47. Jamzad, Z and Sefidkon, F (2007), "Essential Oil Composition of Four Iranian Nepeta Species (N.cephalotes, N. bornmuelleri, N. mirzayanii and N. bracteata)", J Essent Oil Res, Vol. 19 (3), 262-265.
- 48. Dabiri, M and Sefidkon, F (2003),
 "Chemical composition of *Nepeta* crassifolia Boiss. & Buhse oil from Iran", *Flav Fragr J*, Vol. 18, 225–227.
- 49. Blagojevic, PD; Menezes, FS; Rabbitt, K and Radulovic, N (2011), "Essential oil of *Nepeta x faassenii* Bergmans ex Stearn (*N. mussinii* Spreng. x *N. nepetella* L.): a comparison study", *Nat Prod Commun*, Vol. 6 (7), 1015-1022.
- 50. Agarwal, SG; Kapahi, BK; Srivastava, TN and Thappa, RK (2001), "Essential oil of four Himalayan *Nepeta* Species," *J Essent Oil Res*, Vol. 13 (3), 189-191.
- 51. Ghazian, F; Kazempour, N; Mahboubi, M and Taghizadeh, M (2011), "Chemical composition, antioxidant and antimicrobial activity of *Nepeta persica* Boiss. essential oil", *Herba polonica*, Vol. 57 (1), 62-71.
- 52. Khosravi, M; Larijany, K and Rustaiyan, A (2000b), "Composition of the essential oil of *Nepeta racemosa* Lam. from Iran", *J Essent Oil Res*, Vol. 12, 151-152.
- 53. Perez-Alonso, MJ; Rodriquez, AB and Velasco-Negueruela, A (1989), "Essential oil analysis of *Nepeta teydea* Webb. & Berth", *Flav Fragr J*, Vol. 4 (4), 197–199.
- 54. Galati, EM; Harvala, C; Sanogo, R and Tzakou, O (2000) Essential oil composition of *Nepeta argolica* Bory et Chaub. subsp. *argolica*", *Flav Fragr J*, Vol. 15, 115-118.
- 55. Amzal, H; Bakri, Y; Boudhane, A; Zenasni, L *et al.* (2008), "The essentials oils and antimicrobial activity of four *Nepeta* species from Morocco", *J Med Plants Res*, Vol. 2, 111–114.
- 56. Adiguzel, A; Gulluce, M; Ozer, H; Sokmen, M; *et al.* (2009), "Antimicrobial and antioxidant activity of the essential oil and methanol extract

of *Nepeta cataria*", *Polish J Microb*, 58 (1), 69-76.

- 57. Morteza-Semnani, K and Saeedi, M (2004), "Essential oils composition of *Nepeta cataria* L. and *Nepeta crassifolia* Boiss. and Buhse from Iran", *J Essent Oil Bearing Plants*, Vol. 7 (2), 120-124.
- 58. Benito, PB; Perez-Alonso, MJ; Rico, MM and Velasco-Negueruela, A (1998), "Composicion de los aceites esenciales de Nepeta nepetella subsp. aragonensis, Nepeta coerulea subsp. coerulea y Nepeta cataria", Giorn Bot Ital, Vol. 122, 295-302.
- 59. Alpsoy, L; Atici, O; Kekec, G; Mutlu, S et al. (2012), "Genotoxic effects of catmint (*Nepeta meyeri* Benth.) essential oils on some weed and crop plants", *Toxicology and Industrial Health*.
- 60. Bozin, B; Gkinis, G; Mimica-Dukic, N and Tzakou, O (2010), "Antioxidant activity of *Nepeta nuda L. ssp. nuda* essential oil rich in nepetalactones from Greece", *J Med Food*, Vol. 13 (5), 1176-1181.
- Kokdil, G; Kurucu, S and Topcu, G (1996), "Composition of the essential oil of *Nepeta nuda* L. ssp. *Albiflora* (Boiss.) Gams", *Flav Fragr J*, Vol. 11, 167-169.
- 62. Akgul, A; Baser, KHC; Ozek, T and Tumen, G (1993), "Composition of the Essential Oil of *Nepeta racemosa* Lam", *J Essent Oil Res*, Vol. 5, 215-217.
- Chalchat, JC; Gorunovic, MS; Maksimovic, ZA and Petrovic, SD (2000), "Composition of the essential oils of *Nepeta rtanjensis* Diklic et Milojevic, Lamiaceae from Serbia", J *Essent Oil Res*, 12 (2), 238-240.
- 64. Grbic, ML; Sokovic, M; Stupar, M; Vukojevic, J *et al.* (2008), "Antifungal activity of *Nepeta rtanjensis* essential oil", *J Serb Chem Soc*, Vol. 73 (10), 961–965.
- 65. El-Moaty, HIA (2010), "Essential oil and iridoide glycosides of *Nepeta septemcrenata* Erenb", *J Nat Pro*, Vol. 3, 103-11.

- 66. Baser, KH; Demirci, B; Iscan, G and Kose, YB (2011), "Anticandidal activity of the essential oil of *Nepeta transcaucasica* Grossh", *Chem Biodivers*, Vol. 8 (11), 2144-2148.
- 67. Jamzad, Z; Mirza, M and Sefidkon, F (2006), "Chemical composition of the essential oil of five Iranian Nepeta species (N. crispa, N. mahanensis, N. ispahanica, N. eremophila and N. rivularis)", Flav Fragr J, Vol. 21, 764–767.
- 68. Oji, K and Shafaghat, A (2010), "Nepetalactone content and antibacterial activity of the essential oils from different parts of *Nepeta persica*", *Nat Prod Commun*, Vol. 5(4), 625-628.
- Khaligh, P; Mirzajani, F; Salehi, P and Sonboli, A (2012), "Essential oil composition and antioxidant activity of different extracts of *Nepeta betonicifolia* C.A. Meyer and *Nepeta saccharata* Bunge", *Nat Prod Res*, Vol. 26 (8), 736-743.
- 70. Costantinidis, T; Lazari, DM; Loukis, AE and Skaltsa, HD (2000), "Essential oil analysis of *Nepeta argolica* Bory et Chaub. subsp. *argolica* (Lamiaceae) growing wild in Greece", *Flav Fragr J*, Vol. 15, 96-99.
- 71. Handjieva, NV and Popon, SS (1996), "Constituents of essential oils from *Nepeta cataria* L., *N. grandiflora* M. B., and *N. nuda* L", *J Essent Oil Res*, Vol. 8, 639-643.
- 72. Baharvand, B; Deyhimi, F; Nori-Shargh, D and Raftari, S (2006), "The Volatile Constituents Analysis of Nepeta kotschyi Boiss. from Iran", J Essent Oil Res, Vol. 18 (3), 237-238.
- 73. Hosseini, M and Moghaddam, FM (1996), "Composition of the essential oil from *Nepeta crassifolia* Boiss. & Buhse", *Flav Fragr J*, Vol. 11, 113-115.
- 74. Agar, G; Aksakal, O; Bozari, S and Erturk, FA (2012), "Determination of chemical composition and genotoxic effects of essential oil obtained from *Nepeta nuda* on *Zea mays* seedlings",

Toxicol Ind Health, doi: 10.1177/0748233711433939.

- 75. Jack (2012), "Study on Chemical Composition of the *Nepeta Angustifoliathe* GY. WU. and the Extraction Technique", *Med Res*, 127.
- 76. Bagci, E; Behcet, L and Kilic, O (2013), "Essential Oil Compounds of Three Nepeta L. Taxa from Turkey and their Chemotaxonomy", Asian J Chem, Vol. 25 (14).
- 77. Bicchi, C; Mashaly, M and Sandra, P (1984), "Constituents of essential oil of *Nepeta nepetella*", *Planta Med*, Vol. 50 (1), 96-98.
- Barroso, JC; Cotrim, HC; Figueiredo, AC; Pais, MSS et al. (1994), "Composition of the essential oil from inflorescences of *Nepeta tuberosa* L. ssp. tuberose", *Flav Fragr J*, Vol. 9, 71-73.
- 79. Bernardi, MM; Lago, JHG; Ricci, EL; Romoff, P *et al.* (2010), "Antinociceptive and anti-inflammatory actions of *Nepeta cataria* L. var. citriodora (Becker) Balb. essential oil in mice", *J Health Sci Inst*, Vol. 28 (3), 289-93.
- Bisht, DS; Lal, P; Mathela, CS; Padalia, RC *et al.* (2010), "Constituents and antimicrobial activity of the essential oils of six Himalayan *Nepeta* species", *J Serb Chem Soc*, Vol. 75 (6), 739–747.
- 81. El-Hamouly; El-Hela, AA and Mohamed MA (2004), "Phytochemical and biological investigation of the volatile constituents of Nepeta septemcrenata ehrenb., growing in Sci. Egypt", **Bull Pharm** Assiut University, Vol. 27 (1), 95-98.
- Amold, NA; Feo, VD; Formisano, C; Mancini, E et al. (2009), "Phytotoxic effects of essential oils of Nepeta curviflora Boiss. and Nepeta nuda L. subsp. albiflora growing wild in Lebanon", J Plant Interactions, Vol. 4 (4), 253-259.
- 83. Arnold, NA; Piozzi, F and Senatore, F (2005), "Composition of the Essential Oil of *Nepeta curviflora* Boiss.

(Lamiaceae) from Lebanon", *J Essent Oil Res*, Vol. 17 (3), 268.

- 84. Mehregan, I and Sajjadi, SE (2005), "Chemical Constituents of the Essential Oil of *Nepeta daenensis* Boiss", *J Essent Oil Res*, Vol. 17 (5), 563.
- Alamshahi, A; Dabiri, M and Sefidkon, F (2002), "Analysis of the essential oil of *Nepeta fissa* C.A. Mey from Iran", *Flav Fragr J*, Vol. 17, 89-90.
- 86. Alim, A; Atas, AD; Cetin, A; Cetinus, SA et al. (2009), "Chemical composition and *in vitro* antimicrobial and antioxidant activities of the essential oil of *Nepeta nuda* L. subsp. *Albiflora* (Boiss.) gams", *Afr J Microbiol Res*, Vol. 3 (8), 463-467.
- 87. Eskandari, B and Sajjadi, SE (2005), "Chemical constituents of the essential oil of *Nepeta oxyodonta*", *Chem Nat Compds*, Vol. 41 (2), 175-177.
- 88. Kökdil, G; Kurucu, S; Tanker, M and Topcu, G (1997), "Essential Oil Analysis of *Nepeta cilicia* Boiss", *Flav Fragr J*, Vol. 12, 99–101.
- 89. Baser, KHC; Bemirci, B; Ozek, T and Tumen, G (2001), "Composition of the essential oil of *Nepeta betonicifolia* C. A. Meyer from Turkey", *J Essent Oil Res*, Vol. 13, 35-36.
- 90. Kokdil, G; Kurucu, S and Yıldız, A (1998), "Essential oil composition of *Nepeta nuda* L. ssp. *nuda*", *Flav Fragr J*, Vol. 13, 233–234.
- 91. Nadji, N and Rustaiyan, A (1999), "Composition of the essential oils of Nepeta ispahanica Boiss. and Nepeta binaludensis Jamzad from Iran", Flav Fragr J, Vol. 14, 35-37.
- 92. Honermeier, B; Koocheki, A; Moghadam, PR and Nadjafi, F (2012), "First experiments on cultivation of *Nepeta binaludensis* Jamzad - an example of domestication of a highly endangered medicinal plant of Iran", Z *Arznei- Gewurzpfla*, Vol. 17 (2), 64 – 71.
- 93. Baser, KHC; Demirci, B and Kaya, A (2007), "Micromorphology of glandular trichomes of *Nepeta congesta* Fisch. & Mey. var. congesta (Lamiaceae) and chemical analysis of

the essential oils", *South African J Bot*, Vol. 73 (1), 29-34.

- 94. Nickavar, B; Mojab, F and Tehrani, HH (2009), "Essential oil analysis of *Nepeta crispa* and *N. menthoides* from Iran", *Iranian J Pharma Sci Winter*, Vol. 5 (1), 43-46.
- 95. Salehi, P; Sonboli, A and Yousefzadi, M (2004), "Antimicrobial activity and chemical composition of the essential oil of *Nepeta crispa* Willd. from Iran", *Z Naturforsch*, Vol. 59 (c), 653–656.
- 96. Bamoniri, A; Batooli, H; Haghani, M and Safaei-Ghomi, J (2006), "Essential Oil Composition of Nepeta gloeocephala Rech. f. from Iran", J Essent Oil Res, Vol. 18 (6), 635.
- 97. Jamzad, M; Jamzad, Z; Masoudi, S and Rustaiyan, A (2008), "Composition of the Essential Oils of *Nepeta sessilifolia* Bunge and *Nepeta haussknechtii* Bornm. from Iran", *J Essent Oil Res*, Vol. 20 (6), 533-535.
- 98. Khatamsaz, M and Sajjadi, SE (2001), "Volatile Constituents of Nepeta heliotropifolia Lam", J Essent Oil Res, Vol. 13 (3), 204-205.
- 99. Allahyari, L; Salehi, P and Sonboli, A (2005), "Essential Oil Composition of *Nepeta involucrata* from Iran", *Chem Nat Compds*, Vol. 41 (6), 683-685.
- 100. Kokdil, G; Kurucu, S and Topcu, G (1997), "Chemical constituents of the essential oils of *Nepeta italica* L. and *Nepeta sulfuriflora* P.H. Davis", *Flav Fragr J*, Vol. 12 (1), 33-35.
- 101. Gholipour, A; Mojarrad, M; Sonboli, A and Yousefzadi, M (2009), "Antibacterial activity and composition of the essential oil of *Nepeta menthoides* from Iran", *Nat Prod Commun*, Vol. 4 (2), 283-286.
- 102. Dayan, FE; Kobaisy, M; Mamonov, LK; Mukanova, GS *et al.* (2005), "Composition and Phytotoxic Activity of *Nepeta pannonica* L. Essential Oil", *J. Essent Oil Res*, Vol. 17 (6), 704.
- 103. Ali, T; Javan, M; Semnanian, S and Sonboli, A (2012), "Evaluation of the antinociceptive and anti-inflammatory effects of essential oil of *Nepeta*

pogonosperma Jamzad et Assadi in rats", **Daru**, Vol. 20 (1), 48.

- 104. Akhgar, M R and Moradalizadeh, M (2012), "Chemical composition of the essential oils from stems, flowers and leaves of *Nepeta schiraziana* Boiss", *Iranian J Med Aromatic Plants*, Vol. 28 (1 (55)), 28-34.
- 105. Hadjiakhoondi, A; Gohari, AR and Saeidnia, S (2008), "Trypanocidal Activity of Oil of the Young Leaves of *Nepeta cataria* L. Obtained by Solvent Extraction", *J Med Plants*, Vol. 7 (4), 54-57.
- 106. 106. Bhat, KA; Hassan, T; Rather, MA; Shawl, AS *et al.* (2011), "Chemical composition of the essential oils of *Nepeta laevigata* and *Nepeta elliptica* from India", *Chem Nat Comp*, Vol. 47 (3), 456-458.
- 107. Javidnia, K; Mehregan, I; Miri, R and Sadeghpour, H (2005), "Volatile constituents of the essential oil of *Nepeta ucrainica* L. ssp. *kopetdaghensis* from Iran", *Flavour Fragr J*, Vol. 20, 219–221
- 108. Farjam, MH (2012), "Antibacterial activity and composition of essential oil of *Nepeta pungens* Benth. from Iran", *J Appl Pharmaceut Sci*, Vol. 2 (4), 103-105.
- 109. 109 Mothana, RA (2012), "Chemical composition, antimicrobial and antioxidant activities of the essential oil of *Nepeta deflersiana* growing in Yemen", *Rec Nat Prod*, Vol. 6 (2), 189-193.
- 110. Bisht, DS; Joshi, SC; Mathela, CS and Padalia, RC (2012), "Isoiridomyrmecin rich essential oil from *Nepeta erecta* Benth. and its antioxidant activity", *J Essent Oil Res*, Vol. 26 (1), 29-35.
- 111. Hassan, T and Rather, MA (2011), "Analysis of the diterpene rich essential oil of *Nepeta clarkei* Hooke. from Kashmir Himalayas by capillary GC-MS", *Int J ChemTech Res*, Vol. 3 (2), 959.
- 112. Comlekciogolu, N and Karaman, S (2007), "Essential oil composition of *Nepeta Cilicia* Boiss. A pud bentham

and *Phlomis viscose* Poiret from Turkey", *Int J Bot*, Vol. 3 (1), 122-124.

- 113. Ozcan, M and Senatore, F (2003), "Composition of the essential oil of *Nepeta betonicifolia* C.A. Meyer (Lamiaceae) from Turkey", *J Essent Oil Res*, Vol. 15 (3), 200-201.
- 114. Ebrahimi, SN; Hadian, J; Mirjalili, MH and Sonboli, A (2006), "Essential oil composition of Nepeta satureioides from Iran", Chem Nat Compds, Vol. 42 (2), 175-177.
- 115. Ghassemi, N and Sajjadi, SE (1999),
 "Volatile constituents of Nepeta glumerulosa Boiss. subsp. carmanica", Flav Fragr J, Vol. 14, 265-267.
- 116. Batooli, H; Nahavandi, S and Safaei-Ghomi, J (2011), "Studies on the antioxidant activity of the volatile oil and methanol extracts of *Nepeta laxiflora* benth. and *Nepeta sessilifolia* bunge. from Iran", *J Food Biochem*, Vol. 35, 1486–1492.
- 117. Hassan, T; Dar, BA; Rather, MA; Sofi, SN *et al.* (2011), "GC-FID and GC-MS analysis of the sesquiterpene rich essential oil of *Nepeta govaniana* (Wall. ex Benth.) Benth. from Jammu and Kashmir", *Int J Chem Tech Res*, Vol. 3 (3), 1194-1199.
- 118. Kharkwal, H; Laurent, R and Mathela, CS (1994), "Investigations on Himalayan Nepeta Species. V. Essential Oil of Nepeta govaniana Benth", J Essent Oil Res, Vol. 6 (4), 425-428.
- 119. Xi-feng, G (2010-09), "Research on the Extraction of Essential Oil from Nepeta pratti Levl Based on CO₂ Supercritical Extraction Technology", J Anhui Agri Sci.
- 120. Arnold, NA; Formisano, C; Piozzi, F; Rigano, D et al. (2013), "GC and GC-MS analysis of the essential oil of *Nepeta cilicica* Boiss. ex Benth. from Lebanon", Nat Prod Res.
- 121. Aghazaria, F; Ghannadi, A; Mehrabania, M; Mohagheghzadeh, A *et al.* (2003), "Quantity and composition of the SDE prepared

essential oil of *Nepeta macrosiphon* boiss", *Iranian J Pharma Res*, Vol. 2, 103-105.

- 122. Jafari, A; Javidnia, K; Miri, R and Rezai, H (2004), "Analysis of the volatile constituents of *Nepeta macrosiphon* Boiss. grown in Iran", *Flavour Fragr J*, Vol. 19, 156–158.
- 123. 123, Abad, MHK; Nasrabadi, M and Vahedi, H (2011), "Chemical composition of the essential oil of *Nepeta sintenisii* leaves growing wild in Darkesh protected area (North Khorassan Province Iran)", *Der Pharmacia Sinica*, Vol. 2 (3), 207-210.
- 124. Masoudi, S; Monfared, A and Rustaiyan, A (1999), "Composition of the Essential Oil of *Nepeta asterotrichus* Rech. F. et Aell. from Iran", *J Essent Oil Res*, Vol. 11 (2), 229-230.

- 125. Habibi, Z; Masoudi, S and Rustaiyan, A (2004), "Essential Oil of Nepeta makuensis Jamzad et Mozaffarian from Iran", J Essent Oil Res, Vol. 16 (3), 214-215.
- 126. Angioni, A; Barra, A; Coroneo, V; Dessi, S *et al.* (2006), "Chemical composition, seasonal variability, and antifungal activity of *Lavandula stoechas* L. ssp. stoechas essential oils from stem/leaves and flowers", J Agric Food Chem, Vol. 54, 4364-4370.
- 127. Bessiere, JM; Juteau, F; Masotti, V and Viano, J (2003), "Seasonal and phenological variations of the essential oil from the narrow endemic species *Artemisia molinieri* and its biological activities", *J Agric Food Chem*, Vol. 51, 7115-7121.

Correspondence Author: Damanjit Singh Cannoo Department of Chemistry, Sant Longowal Institute of Engineering and Technology, Longowal, Sangrur, Punjab-148106, India Email: 13daman@gmail.com

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